Tuberculosis

Practical guide for clinicians, nurses, laboratory technicians and medical auxiliaries

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Introduction

Tuberculosis is an infectious disease caused by *Mycobacterium tuberculosis*. Tuberculosis typically attacks the lungs, but can also affect other parts of the body. The disease has become rare in high income countries, but is still a major public health problem in low- and middle-income countries.

It is estimated that between the years 2000 and 2010, eight to nine million new cases emerged each year. Approximately 1.5 million people die from the disease each year. In adults, tuberculosis is the second leading cause of death due to an infectious disease (after AIDS), with 95% of deaths occurring in low-income countries. Tuberculosis is a major problem of children in poor countries where it kills over 100,000 children each year.

The treatment of tuberculosis remains a constraint for patients and a heavy burden for the healthcare system. Drug-susceptible tuberculosis requires at least six months of therapy under close supervision. A treatment for multidrug-resistant tuberculosis requires nearly two years of treatment with poorly tolerated and less effective drugs. In most places the diagnosis still relies mainly on direct microscopy that is unable to detect a large proportion of patients. The BCG vaccine, developed almost a century ago, confers only partial protection.

After 40 years of minimal progress in the tools to fight tuberculosis there are some reasons for hope. A few new drugs are reaching the final phase of development; a new molecular test that can be decentralized to some extent and allows the rapid diagnosis of tuberculosis and of resistance to rifampicin has been introduced. Though this is undeniable progress, much will be needed to bring the new tools and drugs to the patients in need. Furthermore, a true “point of care” diagnostic test still does not exist and little progress has been made in research for a more effective vaccine.

Case management of patients does not necessarily have to involve a major, vertical programme. It should be incorporated into the framework of other medical activities in order to offer comprehensive and integrated treatment even if the number of patients being treated is relatively small.

This guide has been developed jointly by Médecins Sans Frontières and Partners In Health. It aims at providing useful information to the clinicians and health staff for the comprehensive management of tuberculosis. Forms of susceptible and resistant tuberculosis, tuberculosis in children, and HIV co-infection are all fully addressed.

As treatment protocols are constantly changing, medical staff are encouraged to check this website for updates.
Abbreviations and acronyms

Update: June 2023
<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
</tr>
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<tbody>
<tr>
<td>ACH</td>
<td>Air change per hour</td>
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<tr>
<td>AFB</td>
<td>Acid-fast bacilli</td>
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<tr>
<td>ALT</td>
<td>Alanine aminotransferase</td>
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<tr>
<td>Am</td>
<td>Amikacin</td>
</tr>
<tr>
<td>Amx/Clv</td>
<td>Amoxicillin/clavulanic acid</td>
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<td>ART</td>
<td>Antiretroviral therapy</td>
</tr>
<tr>
<td>ARV</td>
<td>Antiretroviral</td>
</tr>
<tr>
<td>AST</td>
<td>Aspartate aminotransferase</td>
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<tr>
<td>BCG</td>
<td>Bacillus Calmette-Guérin</td>
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<td>Bdq</td>
<td>Bedaquiline</td>
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<tr>
<td>BPNS</td>
<td>Brief peripheral neuropathy screen</td>
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<tr>
<td>BSC</td>
<td>Biosafety cabinet</td>
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<tr>
<td>CDC</td>
<td>Centers for Disease Control and Prevention</td>
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<tr>
<td>Cfz</td>
<td>Clofazimine</td>
</tr>
<tr>
<td>CrCl</td>
<td>Creatinine clearance</td>
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<tr>
<td>CMX</td>
<td>Cotrimoxazole</td>
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<tr>
<td>CNS</td>
<td>Central nervous system</td>
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<tr>
<td>CPC</td>
<td>Cetylpyridinium chloride</td>
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<tr>
<td>CPT</td>
<td>Cotrimoxazole preventive therapy</td>
</tr>
<tr>
<td>Cs</td>
<td>Cycloserine</td>
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<td>CSF</td>
<td>Cerebrospinal fluid</td>
</tr>
<tr>
<td>CXR</td>
<td>Chest x-ray</td>
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<tr>
<td>Dlm</td>
<td>Delamanid</td>
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<tr>
<td>DOT</td>
<td>Directly observed therapy</td>
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<tr>
<td>DR</td>
<td>Drug resistance</td>
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<tr>
<td>DR-TB</td>
<td>Drug-resistant tuberculosis</td>
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<tr>
<td>DS-TB</td>
<td>Drug-susceptible tuberculosis</td>
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<tr>
<td>DST</td>
<td>Drug susceptibility test</td>
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<td>--------------------------</td>
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<tr>
<td>Eth</td>
<td>Ethambutol</td>
</tr>
<tr>
<td>ECG</td>
<td>Electrocardiogram</td>
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<tr>
<td>EPTB</td>
<td>Extrapulmonary tuberculosis</td>
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<tr>
<td>Eto</td>
<td>Ethionamide</td>
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<tr>
<td>FDC</td>
<td>Fixed-dose combination</td>
</tr>
<tr>
<td>FNA</td>
<td>Fine needle aspiration</td>
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<tr>
<td>FQ</td>
<td>Fluoroquinolone</td>
</tr>
<tr>
<td>gDST</td>
<td>Genotypic drug susceptibility test</td>
</tr>
<tr>
<td>GUV</td>
<td>Germicidal ultraviolets</td>
</tr>
<tr>
<td>H</td>
<td>Isoniazid (standard dose)</td>
</tr>
<tr>
<td>HbA1c</td>
<td>Glycated haemoglobin</td>
</tr>
<tr>
<td>H^h</td>
<td>Isoniazid (high dose)</td>
</tr>
<tr>
<td>HIV</td>
<td>Human immunodeficiency virus</td>
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<tr>
<td>HPF</td>
<td>High-power field</td>
</tr>
<tr>
<td>Hr</td>
<td>Isoniazid resistance</td>
</tr>
<tr>
<td>Hr-TB</td>
<td>Isoniazid resistant tuberculosis</td>
</tr>
<tr>
<td>IGRA</td>
<td>Interferon gamma release assay</td>
</tr>
<tr>
<td>IPC</td>
<td>Infection prevention and control</td>
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<tr>
<td>IM</td>
<td>Intramuscular</td>
</tr>
<tr>
<td>Imp/Cln</td>
<td>Imipenem/cilastatin</td>
</tr>
<tr>
<td>INI</td>
<td>Integrase inhibitor</td>
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<tr>
<td>IRIS</td>
<td>Immune reconstitution inflammatory syndrome</td>
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<tr>
<td>IV</td>
<td>Intravenous</td>
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<tr>
<td>LF-LAM</td>
<td>Lateral flow urine lipoarabinomannan assay</td>
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<tr>
<td>LFT</td>
<td>Liver function test</td>
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<tr>
<td>Lfx</td>
<td>Levofloxacin</td>
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<tr>
<td>Abbreviation</td>
<td>Description</td>
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</tr>
<tr>
<td>LPA</td>
<td>Line probe assay</td>
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<td>LTBI</td>
<td>Latent tuberculosis infection</td>
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<tr>
<td>LTR</td>
<td>Long treatment regimen</td>
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<tr>
<td>Lzd</td>
<td>Linezolid</td>
</tr>
<tr>
<td>MDR</td>
<td>Multidrug resistance</td>
</tr>
<tr>
<td>MDR-TB</td>
<td>Multidrug-resistant tuberculosis</td>
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<tr>
<td>Mfx</td>
<td>Moxifloxacin (standard dose)</td>
</tr>
<tr>
<td>Mfx&lt;sub&gt;h&lt;/sub&gt;</td>
<td>Moxifloxacin (high dose)</td>
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<tr>
<td>Mpm</td>
<td>Meropenem</td>
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<tr>
<td>MTB</td>
<td>Mycobacterium tuberculosis</td>
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<tr>
<td>NAAT</td>
<td>Nucleic acid amplification test</td>
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<tr>
<td>NGS</td>
<td>Next generation sequencing</td>
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<tr>
<td>NNRTI</td>
<td>Non-nucleoside reverse transcriptase inhibitor</td>
</tr>
<tr>
<td>NRTI</td>
<td>Nucleoside reverse transcriptase inhibitor</td>
</tr>
<tr>
<td>P</td>
<td>Rifapentine</td>
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<tr>
<td>Pa</td>
<td>Pretomanid</td>
</tr>
<tr>
<td>PAS</td>
<td>Para-aminosalicylic acid</td>
</tr>
<tr>
<td>PCP</td>
<td>Pneumocystosis</td>
</tr>
<tr>
<td>PCR</td>
<td>Polymerase chain reaction</td>
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<tr>
<td>pDST</td>
<td>Phenotypic drug susceptibility test</td>
</tr>
<tr>
<td>PI</td>
<td>Protease inhibitor</td>
</tr>
<tr>
<td>PO</td>
<td>Orally (per os)</td>
</tr>
<tr>
<td>PTB</td>
<td>Pulmonary tuberculosis</td>
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<tr>
<td>Pto</td>
<td>Prothionamide</td>
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<tr>
<td>R</td>
<td>Rifampicin</td>
</tr>
<tr>
<td>Rfb</td>
<td>Rifabutin</td>
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<tr>
<td>RMT</td>
<td>Rapid molecular test</td>
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<tr>
<td>RR</td>
<td>Rifampicin resistance</td>
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<tr>
<td>S</td>
<td>Streptomycin</td>
</tr>
<tr>
<td>SAT</td>
<td>Self-administered treatment</td>
</tr>
<tr>
<td>STR</td>
<td>Short treatment regimen</td>
</tr>
<tr>
<td>TB</td>
<td>Tuberculosis</td>
</tr>
<tr>
<td>tNGS</td>
<td>Targeted next generation sequencing</td>
</tr>
<tr>
<td>Trd</td>
<td>Terizidone</td>
</tr>
<tr>
<td>TSH</td>
<td>Thyroid-stimulating hormone</td>
</tr>
<tr>
<td>TST</td>
<td>Tuberculin skin test</td>
</tr>
<tr>
<td>ULN</td>
<td>Upper limit of normal</td>
</tr>
<tr>
<td>VVS</td>
<td>Ventilated workstation</td>
</tr>
<tr>
<td>WGS</td>
<td>Whole genome sequencing</td>
</tr>
<tr>
<td>WHO</td>
<td>World Health Organization</td>
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<tr>
<td>XDR</td>
<td>Extensive drug resistance</td>
</tr>
<tr>
<td>XDR-TB</td>
<td>Extensively drug-resistant tuberculosis</td>
</tr>
<tr>
<td>Z</td>
<td>Pyrazinamide</td>
</tr>
</tbody>
</table>
Chapter 1: Introduction and epidemiology

1.1 Characteristics of Mycobacterium tuberculosis bacillus
1.2 Transmission
1.3 Evolution of tuberculosis infection and disease in humans
1.4 Prognosis
1.5 Factors modifying tuberculosis epidemiology
1.6 Epidemiological indicators
1.7 Global burden of tuberculosis

Update: January 2022
1.1 Characteristics of Mycobacterium tuberculosis bacillus

*Mycobacterium tuberculosis*, along with *M. bovis*, *M. africanum*, *M. microti* and others, make up the *Mycobacterium tuberculosis* complex, a group of bacteria that cause clinical tuberculosis (TB) in humans. Most TB cases are caused by *M. tuberculosis*. Cases due to other species are far less prevalent.

*M. tuberculosis* is a small, rod-shaped, strictly aerobic, acid-fast bacillus\(^a\). Like other mycobacteria, it is slow growing, resulting in more gradual development of disease when compared with other bacterial infections.

**Footnotes**

(a) Acid-fast bacilli are bacilli, which once stained, resist discoloration by acid and alcohol.
1.2 Transmission

*M. tuberculosis* is transmitted from human-to-human and spread is mainly airborne. The source of infection is usually a person with pulmonary or laryngeal TB. During coughing, speaking, or sneezing, the person produces tiny infectious droplets. These particles, called droplet nuclei, are about 1 to 5 microns in diameter. Depending on the environment, they can remain suspended in the air for several hours.

Transmission may occur when these infectious droplets are inhaled. UV light (sunshine or artificial sources) and ventilation reduce the probability of transmission (Chapter 14).

Other modes of transmission are far less common. Inoculation of cutaneous or mucous membranes rarely occurs, although such cases have been observed in laboratory personnel. Congenital infection (by transplacental transmission or via aspiration or swallowing of infected amniotic fluid at birth) has been reported, but is very rare. Transmission through breast milk does not occur.

The infectiousness of a person is associated with the quantity of bacilli contained in their sputum. Patients with smear-positive sputum on microscopy are by far the most infectious. Those with smear-negative/culture-positive results are less infectious, but still contribute to TB transmission due to more frequent delays in diagnosis.

Persons infected with *M. tuberculosis*, but who have not developed active TB (latent tuberculosis infection), are not infectious. Persons with extrapulmonary TB (EPTB) are only infectious in exceptional circumstances.

Children are generally much less infectious than adults. This may be due to weaker cough mechanics, less sputum production and lower bacillary load.

Not everyone who is exposed to an infectious TB patient becomes infected with *M. tuberculosis*. The probability that TB will be transmitted depends on several factors:

**Infectiousness of the source** (the most important factor)
- Bacteriological status: smear-positive patients are the most infectious.
- Virulence of the bacilli: some strains are highly transmissible (and/or more likely to cause active TB).

**Environment where the exposure occurred**
- Outdoor environments or those with good ventilation and sunlight are less likely to lead to transmission. Small rooms or rooms with no ventilation are conditions most likely to lead to transmission.
- The proximity of the contact person to the patient is also important (e.g. the risk is higher if the person sleeps next to the patient than if they sleep 20 metres away from the patient).

**Duration of exposure**
People in close and prolonged contact with a person with TB are at highest risk of becoming infected with *M. tuberculosis*. They may be family members, roommates, friends, co-workers or other people who spend several hours a day with the infectious patient.

The best way to stop transmission is to start effective TB treatment as soon as possible. It is estimated that a person with untreated smear-positive TB transmits the bacillus to 10 to 20 people a year (with variations according to living conditions and environment).
1.3 Evolution of tuberculosis infection and disease in humans

When a person inhales infectious droplets containing *M. tuberculosis*, most of the larger droplets become lodged in the upper respiratory tract (nose and throat) where infection is unlikely to develop. However, smaller droplet nuclei may reach the small air sacs of the lung (the alveoli) where infection can occur.

1.3.1 Primary infection and latent tuberculosis infection

After transmission, *M. tuberculosis* multiplies slowly, in most cases in the terminal alveoli of the lungs (primary focus) and in the lymph nodes of corresponding drainage areas: this is the primary infection. The primary focus and related hilar lymphadenopathy form the primary complex.

In one to two months, due to the action of lymphocytes and macrophages (cellular immunity), the primary focus is contained and encapsulated, with a central zone of parenchymal necrosis (caseous lesions). It is not usually detectable on chest x-ray, unless it calcifies or grows substantially. Primary infection is usually asymptomatic. In most cases (90 to 95% of non-HIV infected patients), the pulmonary lesions gradually heal.

During the primary infection, specific immunity develops and a positive skin reaction to tuberculin is observed[1]. This immune response may persist without clinical signs of TB. The patient is infected by *M. tuberculosis*, but does not develop the disease. This is referred to as latent tuberculosis infection (LTBI).

In 5 to 10% of infected people, primary infection and/or LTBI progresses to active TB over their lifetime. For HIV co-infected patients, this risk is much higher.

1.3.2 Active tuberculosis

Before immunity is established, bacilli from the primary infectious focus or from a near-by lymph node can be transported and disseminated throughout the body via the lymph system or the bloodstream.

Secondary foci can develop this way, particularly in the lungs, lymph nodes, serous membranes, meninges, bones and kidneys. As soon as an immune response is mounted, most of these foci resolve spontaneously. However, some bacilli may remain dormant in the secondary foci for months and sometimes years.

Different factors can reduce the immune response (e.g. HIV infection) and lead to reactivation of the bacilli and their multiplication in one or more of these foci. This reactivation or progression of the primary or secondary foci results in active TB[2].

An active TB lesion contains actively, slowly or sporadically multiplying bacilli as well as dormant bacilli.

While active TB may occur months or years following primary infection, half of TB cases appear in the year following infection.

1.3.3 Risk factors for developing active tuberculosis

Certain factors increase the risk of developing active TB within the first two years of being infected. These factors include any factor that results in a weakened immune system, damaged lungs and the intensity and duration of exposure.

**Host immune response factors:**
- HIV infection
- Children under 5 years[3][4]
- Malnutrition
- Persons over 60 years
- Diabetes mellitus
- Other risk factors: prolonged corticosteroid therapy (e.g. prednisolone) and other immunosuppressive therapies, severe kidney disease, alcoholism, substance abuse, certain types of cancer (e.g. leukaemia, Hodgkin’s lymphoma, cancer of the head and neck); pregnancy

**Conditions that damage the lung:**
- Tobacco smoking
- Silicosis
- Chronic obstructive pulmonary disease (COPD)

**Intensity of exposure** (high number of inhaled bacilli):
- Highly infectious source
- Poorly ventilated environment
- Proximity with infectious source, including residents and employees of institutions such as prisons, boarding schools and residential care facilities
- Long duration of exposure

**References**


1.4 Prognosis

Without treatment, TB is a severe and potentially fatal disease. After 5 years without treatment, the outcome of smear-positive pulmonary TB (PTB) in non-HIV-infected patients is as follows:\cite{1}:
- 50 to 60% die (case fatality ratio (CFR) for untreated TB);
- 20 to 25% are cured (spontaneous cure);
- 20 to 25% continue to have symptoms.

Untreated TB in HIV-infected patients (not on antiretroviral therapy) is almost always fatal.
The CFR is estimated at 3% in non-HIV infected patients on TB treatment\cite{2}.
The CFR in HIV-infected patients on TB treatment (even on antiretroviral therapy) is higher than in non-HIV infected patients\cite{3}\cite{4}.

Risk factors for poor outcomes of TB treatment (death and relapse) include co-morbidities (e.g. HIV infection, diabetes, COPD), cavities on chest x-ray, high bacillary load and resistance to TB drugs.

References


1.5 Factors modifying tuberculosis epidemiology

Five major factors influence TB epidemiology: (1) socioeconomic conditions, (2) TB treatment, (3) HIV infection, (4) diabetes and (5) BCG vaccination.

1.5.1 Socioeconomic conditions

The principal factors leading to a reduction in TB cases are improved social and housing conditions. Most cases occur in low-income countries. In industrialised countries, TB generally affects the most disadvantaged social groups.

1.5.2 Tuberculosis treatment

Diagnosing and initiating effective treatment in a patient early during their TB disease before they can infect multiple people is considered the most effective preventive measure against TB. Once an effective TB treatment is started, there is a rapid reduction in transmission. Since the introduction of TB treatment, the risk of TB infection decreased by approximately 10% per year in industrialised countries. This trend was observed in countries with a BCG vaccination programme as well as in those without one. Detection programmes, diagnosis and treatment of TB contributed to this reduction in the risk of TB infection.

1.5.3 HIV infection

Immunodeficiency induced by HIV infection is a major risk factor for progression to active TB and has a considerable impact on the epidemiology of TB. While the lifetime risk of developing active TB in the general population is 5 to 10% after infection with M. tuberculosis, this risk is approximately 10% per year in patients co-infected with HIV and M. tuberculosis. Approximately 8% of incident TB cases in the world are among HIV-infected patients (highest in the WHO African Region, more than 50% in parts of southern Africa).

1.5.4 Diabetes

The risk of TB among people with diabetes is higher than among those without diabetes. It is estimated that diabetes contributes to 15% of TB cases worldwide. Diabetes is also associated with poor absorption of TB drugs and therefore higher rates of drug resistant tuberculosis (DR-TB).

1.5.5 BCG vaccination

Effectiveness of BCG at the individual level

BCG vaccination, if given at birth, is highly effective against the severe forms of TB (miliary and meningitis) in children.

Epidemiological impact of vaccination

Despite some protection from the BCG vaccination, the impact of BCG vaccination on TB transmission and the TB epidemic is considered negligible.

1.5.6 Other factors

Other modifying factors include infection prevention and control measures (Chapter 14) and treatment of LTBI (Chapter 16). The degree to which in a given context the TB epidemiology is affected by these measures is not known.

References


1.6 Epidemiological indicators

When a National TB Programme (NTP) functions well, indicators can be obtained from the local authorities and NTP. The WHO tuberculosis country profiles also provide an estimation of TB indicators by individual country\(^a\).

**Box 1.1 - Most common indicators**

<table>
<thead>
<tr>
<th>Indicator</th>
<th>Description</th>
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<tbody>
<tr>
<td><strong>Annual incidence rate of TB cases(^a)</strong></td>
<td><em>Numerator:</em> number of new TB cases (all forms) that occur in a population over one year  &lt;br&gt;  <em>Denominator:</em> population at the start of the year</td>
</tr>
<tr>
<td><strong>Annual incidence rate of smear-positive PTB cases(^a)</strong></td>
<td><em>Numerator:</em> number of new smear-positive PTB cases that occur in a population over one year  &lt;br&gt;  <em>Denominator:</em> population at the start of the year</td>
</tr>
<tr>
<td><strong>Prevalence of smear-positive PTB cases</strong></td>
<td><em>Numerator:</em> number of smear-positive PTB cases  &lt;br&gt;  <em>Denominator:</em> population at the start of the period of time</td>
</tr>
<tr>
<td><strong>Proportion of multidrug- and rifampicin-resistant TB cases among TB cases</strong></td>
<td><em>Numerator:</em> number of multidrug- and rifampicin-resistant TB cases  &lt;br&gt;  <em>Denominators:</em>  &lt;br&gt;  - Total number of TB cases  &lt;br&gt;  - Number of new TB cases  &lt;br&gt;  - Number of previously treated TB cases</td>
</tr>
<tr>
<td><strong>Proportion of extensively drug-resistant TB cases among TB cases</strong></td>
<td><em>Numerator:</em> number of extensively drug-resistant cases  &lt;br&gt;  <em>Denominators:</em> as for multidrug- and rifampicin-resistant TB cases</td>
</tr>
<tr>
<td><strong>Proportion of HIV-infected patients among new TB cases</strong></td>
<td><em>Numerator:</em> number of HIV-infected patients  &lt;br&gt;  <em>Denominator:</em> number of new TB cases</td>
</tr>
</tbody>
</table>

(a) The rate is expressed as the number of new TB cases (or new smear-positive PTB cases) per 100,000 population.
(b) Prevalence is expressed as the number of smear-positive PTB cases per 100,000 population. It includes new and pre-existing cases. Prevalence represents approximately double the incidence rate.
(c) Proportion is expressed in %.

**Footnotes**

(a) For more information: [https://worldhealthorg.shinyapps.io/tb_profiles/?_inputs_&entity_type=%22group%22&lan=%22FR%22](https://worldhealthorg.shinyapps.io/tb_profiles/?_inputs_&entity_type=%22group%22&lan=%22FR%22)
1.7 Global burden of tuberculosis

1.7.1 Latent tuberculosis infection

The global prevalence of LTBI is unknown due to difficulties in diagnosis. However, WHO estimates that one-quarter of the world population has LTBI.[1]

1.7.2 Active tuberculosis

Globally, active TB remains a leading cause of death from infectious disease.

WHO estimates that each year there are approximately 10 million incident cases of TB and 1.5 million deaths due to TB, including 1.3 million among HIV-negative individuals and 214,000 among HIV-infected individuals.[2]

Patients under 15 years account for 11% of all estimated TB cases.[2] However, TB cases in children are frequently undiagnosed and unreported.

While the absolute number of global TB cases is stable, there are large individual country and regional differences in incidence and prevalence.

Most TB cases are in Southeast Asia (43%), Africa (25%) and the Western Pacific (18%), with lower percentages in the Eastern Mediterranean, the Americas and Europe.[2]

1.7.3 Drug-resistant tuberculosis

Drug-resistant TB (DR-TB) is a growing worldwide problem, and no region is spared.

WHO estimates that annually worldwide there are[3]:

• More than one million rifampicin-susceptible and isoniazid-resistant TB (Hr-TB) cases (11% of all incident TB cases).
• 3.3% of new cases and 18% of previously treated cases, with multidrug-resistant TB (MDR-TB) and rifampicin-resistant TB (RR-TB) representing 465,000 cases and 182,000 deaths.

In Eastern Europe and Central Asia, TB incidence is lower than in Southeast Asia and Africa, but up to 30% of new and 65% of retreatment cases exhibit rifampicin-resistance.

In China and India, there is a low proportion of rifampicin-resistant cases among all TB cases. However, because of their large populations, these two countries represent 41% of global MDR/RR-TB cases.

Resources for detecting drug resistance are limited in many parts of Africa. However, available data suggest that the MDR-TB burden is significant, especially in the south.

The prevalence of extensively drug-resistant TB (XDR-TB), according to the new WHO definition, is currently unknown.

Footnotes
(a) Multidrug-resistant: resistance to at least rifampicin and isoniazid.
(b) Rifampicin-resistant: resistance to rifampicin, with or without resistance to other TB drugs.
(c) Extensively drug-resistant: rifampicin-resistance with resistance to any fluoroquinolone, and at least either bedaquiline or linezolid.

References

https://apps.who.int/iris/rest/bitstreams/1312164/retrieve
Chapter 2: Clinical presentation

2.1 Pulmonary tuberculosis

2.2 Extrapulmonary tuberculosis

2.3 Disseminated or miliary tuberculosis

2.4 Clinical presentation in HIV-infected patients

2.5 Summary of clinical presentations of tuberculosis

Update: January 2022
2.1 Pulmonary tuberculosis

Prolonged cough (more than 2 weeks), with or without sputum production, is a common symptom in patients with pulmonary tuberculosis (PTB).

Other frequent, less specific, signs and symptoms include weight loss, anorexia, fatigue, haemoptysis (blood in sputum), shortness of breath, chest pain, moderate fever and night sweats.

Signs and symptoms may vary between individuals and generally evolve in a chronic, insidious manner. History-taking is therefore of the utmost importance.

Advanced forms and complications are common:
- Respiratory insufficiency due to extensive lesions and destroyed lungs;
- Massive haemoptysis due to large cavities with hyper-vascularisation and erosion of vessels;
- Pneumothorax due to the rupture of a cavity in the pleural space.

In endemic areas, the diagnosis of PTB should be considered in any patient consulting for respiratory symptoms lasting more than 2 weeks.

Table 2.1 provides a differential diagnosis of PTB for non-HIV infected patients.

**Table 2.1 - Differential diagnosis for PTB (non-HIV infected patients)**
<table>
<thead>
<tr>
<th>Diseases</th>
<th>Remarks</th>
</tr>
</thead>
</table>
| **Bacterial pneumonia**        | • Usually more acute and shorter in duration; high fever often present.  
                                 | • Response to antibiotics with no anti-TB activity suggests bacterial pneumonia.  
                                 | • Lobar consolidation is typical of bacterial pneumonia; however, CXR alone cannot differentiate PTB from bacterial pneumonia. |
| **Pulmonary abscess**          | • May arise from aspiration in individuals with impaired consciousness (coma, intoxication with alcohol/drugs, etc.).  
                                 | • Foul-smelling, purulent sputum.  
                                 | • Cavities typically have a thick wall and air fluid levels. |
| **Bronchiectasis**             | • Frequent complication of successive, poorly-treated bronchopulmonary infections in tropical regions.  
                                 | • Characterised by chronic or repeated episodes of productive cough.  
                                 | • Hemoptysis, usually mild, can be present. |
| **Lung cancer**                | • History of smoking or environmental exposure (working in a mine, etc.).  
                                 | • Haemoptysis in 20 to 50% of patients. |
| **Paragonimiasis** (lung flukes)| • To be ruled out in presumed PTB cases in endemic areas (certain areas of Southeast Asia, West Africa and Latin America). |
| **Pulmonary echinococcosis**   | • In Latin America, the Middle East, some Sub-Saharan African countries and China.  
                                 | • Lung involvement may cause chronic cough, with or without haemoptysis.  
                                 | • Cysts can mimic TB cavities. |
| **Pneumocystosis**             | • Common in patients with advanced HIV disease and patients receiving long-term, even low dose, corticosteroid therapy. |
| **Less common diseases**       | • Silicosis, sarcoidosis, melioidosis.  
                                 | • Cryptococcosis, aspergillosis, histoplasmosis. |

For differential diagnosis in HIV-infected patients see Section 2.4.
2.2 Extrapulmonary tuberculosis

Starting from a pulmonary localisation (primary infection), *M. tuberculosis* can spread to other organs during a silent phase, usually soon after primary infection (Chapter 1). Active TB can develop in many other parts of the body, particularly in lymph nodes, meninges, bones and joints, kidneys, genital organs and the abdominal cavity.

Extrapulmonary tuberculosis (EPTB) can develop at any age. Due to relative immunodeficiency, young children, HIV-infected and malnourished patients are more at risk of developing EPTB.

Approximately 16% of global TB cases are classified as EPTB, although this figure varies according the local epidemiology[^1].

A patient with EPTB may also have pulmonary involvement, which should be searched for whenever EPTB is diagnosed or suspected. Table 2.3 at the end of this chapter summarises the characteristics of EPTB.

### 2.2.1 Lymph node tuberculosis

Lymph node TB is common, particularly in certain areas of Africa and Asia, and especially in children and HIV-infected patients.

The presentation of lymph node TB is a non-inflammatory adenopathy. Nodes are cold and painless, multiple (usually bilateral) or single, evolving in a chronic mode towards softening and fistulisation. Cervical localisation is most frequent. Axillary and mediastinal localisations are also common. Other sites may be involved.

Diagnosis may be clinical, but whenever possible, fine needle aspiration should be performed (Chapter 3 and Appendix 7).

Adenopathy usually disappears within 3 months of treatment initiation. Paradoxical reactions may occur at the beginning of treatment (appearance of abscesses, fistulas or other lymph nodes), but a change in the treatment is not required.

Differential diagnoses include malignancies (lymphoma, leukaemia, ear/nose/throat tumours, Kaposi sarcoma) and other infections (bacterial, viral, non-tuberculosis mycobacteria, toxoplasmosis, HIV infection, syphilis, African trypanosomiasis).

### 2.2.2 Tuberculous meningitis

TB meningitis is a serious form of TB that affects the meninges. It is most common in children under 2 years and in HIV-infected patients. It is a medical emergency. Any delay in diagnosis or treatment will result in irreversible neurological sequelae or death[^2].

TB meningitis typically has a subacute insidious course over days or weeks. Symptoms include headaches, irritability, fever, vomiting and altered mental status, which worsen if treatment is delayed. The meningeal syndrome (stiff neck, hypotonia in infants, photophobia and headache) is present in most cases. Third cranial nerve palsy (oculomotor paralysis) may occur.

Diagnosis is assisted by examination of cerebrospinal fluid (Chapter 3). The main differential diagnoses are other forms of meningitis.

### 2.2.3 Tuberculosis of bones and joints

Up to 40% of patients with TB of bones and joints have concurrent PTB[^3].

**Spinal TB (spondylodiscitis or Pott’s disease)**

TB can affect vertebrae and intervertebral disks, causing destruction and deformation of the spine. The thoracic spine is the most frequently affected.

Localised back pain may precede by several months the appearance of the first radiological anomalies (destruction of an intervertebral disk).

A spinal prominence (gibbus) due to destruction and deformity of the vertebral bodies may be felt.

Paravertebral cold abscesses and/or neurological complications can develop.

A missed diagnosis of thoracic or cervical spinal TB can result in paralysis.

**Arthritis**

TB most frequently causes a chronic mono-arthritis, starting insidiously, with little or no pain and accompanied by joint destruction. The joints most often affected are the hips, knees, elbows and wrists.
Osteitis
Osteitis is the least common presentation of TB of the bones. It may be a primary osteitis or an osteitis secondary to TB arthritis. Typically, long bones are affected. Cold abscesses may occasionally occur. Like arthritis, it is distinguished from common bacterial infections by the presence of mild symptoms, despite bone and joint destruction.

The diagnosis is based on the patient’s history, clinical examination and radiography, as biopsy and culture are difficult to perform in many settings. A history of prolonged and insidious osteitis or arthritis associated with a deterioration of the general physical condition favours TB aetiology, as opposed to bacterial osteomyelitis or brucellosis. The patient may have a history of non-response to antibiotics.

2.2.4 Urogenital tuberculosis
Renal involvement is frequent and may be asymptomatic for a long period, with a slow development of signs and symptoms: painful urination (dysuria), urinary urgency and frequency (pollakiuria), including during the night (nocturia); back/abdominal pain; tenderness/swelling of the testes or epididymitis or haematuria. General physical condition is generally preserved. Diagnosis is suspected in the presence of pyuria (white blood cells in the urine) and micro- or macroscopic haematuria, which does not respond to antibiotics. Examination of the urine helps with diagnosis (Chapter 3).

In men, genital localisation is secondary to renal involvement. Signs are most often epididymitis with scrotal pain.

In women, genital tract infection can also occur by a hematogenous path. Signs are non-specific: pelvic pain, leucorrhoea and abnormal vaginal bleeding. Infertility is often the reason leading women to seek medical attention.

Extension may be found in the peritoneum, with resulting ascites.

2.2.5 Abdominal tuberculosis
Abdominal TB commonly presents as ascites resulting from the peritoneal localisation of the infection. Abdominal mass (often in the right lower quadrant), pain and diarrhoea may be present. The frequency of chronic ascites in tropical regions, with its many different causes, makes this relatively uncommon form of TB difficult to diagnose.

Diagnosis is assisted by examination of the ascitic fluid via paracentesis (Chapter 3). Constitutional symptoms (fever, night sweats, malaise and weight loss) may be present. Accumulation of ascites may mask weight loss.

2.2.6 Tuberculous pleural effusion
Tuberculous pleural effusion is one of the most common forms of EPTB. It is often asymptomatic, especially if less than 300 ml. Shortness of breath and chest pain (often unilateral) occur when the effusion is large. Sputum production and cough are present in the case of concurrent PTB, which is common.

Constitutional symptoms such as fever, night sweats, malaise and weight loss may also be present.

Effusion can progress to tuberculous empyema, characterised by purulent fluid containing large numbers of bacilli. Tuberculous empyema is often associated with thickened, scarred and calcified pleura.

Diagnosis is assisted by examination of the pleural fluid via paracentesis and chest x-ray (CXR). See Chapter 3.

2.2.7 Tuberculous pericardial effusion
Clinical signs of a tuberculous pericardial effusion include chest pain, shortness of breath, oedema of the lower limbs and sometimes ascites.

Clinical examination may show pericardial friction rub, raised jugular pressure and tachycardia. CXR and ultrasound are key elements for diagnosis (Chapter 3).

Pericardiocentesis may be necessary in the event of acute heart failure with haemodynamic compromise. It must be performed by experienced personnel in well-equipped hospitals, and when possible, under direct visualisation with ultrasound.

2.2.8 Cutaneous tuberculosis
The clinical presentation of cutaneous TB is chronic, painless, non-pathognomonic lesions, ranging from small papula and erythema to large tuberculomas.

The diagnosis is based on culture from a biopsy.
References

   [https://apps.who.int/iris/rest/bitstreams/1312164/retrieve](https://apps.who.int/iris/rest/bitstreams/1312164/retrieve)


   [https://doi.org/10.1186/1471-2334-9-44](https://doi.org/10.1186/1471-2334-9-44)
2.3 Disseminated or miliary tuberculosis

Miliary TB is a generalised massive infection characterised by hematogenous diffusion of *M. tuberculosis* throughout the body. It is a medical emergency.

The disease may manifest as a miliary pattern, or very small nodular elements ('millet seeds') in the lungs.

The classic acute form is mostly found in children, young adults and HIV-infected patients. The presentation can be either abrupt or insidious, with progressive deterioration in the patient's physical condition. The clinical picture is often completed within one to two weeks and is characterised by a profoundly altered physical condition, marked wasting, headache and constant high fever. Discrete dyspnoea and coughing suggest a pulmonary focus; however, lungs can often be clear on auscultation. A moderate hepatosplenomegaly is occasionally found. Certain forms of miliary TB evolve in a subacute manner over several months. Given this non-specific clinical picture, typhoid fever and septicaemia should be considered in the differential diagnosis.

Diagnosis of miliary TB is confirmed by CXR (Chapter 3).

When feasible, fundoscopy may reveal choroidal tubercles.

Sputum smear examination is usually negative.

When there is no possibility of obtaining CXR, the lack of response to antibiotics is an argument in favour of miliary TB.

The tuberculin skin test is more likely to be falsely negative than in any other form of TB.

In children, the risk of meningitis (20-40%)[^1] is high. Lumbar puncture should be routinely performed if miliary TB is suspected.

References

2.4 Clinical presentation in HIV-infected patients

Among HIV-infected patients, TB is the most common opportunistic infection and the leading cause of morbidity and mortality\[^{[1]}\]. According to the WHO clinical staging system for HIV/AIDS, patients with PTB are in clinical stage 3 and patients with EPTB in clinical stage 4\[^{[2]}\].

In the early stages of HIV infection, when the immune system is functioning relatively normally, the clinical signs of TB are similar to those in seronegative individuals.

As the immune system deteriorates in later stages of the disease, smear-negative PTB, disseminated TB and EPTB become more common. These cases are more difficult to diagnose, and have a higher fatality rate than smear-positive PTB cases. Patients may have difficulty expectorating, so more advanced sputum collection techniques may be necessary (Chapter 3 and Appendix 3).

Algorithms presented in Chapter 4 use clinical criteria combined with laboratory and other investigations to help diagnose TB in HIV-infected individuals.

Table 2.2 provides a differential diagnosis of PTB in HIV-infected patients.

<table>
<thead>
<tr>
<th>Diseases</th>
<th>Comments</th>
</tr>
</thead>
</table>
| Other pneumonia (bacterial, viral, atypical) | • Bacterial pneumonia (most often *S. pneumoniae, H. influenzae*) is common at all stages of HIV infection.  
• Atypical pneumonia (*M. pneumoniae, C. pneumoniae*) and viral pneumonia are possible at any CD4 count, except in the case of cytomegalovirus, which occurs at CD4 < 50. |
| Pneumocystosis (*Pneumocystis jirovecii* pneumonia or PCP or PJP) | • Pneumocystis has many characteristics in common with PTB (insidious onset, persistent cough, fever) but tends to occur in the advanced stages of HIV infection (CD4 < 200).  
• Pneumocystis is unlikely in patients taking co-trimoxazole prophylaxis.  
• It imparts a greater degree of dyspnoea, rarely produces effusions, and is not usually accompanied by haemoptysis. |
| Pulmonary Kaposi’s sarcoma (KS) | • KS can resemble PTB, with slow onset of cough, fever, haemoptysis, night sweats and weight loss. It is a disease of advanced stage HIV, and in most cases, is preceded or accompanied by lesions involving the skin and mucus membranes. |
| Less common diseases | • Pulmonary cryptococcosis, histoplasmosis and other fungal infections.  
• Pulmonary nocardiosis: on direct smear, nocardia are weakly acid-fast, and similar in appearance to mycobacteria (although they are branching filamentous bacilli, particularly on Gram staining). |

The most common EPTB in HIV-infected patients are miliary TB, TB meningitis and diffuse lymphadenopathy in children, and lymph node TB, pleural effusion, pericarditis, TB meningitis and miliary TB in adults.

Immune reconstitution inflammatory syndrome (IRIS) is a clinical presentation of TB in patients starting antiretroviral therapy. See Chapter 12 for clinical presentation and management of IRIS.

References

https://apps.who.int/iris/handle/10665/43699
2.5 Summary of clinical presentations of tuberculosis

Table 2.3 - Clinical presentations and considerations for HIV-infected patients
<table>
<thead>
<tr>
<th>Sites</th>
<th>Clinical presentations</th>
<th>Considerations for HIV patients</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Pulmonary TB</strong></td>
<td>• Prolonged cough (&gt; 2 weeks), with or without sputum production.</td>
<td>• Fever and weight loss are more common and pronounced.</td>
</tr>
<tr>
<td></td>
<td>• Weight loss, anorexia, fatigue, shortness of breath, chest pain, moderate fever, night coughs, haemoptysis.</td>
<td>• Cough and haemoptysis may be less common (less inflammation and cavity formation).</td>
</tr>
<tr>
<td></td>
<td></td>
<td>• See algorithms, Chapter 4.</td>
</tr>
<tr>
<td><strong>Disseminated miliary TB</strong></td>
<td>• Non-specific symptoms: high fever, headache, weight loss.</td>
<td>• May be confused with severe wasting in advanced HIV disease.</td>
</tr>
<tr>
<td></td>
<td>• Deterioration over days or weeks.</td>
<td>• <em>M. tuberculosis</em> sometimes isolated from blood cultures.</td>
</tr>
<tr>
<td></td>
<td>• Simultaneous involvement of multiple organs.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>• High risk of meningitis in children.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>• Miliary findings CXR.</td>
<td></td>
</tr>
<tr>
<td><strong>Lymph nodes TB</strong></td>
<td>• Most often in cervical region.</td>
<td>• HIV infection can cause persistent generalised lymphadenopathy (PGL). PGL lymph nodes are painless, and symmetrical. Posterior cervical or epitrochlear nodes are often involved.</td>
</tr>
<tr>
<td></td>
<td>• Non-inflammatory, painless node &gt; 2 cm, chronic (&gt; 4 weeks); fistulisation possible.</td>
<td>• Other common causes of lymphadenopathy include lymphoma, carcinomatous metastases, Kaposi sarcoma.</td>
</tr>
<tr>
<td><strong>TB meningitis</strong></td>
<td>• Subacute, insidious.</td>
<td>• Rule out cryptococcal meningitis: perform antigen test on serum and CSF.</td>
</tr>
<tr>
<td></td>
<td>• Headaches, irritability, fever, altered mental status.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>• Meningeal syndrome usually present.</td>
<td></td>
</tr>
<tr>
<td><strong>Bone and joint TB</strong></td>
<td>• Monoarthritis with joint destruction and little or no pain.</td>
<td>• Multifocal disease more common.</td>
</tr>
<tr>
<td></td>
<td>• Deformity of the spine (Pott’s disease).</td>
<td></td>
</tr>
<tr>
<td><strong>Urogenital TB</strong></td>
<td>• Renal: urinary symptoms, few constitutional symptoms; suspected when no response to antibiotics for urinary infection.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>• Non-specific gynaecological symptoms, infertility or epididymitis with scrotal pain.</td>
<td></td>
</tr>
<tr>
<td><strong>Abdominal TB</strong></td>
<td>• Ascites (may mask weight loss).</td>
<td>• PTB is more frequently associated.</td>
</tr>
<tr>
<td></td>
<td>• Abdominal mass, pain, diarrhoea.</td>
<td></td>
</tr>
<tr>
<td><strong>Effusions</strong></td>
<td>• Pleural: pleuritic chest pain, dyspnoea.</td>
<td>• Serious effusions are common.</td>
</tr>
<tr>
<td></td>
<td>• Pericardial: chest pain, dyspnoea, lower limb oedema or ascites, pericardial friction rub.</td>
<td>• TB is the most likely aetiology in high TB–HIV prevalence settings.</td>
</tr>
</tbody>
</table>
TB is considered as non-severe if the following criteria are met:

- negative smear microscopy, and
- uncomplicated PTB with a small infiltrate confined to one lobe and no cavities, or
- uncomplicated extra-thoracic lymph node TB, or
- uncomplicated intrathoracic lymph node TB.
Chapter 3: Diagnosis and follow-up investigations

3.1 Active tuberculosis

3.2 Latent tuberculosis infection

3.3 Other investigations

Update: October 2022
3.1 Active tuberculosis

3.1.1 Introduction

Active tuberculosis (TB) is bacteriologically confirmed by the detection of *M. tuberculosis* complex through different bacteriological tests. These tests detect either the organism (smear microscopy and culture), or some of its genetic material (genotypic tests, including rapid molecular tests and genome sequencing).

Specimens used for bacteriological testing include respiratory specimens (sputum, nasopharyngeal aspirate and, in children, gastric aspirate) and extrapulmonary specimens (Table 3.6).

Drug susceptibility testing (DST) is indicated for all patients with confirmed TB. It can be performed using genotypic or phenotypic tests:
- Genotypic DST (gDST) can detect resistance to TB drugs by identifying specific gene mutations.
- Phenotypic DST (pDST) can detect resistance to TB drugs by measuring the growth of *M. tuberculosis* in the presence of the drug.

To diagnose TB and determine the appropriate regimen at baseline:
- All patients should be tested with a rapid molecular test (RMT) to detect *M. tuberculosis* and rifampicin resistance.
- Whatever the result of the rifampicin susceptibility test (resistance detected or not), all patients, if possible, should be tested with an RMT for isoniazid resistance and at least those with high risk of isoniazid resistance (for definition of high risk of resistance, see below).
- All patients with rifampicin resistance should be tested for resistance to fluoroquinolones and other TB drugs.
- All patients with isoniazid resistance and rifampicin susceptibility should be tested for resistance to fluoroquinolones.
- Culture, pDST and genome sequencing may be required.

In limited-resource settings, resistance to TB drugs should be investigated in priority in patients with:
- High risk of mortality: e.g. HIV-infected patients or patients with extensive disease.
- High risk of resistance: patients with previous TB treatment, or in contact with a TB case resistant to TB drug(s), or coming from an area of high prevalence of resistance to TB drug(s).

Notes:
- Negative bacteriological tests for *M. tuberculosis* does not rule out TB.
- A negative DST does not necessarily rule out drug resistance.

Other investigations can assist TB diagnosis. These investigations include: lateral flow urine lipooarabinomannan assay (LF-LAM) which detects an antigen of *M. tuberculosis* cell wall excreted in urine, medical imaging, and some biological tests.

3.1.2 Rapid molecular tests

RMTs are nucleic acid amplification tests (NAATs). They can detect *M. tuberculosis* and drug resistance by identifying resistance-conferring mutations in certain genes (Table 3.1). Other drug resistance-conferring mutations may be present, but not detected by RMTs. In areas where prevalence of these mutations is high, RMT sensitivity may be decreased.

**Table 3.1** – Rapid molecular tests and detection of drug resistance
RMTs have a good specificity, but are less sensitive than culture. Their various levels of complexity determine their use at different levels of health facilities. Low complexity RMTs are preferred in routine practice.

### Low complexity nucleic acid amplification tests

#### Xpert assays

Xpert assays are almost fully automated. An uninterrupted power supply and a computer are required to perform and read assays.

Xpert assays can be performed on:
- Respiratory specimens
- Extrapulmonary (EP) specimens:
  - Lymph node biopsy or aspirate: suspicion of lymph node TB or detection of rifampicin resistance in clinically diagnosed lymph node TB;
  - Cerebrospinal fluid (CSF): suspicion of TB meningitis;
  - Pleural fluid: suspicion of TB with pleural effusion;
  - Stool: suspicion of PTB in children;
  - Pericardial fluid: suspicion of TB with pericardial effusion (sampling to be performed only by experienced clinicians);
  - Urine: suspicion of genitourinary TB; suspicion of disseminated TB in HIV-infected patients;
  - Synovial fluid: suspicion of TB arthritis;
  - Peritoneal fluid: suspicion of abdominal TB;
  - Blood: suspicion of disseminated TB in HIV-infected patients.

<table>
<thead>
<tr>
<th>Tests</th>
<th>TB drug resistance (targeted genes)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Low complexity NAATs</strong></td>
<td>Rifaampicin (rpoB)</td>
</tr>
<tr>
<td>• Xpert MTB/RIF</td>
<td></td>
</tr>
<tr>
<td>• Xpert MTB/RIF Ultra</td>
<td></td>
</tr>
<tr>
<td>• Truenat MTB-RIF Dx</td>
<td></td>
</tr>
<tr>
<td><strong>Moderate complexity NAATs</strong></td>
<td></td>
</tr>
<tr>
<td><strong>High complexity NAATs</strong></td>
<td></td>
</tr>
<tr>
<td>• GenoType MTBDRplus (V2.0)</td>
<td></td>
</tr>
<tr>
<td>• Genoscholar NTM+MDRTB II</td>
<td></td>
</tr>
<tr>
<td><strong>Low complexity NAATs</strong></td>
<td>Isoniazid high-level resistance (katG)</td>
</tr>
<tr>
<td>• Xpert MTB/XDR</td>
<td></td>
</tr>
<tr>
<td><strong>Moderate complexity NAATs</strong></td>
<td>Isoniazid low-level resistance, thionamides (a) (inhA promoter)</td>
</tr>
<tr>
<td><strong>High complexity NAATs</strong></td>
<td></td>
</tr>
<tr>
<td>• GenoType MTBDRplus (V2.0)</td>
<td></td>
</tr>
<tr>
<td>• Genoscholar NTM+MDRTB II</td>
<td></td>
</tr>
<tr>
<td><strong>Low complexity NAATs</strong></td>
<td>Fluoroquinolones (gyrA, gyrB) (b)</td>
</tr>
<tr>
<td>• Xpert MTB/XDR</td>
<td></td>
</tr>
<tr>
<td><strong>High complexity NAATs</strong></td>
<td>Aminoglycosides (rrs, eis)</td>
</tr>
<tr>
<td>• GenoType MTBDRsl (V2.0)</td>
<td></td>
</tr>
<tr>
<td><strong>High complexity NAATs</strong></td>
<td>Pyrazinamide (pncA)</td>
</tr>
<tr>
<td>• Genoscholar PZA-TB II</td>
<td></td>
</tr>
</tbody>
</table>

(a) Mutations in other genes can result in resistance to thionamides. Consequently, absence of inhA mutation does not rule out resistance.

(b) Specific mutations in gyrA (e.g. mutations recognized by the probes MUT3B, 3C, 3D) are associated with high-level fluoroquinolones resistance.

RMTs have a good specificity, but are less sensitive than culture. Their various levels of complexity determine their use at different levels of health facilities. Low complexity RMTs are preferred in routine practice.
Xpert MTB/RIF and Xpert MTB/RIF Ultra assays provide simultaneously results for *M. tuberculosis* detection and rifampicin resistance.

Sensitivity of Xpert MTB/RIF Ultra assay is higher than that of Xpert MTB/RIF assay. It provides a result “trace” corresponding to the lowest bacillary load for *M. tuberculosis* detection. It is preferred for HIV-infected patients, children, EP specimens, and sputum smear-negative specimens. Its specificity is lower in patients with a history of TB, as a “trace” result may indicate that the specimen contains fragments of dead bacilli.

WHO has validated their use on lymph node biopsy or aspirate, CSF, and pleural fluid, synovial fluid[2]. Xpert assays can be performed on any biopsy specimens (lymph node, bone, skin, resection material, etc.) with good performance. Xpert assays have shown acceptable performances in various studies on other specimens (peritoneal[9] and pericardial[4] fluids, stools[5][6][7], and urine[8][9]). Xpert assays on blood have a low sensitivity compared to culture and are not routinely recommended[10].

Xpert MTB/XDR assay detects resistance to isoniazid (low- and high-level), fluoroquinolones (low- and high-level), aminoglycosides, and thionamides. It does not detect resistance to rifampicin.

Xpert MTB/XDR assay employ the same platform as other Xpert assays, but require a 10-colour module instead of the 6-colour module used for Xpert MTB/RIF and Xpert MTB/RIF Ultra assays. The 10-colour module can also read Xpert MTB/RIF and Xpert MTB/RIF Ultra assays.

Xpert MTB/XDR assay should be used:

- When resistance to rifampicin has been detected by Xpert MTB/RIF or Xpert MTB/RIF Ultra, to detect resistance to other drugs.
- When *M. tuberculosis* has been detected by Xpert MTB/RIF or Xpert MTB/RIF Ultra or culture, to detect resistance to isoniazid in all patients, if possible, and at least those with high risk of isoniazid resistance (Section 3.1.1).
- Before using a fluoroquinolone containing regimen in isoniazid-resistant TB (Hr-TB), multidrug-resistant (MDR-TB), rifampicin-resistant TB (RR-TB) or drug-susceptible TB treated with the regimen 2HPZ-Mfx/2HP-Mfx.
- Before treating drug-susceptible TB meningitis with the regimen 6HRZ-Eto.
- In patients with fluoroquinolone-susceptible TB, initially treated with a fluoroquinolone-containing regimen, and presenting a smear-positive microscopy at Month 2 or later.

Table 3.2 – Main performances of Xpert assays
<table>
<thead>
<tr>
<th>Xpert assays</th>
<th>Performances</th>
</tr>
</thead>
</table>
| **MTB/RIF** | Detection of *M. tuberculosis* (MTB) compared to culture:  
  - Sensitivity in respiratory specimens\(^{[11]}\):  
    - sputum-smear positive: 99%  
    - sputum smear-negative: 68%  
    - HIV-infected patients: 79%  
    - children: see Appendix 1.  
  - Sensitivity in EP specimen: see Appendix 1.  
  - Specificity: very high in all specimens (99%), i.e. a positive result is unlikely to be a false positive.  
  
  Detection of rifampicin resistance compared to pDST\(^{[11]}\):  
  Sensitivity: 95%; specificity: 98% |
| **MTB/RIF Ultra** | Detection of MTB in respiratory and EP specimens\(^{[12]}\):  
  - Sensitivity: + 5% compared to Xpert MTB/RIF  
  - Specificity: - 3.2% compared to Xpert MTB/RIF; - 5.4% in patients with a history of TB  
  No result for rifampicin resistance if “trace” result. |
| **MTB/XDR** | Detection of MTB in respiratory and EP specimens (children and adults):  
  As Xpert MTB/RIF.  
  Detection of resistances compared to pDST\(^{[13]}\):  
  - To isoniazid (low- and high-level): sensitivity: 94.2%; specificity: 98%  
  - To fluoroquinolones (low- and high-level): sensitivity: 93.1%; specificity: 98.3%  
  - To aminoglycosides: sensitivity: 86.1%; specificity: 98.9%  
  - To thionamides: sensitivity: 51.7%; specificity: 98.3% |

For more information on specimen processing and Xpert instruments see Appendix 1.
For interpretation of Xpert assay results see Appendix 2.
For request form see Appendix 3.

**2) Truenat assays**

Truenat assays require:
- Several manual steps (pipetting).
- Sequential testing for *M. tuberculosis* detection (Truenat MTB Plus), then for rifampicin resistance detection (Truenat MTB-RIF Dx).
- Separate kits for specimen preparation, DNA extraction, DNA amplification, and detection of *M. tuberculosis* and rifampicin resistance.

Truenat MTB Plus can only be performed on sputum specimens (positive or negative smear microscopy). It is not recommended for other respiratory specimens or EP specimens\(^{[2][7]}\).  
Specificity is high, i.e. a positive result is unlikely to be a false positive\(^{[14]}\).  
Tests can be run at room temperatures of up to 40 °C and humidity of up to 80%. Truenat instruments are battery-operated and can be used in peripheral or mobile health facilities.
Interpretation of results is the same as for Xpert (Appendix 2).

**Table 3.3** – Main performances of Truenat assays
3) TB-LAMP

Although validated by WHO, this test has major limitations:
- It does not detect rifampicin resistance.
- Its sensitivity is lower than that of other low complexity NAATs in HIV-infected or smear-negative patients.
- It cannot be used for the diagnosis of extrapulmonary TB (EPTB).

Box 3.1 – Choice of low complexity NAATs

<table>
<thead>
<tr>
<th>Truenat assays</th>
<th>Performances</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>MTB Plus</strong></td>
<td>Detection of MTB in sputum specimens (children and adults) compared to culture:</td>
</tr>
<tr>
<td></td>
<td>- Sensitivity:</td>
</tr>
<tr>
<td></td>
<td>- sputum smear-positive: 80%</td>
</tr>
<tr>
<td></td>
<td>- sputum smear-negative: 55%</td>
</tr>
<tr>
<td></td>
<td>- Specificity: 96% [14]</td>
</tr>
<tr>
<td><strong>MTB-RIF Dx</strong></td>
<td>Detection of rifampicin resistance compared to pDST:</td>
</tr>
<tr>
<td></td>
<td>- Performed on the DNA isolated from sputum specimens with Truenat MTB Plus positive result.</td>
</tr>
<tr>
<td></td>
<td>- Sensitivity: 84%</td>
</tr>
<tr>
<td></td>
<td>- Specificity: 97%</td>
</tr>
</tbody>
</table>

**Moderate complexity nucleic acid amplification tests**

WHO recommends these tests for the simultaneous detection of *M. tuberculosis* and resistance to rifampicin and isoniazid, from smear-positive and negative respiratory specimens, in children and adults, including HIV-infected patients.

Table 3.4 – Performances of moderate complexity NAATs

<table>
<thead>
<tr>
<th>Tests</th>
<th>Performances</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Abbott Real Time MTB and MTB RIF/INH BD MAX MDR-TB</strong></td>
<td>Detection of MTB compared to culture:</td>
</tr>
<tr>
<td><strong>Hain FluoroType MTB and MTBDR</strong></td>
<td>- Sensitivity 93%</td>
</tr>
<tr>
<td><strong>Roche cobas MTB and MTB-INH/RIF</strong></td>
<td>- Specificity 97.7%</td>
</tr>
<tr>
<td><strong>Detection of rifampicin resistance</strong></td>
<td>Compared to pDST:</td>
</tr>
<tr>
<td><strong>Roche cobas MTB and MTB-INH/RIF</strong></td>
<td>- Sensitivity 96.7%</td>
</tr>
<tr>
<td><strong>Hain FluoroType MTB and MTBDR</strong></td>
<td>- Specificity 98.9%</td>
</tr>
<tr>
<td><strong>Detection of isoniazid resistance</strong></td>
<td>Compared to pDST:</td>
</tr>
<tr>
<td><strong>Roche cobas MTB and MTB-INH/RIF</strong></td>
<td>- Sensitivity 86.4%</td>
</tr>
<tr>
<td><strong>Hain FluoroType MTB and MTBDR</strong></td>
<td>- Specificity 99.8%</td>
</tr>
</tbody>
</table>

NAATs of moderate complexity have several limitations:
- Need for space, equipment, qualified staff; only feasible in regional laboratories.
- Their use does not eliminate the need for pDST, high complexity NAATs, or genome sequencing to:
  - test susceptibility to other TB drugs;
  - confirm a negative result in patients at high risk of drug resistance.
- Their use on EP specimens is not validated.
High complexity nucleic acid amplification tests

Line probe assays (LPA) can detect specific rifampicin, isoniazid, fluoroquinolones, aminoglycosides, and pyrazinamide resistance encoding mutations in *M. tuberculosis*.

These tests can be performed on isolates of *M. tuberculosis* (indirect testing). Some can be performed on sputum specimens (direct testing).

NAATs of high complexity have several limitations:
- Need for space, equipment, highly qualified staff; only feasible in reference and national laboratories.
- Risk of cross-contamination (tests are performed in an open system that can lead to the detection of DNA from sources other than the specimen).
- To benefit from the short turnaround time of these tests, efficient logistical support is required to ensure specimens are transported to the laboratory and the results are delivered in a timely manner.
- Their use does not eliminate the need for pDST or genome sequencing to:
  - test sensitivity to other TB drugs;
  - confirm a negative result in patients at high risk of drug resistance.
- Although direct test results can be obtained in 1 to 2 days, for indirect tests, it is necessary to wait the time required for bacterial growth (Appendix 5).
- Their use on respiratory (non-sputum) or EP specimens is not validated.

**Box 3.2 – WHO validated LPAs**

**First-line LPAs**
- GenoType MTBDR plus version 2 (“Hain first line test”): initial test to detect resistance to rifampicin and isoniazid on smear-positive sputum specimens and *M. tuberculosis* isolates. Compared to pDST, sensitivity is 98.2% for rifampicin, and 97.8% for isoniazid; specificity is 95.4% for rifampicin, and 98.8% for isoniazid[18]. On smear-negative sputum specimens, sensitivity is low (44.4%), and its use is not recommended.[13]
- Genoscholar NTM+MDRTB II (“Nipro test”): performances comparable to GenoType MTBDRplus to detect resistance to rifampicin and isoniazid on smear-positive sputum specimens and *M. tuberculosis* isolates. Not recommended on smear-negative sputum specimens. Can differentiate *M. avium*, *M. intracellulare* and *M. kansasii* from other non-tuberculous mycobacteria.
- Genoscholar PZA-TB ll: to detect resistance to pyrazinamide on *M. tuberculosis* isolates. Compared to pDST, sensitivity is 81%, and specificity is 97%.[13]

**Second-line LPA**
GenoType MTBDRs/ version 2 (“Hain second line test”): in patients with confirmed MDR/RR-TB, to detect resistance to fluoroquinolones (high- and low-level) and aminoglycosides on smear-positive or smear-negative sputum specimens and *M. tuberculosis* isolates. The number of “indeterminate” results is higher for smear-negative than for smear-positive sputum specimens. For smear-positive sputum specimens, sensitivity is 93% for fluoroquinolones, and 88.9% for aminoglycosides; specificity is 98.3% for fluoroquinolones, and 91.7% for aminoglycosides[16].

### 3.1.3 Genome sequencing

Genome sequencing can only be performed in highly specialized reference laboratories. It can rapidly:
- Detect mutations associated with TB drug resistance. When available, it is particularly useful to identify:
  - resistance to TB drugs for which pDST is unreliable, or no RMTs are available;
  - mutations missed by RMTs (+ 20% of drug resistance detection compared to RMTs has been described[17]).
- Detect mixed infection (infection with distinct *M. tuberculosis* strains).
- Identify heteroresistance (same strain, with different resistance profiles).
- Differentiate treatment relapse and reinfection with a different strain.

Genome sequencing methods include Sanger sequencing (reference method) and next generation sequencing (NGS). The advantage of NGS is that, unlike Sanger sequencing, it provides results for a large number of genes in a single reaction. NGS results are interpreted by reference laboratories using specific software and mutation databases[6].
Some mutations associated with resistance to recently introduced drugs (e.g. bedaquiline and delamanid) and their therapeutic implications are still not well-known.

The two main NGS techniques are targeted NGS (tNGS) and whole genome sequencing (WGS):
- tNGS (on smear-positive sputum specimens or culture isolates): detection of resistance conferring mutations on 18 selected genes: first-line TB drugs, fluoroquinolones, aminoglycosides, linezolid, bedaquiline, clofazimine, ethionamide (Deepplex®Myc-TB). Used in routine.
- WGS (on culture isolates): detection of resistance conferring mutations on whole genome (i.e. potentially all TB drugs). Used for research.

### 3.1.4 Smear microscopy

The purpose of smear microscopy is to detect acid-fast bacilli (AFB) in stained specimens.

Smear microscopy has several limitations:
- It has a sensitivity lower than RMTs and culture in respiratory specimens (65% compared to culture[11]) and EP specimens (48% compared to culture[18]).
- It has a low sensitivity in patients with low bacillary load in sputum (paucibacillary TB), e.g. children and HIV-infected patients.
- It cannot differentiate between *M. tuberculosis* and non-tuberculous mycobacteria. However, in areas with high TB prevalence, AFB detected on smear microscopy are most likely *M. tuberculosis*.
- It does not determine if bacilli are viable (alive) or non-viable (dead).
- It does not determine susceptibility of the bacilli to TB drugs.

Sputum smear microscopy is no longer the recommended initial diagnostic test for PTB. However, it still plays a role:
- When RMTs are not available.
- For assessing the infectiousness of PTB patients.
- For monitoring the response to TB treatment in patients with:
  - drug-susceptible PTB ([Chapter 9](#)).
  - drug-resistant PTB. However, culture is also required for monitoring treatment response in these patients ([Chapter 10](#) and [Chapter 11](#)).

For improving the sensitivity of smear microscopy:
1) Two sputum specimens should be examined. Approximately 86% of sputum smear-positive patients are identified during the first examination, and an additional 12% during the second. It is not necessary to carry out more than 2 examinations[19].
2) Light-emitting diode (LED) fluorescent microscopy to examine auramine-stained smears is preferred to Ziehl-Neelsen microscopy, as it is more sensitive, and reading is more rapid.

Concentration techniques can also increase the sensitivity of smear microscopy[20].

For sputum specimen collection, storage and shipment see [Appendix 3](#). For sputum smear preparation and staining techniques see [Appendix 4](#). For request form see [Appendix 34](#).

### 3.1.5 Culture

Culture consists of growing *M. tuberculosis* in specific liquid or solid media.
Culture on liquid medium (automated or manual mycobacterial growth indicator tube, MGIT) is the reference method for the diagnosis of PTB and EPTB. Given the long turnaround time and equipment required, it is not used as initial diagnostic test.
Culture on solid medium (Lowenstein-Jensen) is cheaper, less prone to contaminations than cultures on liquid media, but its turnaround time is longer.
Other culture techniques are less commonly used[4].

Culture is necessary to:
- Confirm treatment failure.
- Assess treatment response in patients with drug-resistant PTB ([Chapter 10](#) and [Chapter 11](#)).
- Evaluate treatment outcome in patients with drug-resistant PTB ([Chapter 17](#)).
- Provide isolates for the following tests:
  - First-line LPAs on sputum smear-negative and EP specimens
  - Genoscholar PZA-TB II, regardless of sputum smear positivity
Culture may help to diagnose TB when other bacteriological tests are negative or inconclusive:
- In patients with signs and symptoms of TB and a negative RMT, particularly when resistance is suspected.
- In adults with history of TB in the previous 5 years and showing a “trace” result by Xpert MTB/RIF Ultra.

Culture has several limitations:
- Only specialized laboratories implementing systematic quality assurance procedures can be relied upon for culture (often national reference laboratories or supranational).
- *M. tuberculosis* is a slow-growing bacillus. Positive culture results are obtained after 2 to 4 weeks.

For sputum specimen collection, storage and shipment see Appendix 3.
For the time required to obtain the results see Appendix 5.

### 3.1.6 Phenotypic drug susceptibility testing

Phenotypic DST (pDST) determines if a strain is resistant to a TB drug by evaluating the growth in the presence of the drug. It can determine two levels of resistance (low and high) for isoniazid and fluoroquinolones.

The pDST is essential to detect resistance to drugs for which there are no reliable RMTs, and when genome sequencing is not available.

In addition, pDST may be necessary:
- If an RMT indicates *M. tuberculosis* “detected” and drug resistance “indeterminate”.
- If an RMT indicates drug susceptibility in a patient at high risk of resistance.
- In areas with a high prevalence of mutations not detected by RMTs.

Phenotypic DST is performed on culture isolates by specialized laboratories (often national reference laboratories or supranational).

The pDST is not reliable for all drugs, even when performed by a highly qualified laboratory[^21].

<table>
<thead>
<tr>
<th>Reliability of pDST</th>
<th>TB drugs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Highly reliable</td>
<td>Isoniazid</td>
</tr>
<tr>
<td></td>
<td>Rifampicin</td>
</tr>
<tr>
<td></td>
<td>Fluoroquinolones</td>
</tr>
<tr>
<td></td>
<td>Aminoglycosides</td>
</tr>
<tr>
<td>Unreliable (should not be performed)</td>
<td>Ethambutol</td>
</tr>
<tr>
<td></td>
<td>Ethionamide</td>
</tr>
<tr>
<td></td>
<td>Cycloserine or terizidone</td>
</tr>
<tr>
<td></td>
<td>Para-aminosalicylic acid (or sodium)</td>
</tr>
<tr>
<td></td>
<td>Delamanid</td>
</tr>
<tr>
<td>Reliable, but limited access outside of supranational laboratories</td>
<td>Bedaquiline</td>
</tr>
<tr>
<td></td>
<td>Linezolid</td>
</tr>
<tr>
<td></td>
<td>Clofazimine</td>
</tr>
<tr>
<td>Reliable when performed in a high-quality laboratory (difficult to perform)</td>
<td>Pyrazinamide</td>
</tr>
</tbody>
</table>

### 3.1.7 Summary of bacteriological tests


The tables below provide an overview of the specimens that can be used for each test, and of the tests that can detect resistance to each TB drug.

**Table 3.6 – Specimens for bacteriological tests**

<table>
<thead>
<tr>
<th>Tests</th>
<th>Specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Xpert, microscopy, culture</td>
<td>Respiratory or EP specimens</td>
</tr>
<tr>
<td>Truenat</td>
<td>Sputum (smear-positive or negative)</td>
</tr>
<tr>
<td>Moderate complexity NAATs</td>
<td>Respiratory specimens</td>
</tr>
<tr>
<td>GenoType MTBDRplus version 2</td>
<td>Sputum (smear-positive only)</td>
</tr>
<tr>
<td>Genoscholar NTM+MDRTB II</td>
<td><em>M. tuberculosis</em> isolate</td>
</tr>
<tr>
<td>Genoscholar PZA-TB II</td>
<td><em>M. tuberculosis</em> isolate</td>
</tr>
<tr>
<td>GenoType MTBDRs/ version 2</td>
<td>Sputum (smear-positive or negative)</td>
</tr>
<tr>
<td></td>
<td><em>M. tuberculosis</em> isolate</td>
</tr>
<tr>
<td>tNGS</td>
<td>Sputum (smear-positive only)</td>
</tr>
<tr>
<td></td>
<td><em>M. tuberculosis</em> isolate</td>
</tr>
<tr>
<td>WGS</td>
<td><em>M. tuberculosis</em> isolate</td>
</tr>
</tbody>
</table>

**Table 3.7 - Tests to detect specific drug resistance**
### 3.1.8 Lateral flow urine lipoarabinomannan assay

TB lipoarabinomannan (LF-LAM) is a urine-based point-of-care test that detects lipoarabinomannan (LAM) antigen, which is a marker of active TB.

This test is easy to perform by trained staff, including in peripheral health facilities.

Advantages of LF-LAM over sputum-based tests include:

- Urine specimens easier to collect.
- No risk of staff contamination during specimen collection or processing.
- No specific storage requirements for the urine prior to testing.
The urine is applied to the test strip, left at room temperature for 25 minutes, then read by the naked eye by comparing the band for positivity to a grading scale provided by the manufacturer\(^4\).

This rapid test should be used in the diagnosis of PTB and EPTB in HIV-infected children and adults. Its rapidly obtained result can contribute to reducing TB mortality among these patients\(^7\).

Its performances depend on the individual level of immunodeficiency at the time of testing. Its sensitivity is low, but it has an acceptable specificity (see below).

The LF-LAM test is recommended for the following patient groups\[^22]\:
- HIV-infected patients with signs and symptoms of TB or seriously ill, irrespective of CD4 count (sensitivity: 35%; specificity: 95%).
- Hospitalised patients with advanced HIV disease\(^9\) (sensitivity: 64%; specificity: 82%).
- HIV-infected outpatients with CD4 count < 100 cells/mm\(^3\) (sensitivity: 40%; specificity: 87%).

If LF-LAM test is positive: TB treatment should be initiated.

Due to the low sensitivity of the LF-LAM test, a negative result does not rule out TB. The test does not provide information on drug susceptibility. Therefore, all above-mentioned patients should be tested with an RMT, regardless of whether the LF-LAM result is positive or negative.

### 3.1.9 Medical imaging

**Radiography**

Chest x-ray (CXR) is used to:
- Detect abnormalities suggestive of PTB and other intra-thoracic TB localisations (pleural, pericardial, miliary).
- Evaluate the severity of intra-thoracic lesions.

It is particularly useful in the diagnosis of PTB in children (Chapter 5).

For PTB, CXR has a higher sensitivity than TB symptoms\[^23]\: a patient with a normal CXR is unlikely to have PTB. For this reason, it can also be used as a screening tool (Chapter 6) and a triaging tool to identify patients with respiratory symptoms eligible for an RMT.

CXR is also used to:
- Evaluate the response to TB treatment.
- Look for possible complications in case of worsening respiratory symptoms (pneumothorax, tracheal stenosis, etc.).

CXR has several limitations:
- Low specificity: except for cavities or miliary TB, which are specific to TB other abnormalities seen on CXR may be due to other pulmonary diseases.
- Variable quality, depending on several factors:
  - equipment and supply
  - positioning (obtaining quality CXR in children is challenging)
  - reader training and proficiency
- Difficulty distinguishing active from healed lesions
- Error rate of approximately 20%\[^24\] (specialists’ under/over-reading of the film)

When available, digital CXR has advantages over x-ray films:
- Consistent quality
- Easier image archiving
- No need for reagents and films
- Rapid transmission for teleconsultation and specialist advice
- Immediate results; possibility to screen large numbers of people within a short timeframe
- Lower radiation exposure for staff and patients.

Interpretation of digital CXR can be assisted by computer-aided detection (CAD) software packages. CAD analyses CXR for the presence of PTB-compatible abnormalities, and divides images into “normal” and “abnormal”, thereby reducing the number of CXR that need to be read by a clinician. CAD is as sensitive as a radiologist\[^25\].

Computer-aided CXR interpretation assists clinicians when all CXR cannot be read by a radiologist.

However, a radiologist should be consulted locally or via telemedicine to interpret difficult CXR (e.g. in children).
Bone x-ray is used to diagnose and evaluate severity of bone and/or joint TB and assess treatment response.

**Ultrasound**

Ultrasound (including point-of-care ultrasound, POCUS) may be useful in:
- PTB: pulmonary consolidation can support the diagnosis of PTB.
- EPTB: if suspected pleural/pericardial effusion or abdominal TB in children and adults, particularly in immunocompromised patients (e.g. HIV-infection, malnutrition).

**Table 3.8 – Medical imaging findings suggestive of TB**

<table>
<thead>
<tr>
<th>Sites</th>
<th>Findings</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Pulmonary TB</strong></td>
<td><strong>Children</strong>&lt;br&gt;See Chapter 5&lt;br&gt;&lt;br&gt;<strong>Adolescents and adults</strong>&lt;br&gt;CXR can show:</td>
</tr>
<tr>
<td></td>
<td>• Infiltrates typically located in apical and posterior segment of upper lobes and superior segments of lower lobes.</td>
</tr>
<tr>
<td></td>
<td>• Cavities (specific for TB), patchy, poorly defined consolidations.</td>
</tr>
<tr>
<td></td>
<td><strong>Patients with TB/HIV</strong>&lt;br&gt;As above.</td>
</tr>
<tr>
<td></td>
<td>• In advanced immunodeficiency, infiltrates tend to be more homogeneous, diffuse and located in the lower lungs.</td>
</tr>
<tr>
<td></td>
<td>• Less cavities than in non-HIV-infected patients.</td>
</tr>
<tr>
<td></td>
<td>• Mediastinal and hilar lymphadenopathy may be observed.</td>
</tr>
<tr>
<td></td>
<td>• Miliary pattern.</td>
</tr>
<tr>
<td><strong>Miliary TB</strong></td>
<td>CXR can show miliary nodules (1-3 mm in diameter) disseminated in both fields and uniformly distributed throughout the lung.</td>
</tr>
<tr>
<td><strong>Pleural effusion</strong></td>
<td>• CXR: effusion (even with minimal clinical signs): &lt;br&gt;</td>
</tr>
<tr>
<td></td>
<td>▪ Mostly unilateral.</td>
</tr>
<tr>
<td></td>
<td>▪ Obliteration of costophrenic angle.</td>
</tr>
<tr>
<td></td>
<td>▪ Opacity with curved upper margin.</td>
</tr>
<tr>
<td></td>
<td>▪ Ultrasound: anechogenic fluid on the costophrenic angle (may be echogenic in empyema).</td>
</tr>
<tr>
<td><strong>Pericardial effusion</strong></td>
<td>• CXR: cardiac silhouette enlargement, “water bottle” silhouette (very large effusions).</td>
</tr>
<tr>
<td></td>
<td>• Ultrasound: anechogenic fluid around the heart (may be echogenic if purulent).</td>
</tr>
<tr>
<td><strong>Bone/joint TB</strong></td>
<td>X-ray can show:</td>
</tr>
<tr>
<td></td>
<td>• Any bone/joint: osteopenia (demineralization), bone destruction with relative preservation of cartilage space.</td>
</tr>
<tr>
<td></td>
<td>• Spine: destruction of an inter-vertebral disk, osteopenia, irregularity of bone margin, bone destruction, paravertebral abscesses.</td>
</tr>
<tr>
<td><strong>Abdominal TB</strong></td>
<td>Ultrasound can show enlarged lymph nodes consistent with TB (and other diseases, especially in HIV infection), bowel wall thickening (ileo-caecal region), hypoechoic micro-abscesses of liver and/or spleen, ascites.</td>
</tr>
</tbody>
</table>

**Notes:**
- Radiographical and ultrasound findings of EPTB are non-specific. A differential diagnosis should always be considered.
- In HIV-infected patients in settings of high TB prevalence, pleural/pericardial effusion, enlarged abdominal lymph nodes, splenic microabscesses, and ascites are highly suggestive of EPTB [26].
- Adolescents typically have CXR abnormalities similar to those found in adults, however, they may also have abnormalities commonly seen in children, such as enlarged hilar lymph nodes.
3.1.10 Other laboratory tests on tissues and body fluids

The diagnosis of TB can be supported by biological tests performed on tissues or body fluids.

Table 3.9 – Findings suggestive of TB in tissues or body fluids

<table>
<thead>
<tr>
<th>Tissues/fluids</th>
<th>Findings</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lymph node</td>
<td>Cytology: granulomatous tissue, presence of giant Langhans cells, and/or caseous necrosis. AFBs are not always found by microscopy.</td>
</tr>
</tbody>
</table>
| CSF                 | - Clear, hyper-concentrated liquid.  
- High protein level > 0.40 g/l (see Pandy test, Appendix 8).  
- Low glucose < 60 mg/l.  
- Ratio CSF glucose/blood glucose < 0.5.  
- Between 100 and 1,000 white cells/mm³, of which over 80% are lymphocytes. In HIV-infected patients, rule out cryptococcal meningitis. |
| Peritoneal fluid    | Translucent, yellow-coloured liquid.  
- Exudate rich in lymphocytes, usually > 300 white cells/mm³; Rivalta test positive (Appendix 8).  
- Serum-ascites albumin gradient (SAAG):  
  < 1.1 g/dl: consistent with TB (and many other conditions).  
  > 1.1 g/dl: peritoneal TB unlikely.  
- Adenosine deaminase (ADA) > 39 U/l, likely due to TB[S7]. |
| Pleural fluid       | Straw-coloured fluid.  
- High protein level ≥ 30 g/l (Rivalta test, Appendix 8).  
- Rich in white cells (1,000-2,500/mm³), with predominant lymphocytes.  
- ADA typically > 50 U/l. Pleural effusion with an ADA < 40 U/l is much less likely due to TB. The specificity is increased when ADA is > 50 U/l and the lymphocyte-neutrophil ratio is > 0.75[S8]. |

Notes:
- ADA levels increase in TB. ADA is therefore a surrogate marker for TB in pleural and peritoneal fluids. Although not widely available, kits can be purchased to perform the test if a spectrophotometer is available.
- The sensitivity of ADA in peritoneal fluid is lower in patients with cirrhosis.
- HIV-infected patients may have lower levels of ADA.

Footnotes:
(a) When microscopy is the only diagnostic test available, specimens should be sent to a facility with capacity to perform RMTs.
(b) For more information, see: Global Laboratory Initiative. Line probe assays for drug resistant tuberculosis detection Interpretation and reporting guide for laboratory staff and clinicians. http://stoptb.org/wg/gli/assets/documents/LPA_test_web_ready.pdf
(c) For more information:
  • WHO catalogue of mutations in M. tuberculosis complex and their association with drug resistance: https://www.who.int/publications/i/item/9789240028173
(d) Microscopic observation of drug susceptibility (MODS), nitrate reductase assay (NRA), thin layer agar and colorimetric redox indicator (CRI).
(e) LAM antigen is a component of the mycobacterial cell walls released by M. tuberculosis then excreted by the kidneys.
(f) Alere Determine® TB LAM Ag (Alere Inc, Waltham, MA, USA).
(g) Seriously ill: respiratory rate > 30/minute, temperature > 39 °C, heart rate > 120/minute and unable to walk unaided.
(h) For children > 5 years and adults: CD4 count < 200 cells/mm$^3$ or a WHO clinical stage 3 or 4. All children < 5 years are considered as having advanced HIV disease.

(i) HIV-infected patients diagnosed with TB using the LF-LAM should be recorded as bacteriologically confirmed TB cases.

References


3.2 Latent tuberculosis infection

Diagnosis is based on exclusion of active TB and demonstration of latent tuberculosis infection (LTBI).

For demonstrating LTBI, one of the following tests may be performed. However, these tests are not mandatory prior to initiating LTBI treatment in:

- Children under 5 years household contact of a TB case;
- HIV-infected children and adults[1].

### 3.2.1 Tuberculin skin test

A positive tuberculin skin test (TST) indicates that a mycobacterial infection has occurred.

For interpretation of TST results, see Appendix 9.

TST has several limitations:

- It does not distinguish infection by *M. tuberculosis* from exposure to environmental mycobacteria.
- It does not distinguish latent/active TB.
- Prior BCG vaccination can result in a false positive TST.
- False negative TST is common, particularly in HIV-infected patients and malnourished children.

After having ruled out active TB, a positive TST is an indication for treatment of LTBI (Chapter 16).

**Notes:**

- TST is also used to check the absence of TB in neonates on isoniazid monotherapy (Chapter 16).
- Other skin tests are available, but have not yet been evaluated by WHO.

### 3.2.2 Interferon gamma release assays

The test is performed in vitro on blood to which *M. tuberculosis* antigens are added. This results in the rapid stimulation of memory T cells and release of interferon-gamma in patients previously exposed to the bacillus.

The following tests measure:

- QuantIFERON-TB Gold In-Tube: the amount of interferon-gamma released.
- T-SPOT.TB test: the number of interferon-gamma producing T cells[2].

The advantage of IGRAs over TST is the absence of cross-reaction with BCG vaccine and most environmental mycobacteria.

IGRAs have some limitations:

- They do not distinguish latent/active TB.
- They are more complex than TST (equipment and trained laboratory technicians) and are not widely available.

A positive test indicates that LTBI is likely; a negative test indicates that it is unlikely.

After having ruled out active TB, a positive IGRA is an indication for treatment of LTBI (Chapter 16).

**References**


### 3.3 Other investigations

In addition to TB diagnosis tests, the following investigations should be performed at baseline and during treatment. The purpose is to identify common comorbidities, contra-indications, as well as adverse effects of TB drugs.

**Table 3.10 – Other investigations in TB treatment**

<table>
<thead>
<tr>
<th>Tests</th>
<th>Indications</th>
</tr>
</thead>
<tbody>
<tr>
<td>Electrocardiogram (ECG)</td>
<td>Patients on QT prolonging drugs</td>
</tr>
<tr>
<td>Brief peripheral neuropathy screen (BPNS)</td>
<td>Patients on linezolid</td>
</tr>
<tr>
<td>Visual function tests</td>
<td>Patients on MDR/RR-TB treatment including ethambutol, linezolid or thionamides</td>
</tr>
<tr>
<td>Audiometry</td>
<td>Patients on aminoglycosides</td>
</tr>
<tr>
<td>Full blood count</td>
<td>Patients on linezolid (or rifabutin)</td>
</tr>
<tr>
<td>Liver function tests</td>
<td></td>
</tr>
<tr>
<td>Aspartate aminotransferase (AST)</td>
<td>Patients with pre-existing hepatic disease</td>
</tr>
<tr>
<td>Alanine aminotransferase (ALT)</td>
<td>Patients on MDR/RR-TB treatment</td>
</tr>
<tr>
<td>Creatinine</td>
<td>Patients with pre-existing renal disease</td>
</tr>
<tr>
<td>Serum creatinine</td>
<td>Patients on aminoglycosides</td>
</tr>
<tr>
<td>Creatinine clearance</td>
<td>Patients on aminoglycosides</td>
</tr>
<tr>
<td>Serum electrolytes (potassium)</td>
<td>Patients on aminoglycosides</td>
</tr>
<tr>
<td>Glycated haemoglobin (HbA1c), or Blood glucose level (fasting or random)</td>
<td>All patients</td>
</tr>
<tr>
<td>HIV, hepatitis B and C testing</td>
<td>Patients with undocumented HIV, hepatitis B and C status</td>
</tr>
<tr>
<td>CD4 count and viral load</td>
<td>Patients with TB/HIV coinfection</td>
</tr>
<tr>
<td>Thyroid stimulating hormone (TSH)</td>
<td>Patients on thionamides or PAS</td>
</tr>
<tr>
<td>Pregnancy test</td>
<td>Patients of childbearing age with MDR/RR-TB</td>
</tr>
</tbody>
</table>

(a) BPNS is a clinical examination for detecting peripheral neuropathy and grading the severity of symptoms ([Appendix 16](#)).

(b) For children under 5 years, a specialized equipment and consultation are required.

For more information see [Chapter 9](#), [Chapter 10](#) and [Chapter 11](#).
Chapter 4: Diagnostic algorithms for pulmonary tuberculosis (PTB) in adults and adolescents

4.1 Guiding principles for the use of the algorithms

4.2 Adult and adolescent algorithms

Update: January 2014
4.1 Guiding principles for the use of the algorithms

The aim of algorithms is to assist the diagnostic process and minimize incorrect diagnosis. The following algorithms are for adults and adolescents. For diagnostic algorithms for children < 10 years, see Chapter 5.

4.1.1 Clinical assessment

– An assessment for danger signs is the first part of the clinical assessment. The adult or adolescent is classified as seriously ill if one or more of the following danger signs are present:
  • Respiratory rate > 30/minute;
  • Fever > 39°C;
  • Pulse rate > 120/minute;
  • Unable to walk unaided.

– In cases where there is no bacteriological confirmation of TB, the clinical (and radiological) assessment should determine if the patient needs broad-spectrum antibiotics and/or anti-TB drugs.

– HIV testing should be routinely offered to all individuals suspected of having TB. If testing is refused or unavailable, it might be assumed that a certain patient is likely to be HIV-positive (according to context and/or clinical presentation). In this event, follow the algorithm for HIV-infected patients.

4.1.2 Clinical response

For patients who are treated empirically for bacterial pneumonia or pneumocystosis (PCP), a “non-response to antibiotics” increases the likelihood of TB. The converse is not necessarily true, such that a response to antibiotics does not automatically exclude TB in a person suspected of having TB, particularly if respiratory symptoms persist after treatment. Pneumonia or PCP may occur in patients with underlying TB.

Antibiotic treatment is appropriate for HIV-infected patients with cough because bacterial infections are common both with and without TB. All seriously ill patients being started on anti-TB treatment should also be treated empirically, with broad-spectrum antibiotics for bacterial pneumonia because benefits outweigh the risks.

References

4.2 Adult and adolescent algorithms

Diagnostic algorithm 1

PTB in HIV-negative patients with low risk of MDR-TB

---

a. When the patient's serological status is unknown, this algorithm should be used in settings with HIV prevalence < 5%.

b. Patients are considered to be at low risk of multidrug-resistant TB (MDR-TB) if they do not meet one of the following criteria: 1) resident in areas with high MDR-TB prevalence; 2) all retreatment categories; 3) exposure to a known MDR-TB case; 4) patient remaining smear + at 2 months; 5) exposure to institutions with high risk of MDR-TB (e.g. prisons).

c. Danger signs: respiratory rate > 30/min and/or fever > 39°C and/or pulse rate > 120/min and/or unable to walk.

d. Smear microscopy: two sputum examinations performed on the same day.

e. Broad spectrum ATB:
• If no danger signs: amoxicillin for 7 days (NO fluoroquinolones);
• If danger signs: parenteral ATB (e.g. ceftriaxone).

Clinical response to a broad spectrum antibiotic does not rule out TB. Patient should be informed to return for reassessment if symptoms recur.

According to setting:
• Xpert MTB/RIF available: two sputum smear microscopy on the same day and one Xpert MTB/RIF from one of the samples collected for smear microscopy;
• Xpert MTB/RIF not available: two sputum smear microscopy on the same day.

In groups of patients with high level of resistance to isoniazid (> 10%) it is recommended to perform a conventional DST at baseline (and/or a line probe assay) in order to provide adequate treatment.

According to setting:
• In groups of patients with prevalence of MDR-TB < 10%, patients seriously ill should immediately be initiated under empiric MDR-TB treatment. H and R will be included in the regimen until confirmation of MDR-TB by conventional methods. If the patient is stable, the clinician may choose to wait for confirmation before initiating a MDR treatment.
• In groups of patients with prevalence of MDR-TB ≥ 10%, patients should be initiated under empiric MDR-TB treatment. Consider adding H in settings where mono-resistance to R is not uncommon.

Clinical signs and chest X-ray (CXR) findings tend to be more typical in those who are HIV-negative having active TB:

<table>
<thead>
<tr>
<th></th>
<th>TB</th>
<th>Bacterial pneumonia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clinical signs</td>
<td>Weight loss, productive cough, purulent sputum, haemoptysis, pleuritic chest pain</td>
<td>• Acute onset • Fever</td>
</tr>
<tr>
<td>CXR</td>
<td>• Infiltrates, nodules with or without cavitation in the upper lobes and in the superior segments of the lower lobes. • Pleural effusions • Adenopathy in the mediastinum or hila (rare in TB in adults and adolescents) • Miliary disease</td>
<td>• Lobar consolidation</td>
</tr>
</tbody>
</table>

When clinical signs AND CXR are strongly suggestive of active TB, treatment should be initiated without waiting for diagnosis confirmation.

Diagnostic algorithm 2

PTB in HIV-positive patients
When the patient’s serological status is unknown, this algorithm should be used in settings with HIV prevalence > 5%.

TB suspect is defined as: cough for more than 2 weeks or any cough with at least one of the following signs: loss of weight, night sweats, fever, and suspicion based on clinical judgment.

Danger signs: respiratory rate > 30/min and/or fever > 39°C and/or pulse rate > 120/min and/or unable to walk.

According to setting:
- Xpert MTB/RIF available: two sputum smear microscopy on the same day AND one Xpert MTB/RIF from one of the samples collected for smear microscopy;
- Xpert MTB/RIF not available: two sputum smear microscopy on the same day.

In patients groups with high level of resistance to isoniazid (> 10%) it is recommended to perform a conventional DST at baseline (and/or a line probe assay) in order to provide adequate treatment.

When possible a culture should be performed. A positive culture result at any point in time in the algorithm should lead to a full TB treatment.
TB treatment should be started when clinical signs AND chest X-ray (CXR) are suggestive of TB (Note k).

**Broad spectrum ATB/PCP:**
- If no danger signs: amoxicillin for 7 days (or recommended oral agent for community-acquired pneumonia in the area). Do NOT use fluoroquinolones;
- If danger signs: parenteral ATB (e.g. ceftriaxone) AND high dose cotrimoxazole.

If no danger signs: patient should be re-assessed after 7 days.
If danger signs: patient should be assessed daily and if no response, TB treatment should be considered after 3 to 5 days.
Clinical response to broad-spectrum ATB does not rule out TB. Patient should be informed to return for reassessment if symptoms recur.

**Differential diagnosis of a coughing HIV-infected adult/adolescent:** bacterial (including atypical) pneumonia, PCP, fungal infection, non-tuberculous mycobacteria, nocardiosis, Kaposi sarcoma and lymphoma.

The diagnosis should be based on clinical assessment, CXR and CD4 results, whether cotrimoxazole preventive therapy (CPT) was used, and other treatment already used in the patient. If the index of suspicion for active TB is high, empiric TB treatment should be initiated without waiting for diagnosis confirmation. Other treatments such as broad-spectrum ATB or therapy for PCP may be needed in addition to TB treatment.

<table>
<thead>
<tr>
<th>Clinical signs</th>
<th>TB</th>
<th>PCP (HIV+)</th>
<th>Bacterial pneumonia</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Clinical signs</strong></td>
<td><strong>Dry cough</strong></td>
<td><strong>Acute onset</strong></td>
<td></td>
</tr>
<tr>
<td>Clinical signs</td>
<td><strong>Weight loss</strong></td>
<td><strong>High fever</strong></td>
<td></td>
</tr>
<tr>
<td>signs</td>
<td><strong>Purulent sputum and haemoptysis less likely if HIV-positive with low CD4 count</strong></td>
<td><strong>Dyspnoea ++</strong></td>
<td></td>
</tr>
<tr>
<td></td>
<td><strong>Fever</strong></td>
<td><strong>Hypoxemia</strong></td>
<td></td>
</tr>
<tr>
<td></td>
<td><strong>Night sweats</strong></td>
<td><strong>Not on CPT</strong></td>
<td></td>
</tr>
<tr>
<td></td>
<td><strong>Pleuritic chest pain</strong></td>
<td><strong>More likely if low CD4 count</strong></td>
<td></td>
</tr>
<tr>
<td><strong>CXR</strong></td>
<td><strong>Upper lobe infiltrates and cavitation only likely in HIV-positive adults with higher CD4 counts. Any lobe of the lung may be affected</strong></td>
<td><strong>Bilateral interstitial infiltrate with reticulonodular markings that are more pronounced in the lower lobes</strong></td>
<td><strong>Lobar consolidation</strong></td>
</tr>
<tr>
<td></td>
<td><strong>In HIV-positive adults with lower CD4 counts, the following 4 patterns are suggestive of TB:</strong></td>
<td><strong>Findings lag behind symptoms and may be normal early in the disease</strong></td>
<td></td>
</tr>
<tr>
<td></td>
<td>1. miliary pattern</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>2. pleural effusion without airspace (with straw-coloured liquid aspirate)</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>3. hilar and mediastinal adenopathy</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>4. large heart (especially if symmetrical and rounded)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Diagnostic algorithm 3 with Xpert MTB/RIF**

**PTB in patients with high risk of MDR-TB**

In the absence of any improvement of clinical signs (no weight gain, persistent cough, pain, etc.) AND no improvement on CXR after 2 months of a well conducted TB treatment, diagnosis and treatment should be reconsidered. MDR-TB should also be considered.

Immediately start empiric MDR treatment, even if positive predictive value of Xpert MTB/RIF for R resistance is low (this is done to avoid the rapid and high mortality due to untreated MDR-TB in HIV patients). H and R should be included in the regimen until confirmation of MDR-TB by conventional methods if the patient comes from a group with less than a prevalence of MDR-TB < 10%. In groups of patients with prevalence of MDR-TB ≥ 10%, patients should be initiated under an empiric MDR treatment without H or R, although one can consider adding H in settings where mono-resistance to R is not uncommon.
The following patients are considered to be at high risk of MDR-TB: 1) resident in areas with high MDR-TB prevalence; 2) all retreatment categories; 3) exposure to a known MDR-TB case; 4) patient remaining smear-positive at 2 months; 5) exposure to institutions with high risk of MDR-TB (e.g. prisons).

Groups of patients at risk of MDR-TB are also at risk of other types of DR-TB as well. DST to the first-line should be performed in order to provide adequate treatment for possible mono- or poly-drug resistance.

In populations with a prevalence < 10% of MDR-TB, the resistance to R diagnosed by Xpert MTB/RIF must be confirmed by conventional methods. Drug sensitivity testing (DST) to both first-line drugs and second-line TB drugs should be performed if possible.

In groups of patients with prevalence of MDR-TB < 10%, the decision to start the MDR-TB treatment will be made on clinical presentation of the patient and immunological status. Patients seriously ill and/or HIV+ should be initiated immediately under empiric MDR-TB treatment. H and R will be included in the regimen until confirmation by conventional methods.

In groups of patients with prevalence of MDR-TB ≥ 10%, the patient should be initiated using an empiric MDR-TB treatment. Consider adding H in settings where mono-resistance to R is not uncommon.

Baseline sputum smear microscopy result on 1 specimen in order to: 1) allow patient follow-up with microscopy; 2) take immediate decisions related to TB infection control.
Chapter 5: Diagnosis of active tuberculosis in children

5.1 Introduction
5.2 Diagnostic approach
5.3 Paediatric diagnostic algorithms

Update: March 2023
5.1 Introduction

In children, defined in this chapter as patients under 10 years, tuberculosis (TB), pulmonary (PTB) and extrapulmonary (EPTB), is a significant cause of morbidity and mortality.

Globally, WHO estimates that more than one million children develop active TB every year[1] and that 60% of TB cases in children are not diagnosed or not reported[2].

After exposure, the risk of TB infection and progression to active TB is high in children under 5 years[3]. Progression to active TB is rapid (within 12 months) in children under 2 years[4].

HIV infection is a significant risk factor for developing TB in children under 1 year[5].

The risk of miliary TB and EPTB, including severe forms such as TB meningitis, is higher in children under 5 years and in immunocompromised children[6]. The most common forms of EPTB are lymph node TB and TB pleural effusion. Osteoarticular TB represents 1 to 2% of TB in children[4].

The risk of death from TB is higher in children under 2 years and children with HIV infection or severe acute malnutrition (SAM)[3]. Almost all deaths due to TB in children occur in those not receiving TB treatment, and in the vast majority of cases, in children under 5 years[8].

TB treatment should not be delayed if investigations, or results of investigations, are not immediately available in children at high risk of TB or death from TB.

Children often have the same resistance profile as the index case, i.e. the person who is the presumed source of the infection. If the resistance profile is not available for the child, the resistance profile of the index case should be taken into account for the child's TB treatment.

Children are not considered infectious unless they have extensive lung involvement and/or cavitory PTB or positive smear microscopy.

References

5.2 Diagnostic approach

Children with TB usually have non-specific symptoms. Clinicians should therefore look for TB, particularly in children:

- Under 2 years of age, or
- With HIV infection or SAM, or
- In contact with a person with TB, or
- Not responding to antibacterial and/or nutritional treatment.

The diagnosis of TB in children, particularly those under 5 years, is often based on a combination of history of exposure to a person with TB, clinical assessment and investigations, such as radiology, when available.

In children at high risk of death from TB, treatment should be initiated as soon a TB diagnosis is considered likely. In children not at high risk of death from TB, the diagnosis may not be made at the first consultation. A second consultation after one to two weeks is often necessary to reassess the clinical status.

The diagnosis is often made without bacteriological confirmation as:

- Children under 5 years have low bacillary load and bacteriological tests are often negative.
- Specimens for diagnosis of EPTB may be difficult to collect.

TB is bacteriologically confirmed in only 20 to 30% of children[1].

To facilitate the diagnosis of PTB and enable rapid treatment in children, WHO has developed diagnostic algorithms (Section 5.3). The diagnosis of EPTB uses the same diagnostic approach. However, no evidence-based algorithms are currently available.

A trial of treatment with TB drugs is not recommended as a method to diagnose TB. Once a decision is made to treat TB in a child, a full course of treatment should be given.

5.2.1 History of exposure to tuberculosis

Children are at risk of TB if they are exposed to a person with confirmed or presumed TB.

They are at higher risk of TB if:
- The index case is a household or close contact.
- The index case has PTB, sputum smear-positive or cavities on chest x-ray.
- The exposure to the index case has occurred in the past 12 months.

Note: Conversely, when TB is diagnosed in children, it is important to detect the index case and any other undiagnosed household member(s) or close contact(s).

5.2.2 Clinical assessment

Symptoms suggestive of tuberculosis

Ask if the child has symptoms commonly associated with TB:

- Cough for more than 2 weeks.
- Fever for more than 2 weeks.
- Night sweats that soak the bed or clothes.
- Weight loss or poor/no weight gain.
- Fatigue, reduced playfulness, loss of appetite.
- Haemoptysis (rare in children).
- Non-painful, enlarged cervical, submandibular, or axillary lymph nodes.
- Rapid breathing.

Physical examination and growth assessment

Look for signs suggestive of TB:

- Fever, tachypnoea, tachycardia.
- Weight loss, growth curve flattening, underweight or malnourished according to weight for height and/or mid-upper arm circumference.
• Abnormal pulmonary auscultation.
• Signs of respiratory distress and SpO₂ < 90-92%.
• Lethargy, altered mental status (may indicate TB meningitis).
• Signs of EPTB:
  ▪ Highly suggestive, e.g.:
    ▪ Angular deformity of the spine, loss of ability to walk.
    ▪ Cervical lymph node with fistula formation.
  ▪ Requiring further investigation, e.g.:
    ▪ Sub-acute meningitis not responding to antibiotic treatment.
    ▪ Ascites.
    ▪ Lymph node without fistula formation.
    ▪ Non-painful enlarged joint.

HIV status should be assessed in all children with presumed or confirmed TB.

Clinical review

If diagnosis is not made at the first consultation, reassess the child (signs/symptoms suggestive of TB and growth) within one to two weeks maximum.

The following are suggestive of TB:
• Persistent or worsening pneumonia despite non-TB antibiotic treatment.
• No weight gain or weight loss despite nutritional support or treatment.
• Persistent fever after other causes have been ruled out or treated (e.g. malaria).
• Persistent or worsening fatigue, reduced playfulness, loss of appetite.

5.2.3 Baseline investigations

When PTB or EPTB is suspected, perform bacteriological tests, lateral flow urine lipoarabinomannan assay if indicated, and radiography if available.

Bacteriological tests

Rapid molecular tests (RMTs) should be performed on respiratory, stool or extrapulmonary (EP) specimens as the initial diagnostic test.

As the sensitivity of Xpert MTB/RIF Ultra is higher than that of Xpert MTB/RIF, preferably use MTB/RIF Ultra for the detection of TB and rifampicin-resistance (Chapter 3).

Sputum specimens can be difficult to obtain in children. Explanation and encouragement are important. Chest clapping may help expectoration.

If sputum cannot be obtained spontaneously, more invasive procedures, such as nasopharyngeal aspiration, sputum induction or gastric aspiration (Appendix 3), can be performed, but only if the specimen is collected for rapid molecular tests, culture or genome sequencing. These procedures should not be performed for smear microscopy.

Stool specimens (which may contain swallowed sputum) are an alternative to respiratory specimens for the diagnosis of PTB in children. Respiratory specimens are more likely to give a positive result, but the use of stool specimens can avoid invasive collection procedures.

For children at risk of DR-TB, i.e. contact with a person with DR-TB or coming from an area with high DR-TB prevalence:
• Multiple specimens (respiratory, stool and EP) should be tested with RMTs. Multiple testing increases the likelihood of detecting TB and obtaining the resistance profile.
• Every effort should be made to perform culture and phenotypic drug susceptibility tests (Chapter 3).

For the diagnostic accuracy of Xpert MTB/RIF in specimens other than sputum, see Appendix 1.

Lateral flow urine lipoarabinomannan assay (LF-LAM)

LF-LAM should be performed in HIV-infected children:
• With signs and symptoms of TB, or
• Hospitalised with advanced HIV disease, or
• Followed as outpatients with a low CD4 count.
Chest x-ray (CXR)

CXR is particularly useful when bacteriological tests are negative or not available. It is also useful to assess the severity of TB and to determine eligibility for the 4-month drug-susceptible TB regimen.

Children with PTB usually have abnormalities on CXR, but a normal CXR does not rule out TB.

For young children unable to stand alone, perform anteroposterior and lateral CXRs if possible (lateral CXR can improve detection of enlarged hilar/mediastinal lymph nodes).

For other children, perform a standard posteroanterior CXR.

CXR findings suggestive of TB in children include*: enlarged hilar/mediastinal lymph nodes, miliary pattern, and cavities. Although generally less specific, consolidation and pleural/pericardial effusion in a child not acutely ill is also suggestive of TB.

Ultrasound

See Chapter 3.

Tuberculin skin test (TST)

In children, a positive TST may be one element among many to establish the diagnosis of active TB. However, it has many limitations (see Chapter 3 and Appendix 9).

5.2.4 Follow-up investigations

For children able to expectorate spontaneously, smear microscopy is used to monitor treatment progress (Chapter 3).

For children unable to expectorate spontaneously, monitoring of treatment progress is clinical. Invasive procedures should not be performed to obtain respiratory specimens for smear microscopy.

Footnotes


References


https://doi.org/10.5588/ijtld.15.0471
5.3 Paediatric diagnostic algorithms

5.3.1 Diagnosis of PTB in symptomatic children with CXR

5.3.2 Diagnosis of PTB in symptomatic children without CXR

(a) Danger signs:
Children ≤ 5 years: age ≤ 2 months; unable to eat or drink; vomiting up everything; severe dehydration; severe pallor; stridor; SpO₂ < 90%; respiratory distress; seizure; profound lethargy or coma; restless, continuously irritable; neck stiffness or bulging fontanelle; fever > 39 °C; SAM. Children ≥ 3 years: classifiable with fever, dehydration; severe pallor; high (cold extremities, capillary refill time > 3 seconds, weak and fast pulse, obstructed or absent breathing; respiratory distress; central cyanosis; coma (or seriously altered level of consciousness); seizures; restless, continuously irritable; fever > 39 °C; SAM.
(b) SAM: Severe acute malnutrition is defined as weight-for-height in 2-score less than -3 or mid-upper arm circumference less than 111 mm.
Diagnosis of PTB in symptomatic children without CXR

1. **Danger signs** requiring urgent treatment:
   - Age < 2 years or HIV infection or SAM

2. **Stabilise and/or transfer**
   - Yes
   - **Age < 2 years or HIV infection or SAM?**
     - Yes
     - Treat most likely non-TB conditions: Repeat clinical history and physical examination in 1-2 weeks.
     - **Persistent or worsening symptoms?**
       - Yes
       - **M. tuberculosis detected by Xpert or LAM?**
         - Yes
         - **Household/close contact with a TB case in the last 12 months?**
           - Yes
           - Score clinical features
             - Cough > 2 weeks
             - Fever > 2 weeks
             - Leukopoenia
             - Night sweats
             - Weight loss
             - Loss of appetite
             - **Tuberculosis**
             - Start TB treatment(s)
           - Score > 107
         - No
         - Do not start TB treatment; Repeat clinical history, physical examination, and scoring in 1-2 weeks
       - No
       - **Exit**
     - No
     - Exit
   - No
   - **Exit**

3. **Persistent or worsening symptoms?**
   - Yes
   - **M. tuberculosis detected by Xpert or LAM?**
     - Yes
     - **Household/close contact with a TB case in the last 12 months?**
       - Yes
       - Score clinical features
         - Cough > 2 weeks
         - Fever > 2 weeks
         - Leukopoenia
         - Night sweats
         - Weight loss
         - Loss of appetite
         - **Tuberculosis**
         - Start TB treatment(s)
       - Score > 107
     - No
     - **Exit**
   - No
   - **Exit**

4. **Stabilise and/or transfer**
   - Yes
   - **Age < 2 years or HIV infection or SAM?**
     - Yes
     - Treat most likely non-TB conditions: Repeat clinical history and physical examination in 1-2 weeks.
     - **Persistent or worsening symptoms?**
       - Yes
       - **M. tuberculosis detected by Xpert or LAM?**
         - Yes
         - **Household/close contact with a TB case in the last 12 months?**
           - Yes
           - Score clinical features
             - Cough > 2 weeks
             - Fever > 2 weeks
             - Leukopoenia
             - Night sweats
             - Weight loss
             - Loss of appetite
             - **Tuberculosis**
             - Start TB treatment(s)
           - Score > 107
         - No
         - Do not start TB treatment; Repeat clinical history, physical examination, and scoring in 1-2 weeks
       - No
       - **Exit**
     - No
     - Exit
   - No
   - **Exit**

---

(a) **Danger signs**:
- Children < 5 years age < 2 months: unable to eat or drink; vomiting up everything; severe dehydration; severe pallor; urin: SPO2 < 90%; respiratory distress; seizures; profound lethargy or coma; restless, continuously irritable, with drowsiness and increasing lassitude; fever > 38 °C; SAM
- Children 2.5 years: diarrhoea with severe dehydration; severe pallor; shock (cold extremities, capillary refill time > 3 seconds, weak and tachypnoe); obstructed or absent breathing; respiratory distress; central cyanosis; coma (seriously altered level of consciousness) seizures; restless, continuously irritable; fever > 38 °C; SAM.

(b) **SAM**: severe acute malnutrition is defined as weight-for-height in Z-score less than -3 or mid-upper arm circumference less than 215 mm.

(c) Once a decision to treat for TB is made, every effort should be made to obtain a CXR to assess the severity of TB.
Chapter 6: Intensive case finding in HIV-infected individuals

6.1 Routine screening
6.2 Purposes of screening

Update: January 2014
6.1 Routine screening

Intensive case-finding (ICF) should be in place in all projects providing care to HIV-infected patients. Screening can be performed at multiple points in time by different levels of health care workers (e.g. counsellors during HIV testing, health care providers during clinical consultations).

All children and adults should be regularly screened for TB using the following criteria:

Table 6.1 - Screening criteria/symptoms in children and adults¹

<table>
<thead>
<tr>
<th>Children</th>
<th>Adults</th>
</tr>
</thead>
<tbody>
<tr>
<td>• Current cough (a)</td>
<td>• Current cough (a)</td>
</tr>
<tr>
<td>• Fever</td>
<td>• Fever</td>
</tr>
<tr>
<td>• Poor weight gain (b)</td>
<td>• Weight loss</td>
</tr>
<tr>
<td>• Contact with a contagious person</td>
<td>• Night sweats</td>
</tr>
</tbody>
</table>

(a) Asking about “current cough”, rather than cough for 2 weeks, is more sensitive for TB disease in HIV-infected individuals
(b) Poor weight gain is defined as reported weight loss or underweight or confirmed weight loss > 5% since last visit, or growth curve flattening.

References

6.2 Purposes of screening

6.2.1 Early detection and treatment of active TB

Children and adults found to have one or more of the above symptoms/criteria during screening may have active TB, and they should be evaluated with an appropriate TB diagnostic algorithm in order to rapidly diagnose those who do have TB (see Diagnostic algorithms, Chapter 4 and Chapter 5).

6.2.2 Identification of patients eligible for isoniazid preventive therapy (IPT)

The significant proportion of asymptomatic active TB described in patients eligible for ART (15 to 20%)[1][2] leads to use this screening method with caution. It should only be used for ruling out TB in patients not yet eligible for antiretroviral therapy (CD4 greater than 350 and no WHO stage 3 or 4 illnesses) or after three months of treatment in patients started under antiretroviral therapy in order to allow for possible unmasking of TB.

For IPT, see Chapter 16.

References


Chapter 7: Case definitions for registration

7.1 Definition of a tuberculosis case
7.2 History of prior anti-TB treatment
7.3 Anatomical site of the disease
7.4 Bacteriological status
7.5 HIV status
7.6 Other co-morbidities
7.7 Summary of patient registration

Update: January 2014
7.1 Definition of a tuberculosis case

A tuberculosis (TB) case is a patient that has been diagnosed as such by a clinician, regardless if the diagnosis has been confirmed bacteriologically or not.

The elements necessary for defining a TB case are: the TB treatment history, the bacteriological status, the anatomical site of the disease and the patient’s HIV status.

Note: any person receiving treatment for TB should be recorded as a TB case.
7.2 History of prior anti-TB treatment

Patients who have interrupted or failed a previous anti-TB treatment have a higher risk of developing drug-resistance (DR). Therefore, it is important to question patients about their previous treatment prior to treatment initiation.

Case registration distinguishes between[1]:

- **New patients**: patients who have never been treated for TB or have taken anti-TB drugs for less than 1 month.

- **Previously treated patients**: patients who have received 1 month or more of anti-TB drugs in the past. Previously treated patients are further sub-classified into relapse, failure and return after treatment interruption:
  - **Relapse**: patients who were cured or completed treatment on their last TB treatment;
  - **Failure**: patients who have failed their most recent treatment (see Chapter 17 for outcome definitions for failure);
  - **Treatment interruption**: patients who interrupted (see Chapter 17 for outcome definition of treatment interruption) their last treatment should be classified as “Return after treatment interruption”.

- **Others**: patients who cannot be included in one of the above categories (e.g. patients who have previously been treated via an erratic or unknown TB regimen).

References

7.3 Anatomical site of the disease

**Pulmonary TB (PTB)**

Refers to a case of TB presenting with involvement of the lung parenchyma.

**Notes:**
- Miliary TB is also classified as PTB because there are lesions in the lungs.
- Any patient presenting with PTB and an EPTB form at the same time is classified as a PTB case for recording purposes.

**Extrapulmonary TB (EPTB)**

Refers to a case of TB involving organs other than the lungs. Diagnosis is based on clinical signs corresponding to extrapulmonary active TB and a decision by a clinician to treat with a full course of anti-TB drugs.

**Notes:**
- Sputum smear microscopy should always be done, and if possible culture and/or molecular test.
- Patients presenting with tuberculous pleural effusion, or mediastinal lymphadenopathy without evidence of parenchymal localization are classified in this category.

**Footnotes**

(a) If possible, obtain histological or bacteriological evidence (microscopy, culture or molecular test).
7.4 Bacteriological status

Bacteriological status refers to the detection of *M. tuberculosis* by smear, culture or molecular methods. The bacteriological status can be further sub-classified on the basis of drug sensitive and drug resistant cases.

7.4.1 Detection of *M. tuberculosis*

Every TB case should be classified into one of two categories:

- **Confirmed TB case**: a case with a positive bacteriological result (microscopy, culture or molecular method).
- **Non-confirmed TB case**: a case where investigations are negative (microscopy, culture or molecular method) and for whom a clinician prescribes anti-TB treatment.

Confirmed TB cases are further sub-classified as:
1. smear positive/negative/not done
2. culture positive/negative/not done
3. molecular test positive/negative/not done

7.4.2 Strain sensitivity/resistance

When possible, culture and DST should be done to determine if the strain presents resistance to some drugs:

- **Susceptible TB**: the strain is not resistant to any first-line anti-TB drugs.
- **Drug-resistant TB**:
  - Monodrug-resistant TB: resistance to one first-line anti-TB drug only;
  - Polydrug-resistant TB (PDR-TB): resistance to more than one first-line anti-TB drug, other than isoniazid and rifampicin;
  - Multidrug-resistant TB (MDR-TB): resistance to at least isoniazid and rifampicin;
  - Extensively drug-resistant TB (XDR-TB): MDR-TB resistant to at least one fluoroquinolone and at least one second-line injectable drug (Km, Amk, Cm).

Patients with DR-TB should be classified in the following manner:

- **Confirmed isoniazid resistance and rifampicin susceptible**: resistance to isoniazid but not rifampicin. Resistance to first and second-line anti-TB drugs may be present.
- **Confirmed rifampicin resistant TB** (RR-TB): resistance to rifampicin confirmed by phenotypic drug susceptibility test or line probe assay or Xpert MTB/RIF (isoniazid susceptible or unknown).
- **Confirmed MDR-TB**: resistance to isoniazid and rifampicin, with or without resistance to first and second-line anti-TB drugs.
- **Confirmed XDR-TB**: resistance to isoniazid and rifampicin, and to at least one fluoroquinolone, and one second-line injectable drug (Km, Amk, Cm).
- **Unconfirmed DR-TB**: patients treated as DR-TB but without DST results (e.g. children who are contacts of a known case, patients with clinical failure and for whom no DST was available for some reason).
7.5 HIV status

Determining and recording the patient's HIV status is critical for treatment decisions, as well as for assessing programme performances. The TB treatment card and TB register, which should be treated as confidential documents, should include: dates and results of HIV tests, starting date of cotrimoxazole and antiretroviral therapy.
7.6 Other co-morbidities

Any other significant diseases, such as diabetes, hepatitis B or C, cancer and malnutrition, should be noted at registration.
# 7.7 Summary of patient registration

Table 7.1 summarizes the elements necessary for defining a TB case.

**Table 7.1 - Patient registration by outcome of most recent TB treatment**

<table>
<thead>
<tr>
<th>Registration groups based on treatment history</th>
<th>Further classification</th>
</tr>
</thead>
<tbody>
<tr>
<td>New</td>
<td></td>
</tr>
<tr>
<td>Previously treated</td>
<td></td>
</tr>
<tr>
<td>Previously treated Relapse</td>
<td>1. <strong>PTB or EPTB?</strong> If EPTB, indicate site.</td>
</tr>
<tr>
<td>Failed Treatment interruption</td>
<td>2. <strong>Bacteriologically confirmed or non-confirmed TB case?</strong></td>
</tr>
<tr>
<td>Other</td>
<td>3. <strong>Sub-category of bacteriological status:</strong></td>
</tr>
<tr>
<td></td>
<td>▪ Smear positive/negative/not done</td>
</tr>
<tr>
<td></td>
<td>▪ Culture positive/negative/not done</td>
</tr>
<tr>
<td></td>
<td>▪ Molecular test positive/negative/not done</td>
</tr>
<tr>
<td></td>
<td>4. <strong>If previously treated:</strong></td>
</tr>
<tr>
<td></td>
<td>▪ Document last regimen received</td>
</tr>
<tr>
<td></td>
<td>▪ History of second-line drug use</td>
</tr>
<tr>
<td></td>
<td>5. <strong>DST pattern:</strong> susceptible to H and R, confirmed H resistance and R susceptible, RR-TB, MDR-TB or XDR-TB</td>
</tr>
<tr>
<td></td>
<td>6. <strong>HIV status</strong> (negative/positive/not done)</td>
</tr>
<tr>
<td></td>
<td>7. <strong>Other co-morbidities?</strong></td>
</tr>
</tbody>
</table>

## References

Chapter 8: Tuberculosis drugs and treatment regimens

8.1 Introduction
8.2 Standard code for treatment regimens
8.3 Drugs for drug-susceptible tuberculosis
8.4 Drugs for drug-resistant tuberculosis
8.5 Tuberculosis drug formulations

Update: October 2022
8.1 Introduction

A combination of several antituberculosis drugs is needed to treat tuberculosis (TB) and prevent the emergence of resistance. Each TB drug has a specific action on one or more bacillary populations, but none on dormant bacilli.

TB drugs are classified into two categories:
- Drugs for drug-susceptible TB (DS-TB), also referred to as “first-line TB drugs”.
- Drugs for drug-resistant TB (DR-TB), also referred to as “second-line TB drugs”. WHO has further classified DR-TB drugs in groups based on their effectiveness and safety profile.

Treatment regimens define the TB drug combinations used and the intended duration of TB treatment.

For more information on the TB drugs, see Appendix 10.
8.2 Standard code for treatment regimens

8.2.1 Tuberculosis drugs

Each TB drug has an abbreviation.

Table 8.1 - Categories and abbreviations of TB drugs

<table>
<thead>
<tr>
<th>Categories</th>
<th>TB drugs</th>
<th>Abbreviations</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Drug-susceptible TB</strong> (first-line drugs)</td>
<td>Isoniazid (standard dose)</td>
<td>H R Z E Rfb P</td>
</tr>
<tr>
<td></td>
<td>Rifampicin</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Pyrazinamide</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Ethambutol</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Rifabutin</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Rifapentine</td>
<td></td>
</tr>
<tr>
<td><strong>Drug-resistant TB</strong> (second-line drugs)</td>
<td>Levofoxacin or moxifloxacin</td>
<td>Lfx or Mfx Bdq Lzd</td>
</tr>
<tr>
<td>Group A</td>
<td>Bedaquiline</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Linezolid</td>
<td></td>
</tr>
<tr>
<td>Group B</td>
<td>Clofazimine</td>
<td>Cfz Cs or Trd</td>
</tr>
<tr>
<td></td>
<td>Cycloserine or terizidone</td>
<td></td>
</tr>
<tr>
<td>Group C</td>
<td>Delamanid</td>
<td>Dlm E Z</td>
</tr>
<tr>
<td></td>
<td>Ethambutol</td>
<td>Ipm/Cln or Mpm Am or S</td>
</tr>
<tr>
<td></td>
<td>Pyrazinamide</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Imipenem/cilastatin or meropenem</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amikacin or streptomycin</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Ethionamide or prothionamide</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Para-aminosalicylate sodium or para-aminosalicylic acid</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Isoniazid (high-dose)</td>
<td></td>
</tr>
<tr>
<td>Ungrouped</td>
<td>Pretomanid</td>
<td>Pa</td>
</tr>
</tbody>
</table>

Notes:
- High-dose isoniaizd, although not a Group C drug according to the WHO classification, is considered in this guide as a Group C drug as it is used as such when building a treatment regimen for DR-TB.
- Pretomanid is not categorized in the WHO classification and is only used in standard treatment regimens for DR-TB (Chapter 10).

8.2.2 Treatment regimens

TB treatment regimens are expressed as follows:
- Drugs are designated by their abbreviation.
- For some regimens, the treatment is divided into two phases: initial (or intensive) phase, and continuation phase. The phases are separated by a slash /.
- Letters in brackets ( ) indicate fixed-dose combinations (FDCs).
- Letters that are not in brackets indicate individual drugs.
- Second-line drugs are separated by a hyphen.
• Letters in square brackets [ ] indicate that drugs are used, but not considered as likely effective (Chapter 10).
• A superscript h (°) indicates that the drug is administered in a high dose.
• Numbers before letters indicate the duration (in months) of the treatment or of each phase.
• Numbers in subscript and angle brackets < > after a drug indicate the duration (in months) of the treatment with this drug.

Box 8.1 – Examples

2(HRZE)/4(HR): the initial phase lasts 2 months with an FDC containing 4 drugs; the continuation phase lasts 4 months, with an FDC containing 2 drugs.

18Bdq-Lzd-Cfz-Cs-Dlm-[Z]: the treatment lasts 18 months with 6 individual drugs; Z is used, but not considered as a likely effective drug.

4Bdq°6°-Lfx-Cfz-Z-E-H°Lzd°2°/5Lfx-Cfz-Z-E: the initial phase lasts 4 months but bedaquiline is given for 6 months and linezolid for 2 months only; the continuation phase lasts 5 months.
8.3 Drugs for drug-susceptible tuberculosis

All drugs used for DS-TB treatment are taken 7 days a week.

8.3.1 First-line drugs

Table 8.2 – Main characteristics of first-line TB drugs

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Activity</th>
<th>Resistance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Isoniazid</td>
<td>Bactericidal</td>
<td>• High level of resistance in some regions.</td>
</tr>
<tr>
<td></td>
<td></td>
<td>• Cross-resistance with thionamides.</td>
</tr>
<tr>
<td>Rifampicin</td>
<td>Bactericidal</td>
<td>• High level of resistance to rifampicin in some regions.</td>
</tr>
<tr>
<td>Rifabutin</td>
<td>Bactericidal</td>
<td>• High level of cross-resistance between rifamycins.</td>
</tr>
<tr>
<td>Rifapentine</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ethambutol</td>
<td>Bacteriostatic</td>
<td>Unknown (no reliable drug susceptibility test for ethambutol).</td>
</tr>
<tr>
<td>Pyrazinamide</td>
<td>Weakly bactericidal</td>
<td>High level of resistance in regions where rifampicin resistance is frequent.</td>
</tr>
</tbody>
</table>

**Isoniazid**

Isoniazid is usually well tolerated at recommended doses. It may cause peripheral neuropathy, hepatotoxicity, and hypersensitivity reactions. Peripheral neuropathy can be prevented by administration of pyridoxine (vitamin B<sub>6</sub>). See Appendix 17.

**Rifamycins (rifampicin, rifabutin, rifapentine)**

Rifamycins are usually well tolerated at recommended doses. They may cause hypersensitivity reactions, hepatotoxicity, and thrombocytopenia. They are strong inducers of cytochrome P450 and can affect the plasma concentrations of many drugs (Appendix 19). Rifampicin is the most used rifamycin in the treatment of DS-TB. Rifabutin is used instead of rifampicin in patients taking certain antiretrovirals (Appendix 19). Rifapentine is only used in the 4-month regimen 2HPZ-Mfx/2HP-Mfx.

**Note:** rifampicin and rifapentine are also used to treat latent TB infection (Chapter 16).

**Ethambutol**

Ethambutol is usually well tolerated, including in children, particularly with respect to ocular toxicity<sup>1</sup>. Ocular toxicity is dose- and duration-dependent. It is uncommon when ethambutol is used at the recommended dose for 2 months.

**Pyrazinamide**

Pyrazinamide is usually well tolerated however, it may cause hepatotoxicity, gout, arthralgias and photosensitivity.

8.3.2 Other drugs

Two second-line drugs are also used in the treatment of DS-TB: moxifloxacin (Section 8.4.1) and ethionamide (Section 8.4.3).
References

https://apps.who.int/iris/bitstream/handle/10665/112360/9789241548748_eng.pdf?sequence=1
8.4 Drugs for drug-resistant tuberculosis

Drugs used for DR-TB treatment (except bedaquiline) are taken 7 days a week.

8.4.1 Group A drugs

Table 8.3 – Main characteristics of TB drugs Group A

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Classes</th>
<th>Activity</th>
<th>Resistance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Levofloxacin</td>
<td>Fluoroquinolones (FQs)</td>
<td>Bactericidal</td>
<td>• Resistance common in some regions.</td>
</tr>
<tr>
<td>Moxifloxacin</td>
<td></td>
<td></td>
<td>• Cross-resistance between FQs.</td>
</tr>
<tr>
<td>Bedaquiline</td>
<td>Diarylquinolines</td>
<td>Bactericidal</td>
<td>• Partial cross-resistance with Cfz.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>• Growing resistance as use increases.</td>
</tr>
<tr>
<td>Linezolid</td>
<td>Oxazolidinones</td>
<td>Bactericidal</td>
<td>Resistance assumed to be rare due to its limited use.</td>
</tr>
</tbody>
</table>

Fluoroquinolones (levofloxacin, moxifloxacin)

FQs are usually well tolerated. They may cause tendinopathy and QT prolongation. Moxifloxacin is sometimes used at high dose (Mfx⁰) in the presence of low-level resistance to FQs.

Bedaquiline

Bedaquiline is usually well tolerated. It may cause hepatotoxicity and QT prolongation. Bedaquiline has a long half-life (5.5 months). Therefore, adverse effects can persist after the drug is stopped, and if TB is still active, resistance can develop. Bedaquiline is metabolized in the liver by the cytochrome P450 (CYP450) system enzymes. Drugs, which induce or inhibit CYP450, can affect bedaquiline plasma concentrations and should be avoided (Appendix 19).

The extent of cross-resistance bedaquiline/clofazimine and the clinical implications are not fully understood[1][2][3].

Linezolid

Linezolid may cause myelosuppression, dose- and duration-dependent neuropathy and lactic acidosis. Pyridoxine supplementation (vitamin B₆) is recommended for all patients on linezolid, although there is no evidence that pyridoxine can prevent linezolid-induced neuropathy. Adverse effects frequently lead to reducing the dose or discontinuing linezolid. The optimal dose and duration of treatment are not established.

Linezolid has many interactions and overlapping toxicities with other drugs (e.g. risk of serotonin syndrome when administered with serotonergic drugs⁴). However, it is not always possible to avoid concomitant use of these drugs (e.g. even on linezolid, a patient with depression may require an antidepressant).

8.4.2 Group B drugs

Table 8.4 – Main characteristics of TB drugs Group B
Clofazimine

Clofazimine is a QT-prolonging drug. Orange-pink to brownish-black discolouration of the skin and body fluids occur in almost all patients. These changes are reversible and not harmful. Clofazimine has a long half-life (approximately 70 days). Consequently, its adverse effects can persist for several weeks or months after the drug is stopped.

Cycloserine or terizidone

Cycloserine and terizidone are structural analogues used at the same dose. Both drugs may cause neurotoxicity including psychiatric adverse events. To prevent neurotoxicity, pyridoxine (vitamin B₆) should be administered along with these drugs throughout the course of treatment (Appendix 17).

8.4.3 Group C drugs

Table 8.5 – Main characteristics of TB drugs Group C

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Classes</th>
<th>Activity</th>
<th>Resistance</th>
</tr>
</thead>
</table>
| Clofazimine  | Riminophenazine (anti-leprosy drug) | Probably bacteriostatic | • Partial cross-resistance with Bdq.  
• Growing resistance as use increases. |
| Cycloserine  | Analogue of D-alanine             | Bacteriostatic | • Resistance common in areas where it has been used extensively.  
• Full cross-resistance between the 2 drugs. |
| Terizidone   |                                  |               |                                                 |
Delamanid
Delamanid is usually well tolerated. It may cause QT prolongation. It is particularly useful in patients with pre-existing hepatic disease (no reported hepatotoxicity) or HIV infection (no significant drug interactions or overlapping toxicities with antiretrovirals). It is also useful for replacing a Group A or B drug causing toxicity.

Ethambutol
See Section 8.3.1. Vision monitoring is required when ethambutol is administered for more than 2 months (risk of optic neuritis).

Pyrazinamide
See Section 8.3.1.

Carbapenems (imipenem/cilastatin, meropenem)
Imipenem is always combined with cilastatin. Cilastatin has no antibacterial activity, its role is to inhibit a renal enzyme that inactivates imipenem.
Meropenem does not need to be combined with cilastatin, as it is metabolised through a different pathway.
High cost and difficulty with administration limits the use of carbapenems.
Carbapenems may cause gastrointestinal disturbances, neurotoxicity and hypersensitivity reactions.
Meropenem should be used in children and adolescents under 15 years, and if possible, in epileptic patients and patients with TB meningitis (risk of seizures lower than with imipenem/cilastatin).
The first dose is always administered in a health facility so that an eventual hypersensitivity reaction can be managed. If conditions permit, carbapenems can be continued as an outpatient.

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Classes</th>
<th>Activity</th>
<th>Resistance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Delamanid</td>
<td>Nitroimidazooxazines</td>
<td>Bactericidal</td>
<td>• Potential cross-resistance with pretomanid.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>• Resistance assumed to be rare due to its limited use.</td>
</tr>
<tr>
<td>Ethambutol</td>
<td></td>
<td>Bacteriostatic</td>
<td>High prevalence of resistance among MDR/RR-TB patients (&gt; 49% in some settings).</td>
</tr>
<tr>
<td>Pyrazinamide</td>
<td></td>
<td>Bactericidal</td>
<td>High prevalence among MDR/RR-TB patients (&gt; 80% in some areas).</td>
</tr>
<tr>
<td>Imipenem/cilastatin Meropenem</td>
<td>Carbapenems</td>
<td>Full cross-resistance between carbapenems.</td>
<td></td>
</tr>
<tr>
<td>Amikacin Streptomycin</td>
<td>Aminoglycosides</td>
<td>Bactericidal</td>
<td>Partial cross-resistance between the 2 drugs.</td>
</tr>
<tr>
<td>Ethionamide Prothionamide</td>
<td>Thionamides</td>
<td>Weak bacteriostatic</td>
<td>• Full cross-resistance between thionamides.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>• Cross-resistance with isoniazid if inhA mutation present.</td>
</tr>
<tr>
<td>Para-aminosalicylate sodium Para-aminosalicylic acid</td>
<td>Weak bacteriostatic</td>
<td>Common in some regions.</td>
<td></td>
</tr>
<tr>
<td>Isoniazid high-dose</td>
<td></td>
<td></td>
<td>Cross-resistance with thionamides if inhA mutation present.</td>
</tr>
</tbody>
</table>

Delamanid
Delamanid is usually well tolerated. It may cause QT prolongation. It is particularly useful in patients with pre-existing hepatic disease (no reported hepatotoxicity) or HIV infection (no significant drug interactions or overlapping toxicities with antiretrovirals). It is also useful for replacing a Group A or B drug causing toxicity.

Ethambutol
See Section 8.3.1. Vision monitoring is required when ethambutol is administered for more than 2 months (risk of optic neuritis).

Pyrazinamide
See Section 8.3.1.

Carbapenems (imipenem/cilastatin, meropenem)
Imipenem is always combined with cilastatin. Cilastatin has no antibacterial activity, its role is to inhibit a renal enzyme that inactivates imipenem.
Meropenem does not need to be combined with cilastatin, as it is metabolised through a different pathway.
High cost and difficulty with administration limits the use of carbapenems.
Carbapenems may cause gastrointestinal disturbances, neurotoxicity and hypersensitivity reactions.
Meropenem should be used in children and adolescents under 15 years, and if possible, in epileptic patients and patients with TB meningitis (risk of seizures lower than with imipenem/cilastatin).
The first dose is always administered in a health facility so that an eventual hypersensitivity reaction can be managed. If conditions permit, carbapenems can be continued as an outpatient.
Amoxicillin/clavulanic acid is routinely administered prior to carbapenems, as clavulanic acid prevents the development of carbapenem resistance.

**Aminoglycosides (amikacin, streptomycin)**

Aminoglycosides should only be used when no alternative is available. Most DR-TB patients can be treated without aminoglycosides, including some cases of extensively drug-resistant TB (XDR-TB).

Aminoglycosides are nephrotoxic and ototoxic drugs. Streptomycin is less nephrotoxic than other aminoglycosides, but causes vestibular toxicity more frequently. If an aminoglycoside is used, close monitoring is essential (audiometry, electrolytes and renal function). If close monitoring cannot be ensured, aminoglycosides should not be used.

**Note:** kanamycin and capreomycin are no longer recommended, as their use is associated with higher rates of treatment failure and death.

**Thionamides (ethionamide, prothionamide)**

Ethionamide and prothionamide are used at the same dose. They may cause gastrointestinal disturbances, hypothyroidism (especially if co-administered with para-aminosalicylic acid), neuropathy and hepatotoxicity. In diabetic patients, the dose of antidiabetics may need to be adjusted.

**Para-aminosalicylate sodium or para-aminosalicylic acid**

PAS often causes gastrointestinal disturbances and can decrease the absorption of other TB drugs. It may also cause hypothyroidism, especially when co-administered with a thionamide.

**High-dose isoniazid**

See Section 8.3.1. There is limited evidence to support the use of high-dose isoniazid. High-dose isoniazid may cause more adverse effects than the standard dose. It has overlapping toxicity with linezolid (neuropathy) and hepatotoxic drugs. To prevent peripheral neuropathy, pyridoxine (vitamin B₆) should be administered to all patients throughout the course of treatment (Appendix 17).

### 8.4.4 Ungrouped drugs

**Pretomanid**

Pretomanid belongs to the same class as delamanid and has bactericidal activity. It is used only as part of standard regimens for DR-TB in the following combinations: BPaLM, BPaL (and BPaLC in operational research conditions only), see Chapter 10. Regimens that include bedaquiline, pretomanid and linezolid may cause hepatotoxicity, lactic acidosis, myelosuppression, neuropathy and QT prolongation. Pretomanid/delamanid cross-resistance is likely.

### 8.4.5 Other drugs

**Amoxicillin/clavulanic acid**

Amoxicillin/clavulanic acid is administered before each dose of carbapenem. The clavulanic acid component prevents the development of carbapenem resistance. Only formulations with a ratio of 4:1 (e.g. 500/125 mg) or 2:1 (e.g. 250/125 mg) are suitable for this indication. Do not use formulations with a ratio of 8:1 or 7:1.

### References


8.5 Tuberculosis drug formulations

Only quality-assured drugs should be used. Several internationally recognized mechanisms ensure the quality of TB drugs.

8.5.1 Fixed-dose combinations

FDC formulations combine several TB drugs (2, 3 or 4) in the same tablet. FDCs are only available for first-line TB drugs. FDCs improve adherence (decreased pill burden, decreased risk of omission of one or more drugs).

Table 8.6 – Quality-assured FDC formulations

<table>
<thead>
<tr>
<th>FDCs</th>
<th>Available formulations</th>
</tr>
</thead>
<tbody>
<tr>
<td>Children</td>
<td></td>
</tr>
<tr>
<td>HZR</td>
<td>H50 mg/Z150 mg/R75 mg</td>
</tr>
<tr>
<td>HR</td>
<td>H50 mg/R75 mg</td>
</tr>
<tr>
<td>Adults</td>
<td></td>
</tr>
<tr>
<td>EHZR</td>
<td>E275 mg/H75 mg/Z400 mg/R150 mg</td>
</tr>
<tr>
<td>EHR</td>
<td>E275 mg/H75 mg/R150 mg</td>
</tr>
<tr>
<td>HR</td>
<td>H75 mg/R150 mg</td>
</tr>
</tbody>
</table>

Note: when needed in children, ethambutol is given as a single formulation, in addition to the paediatric FDCs.

8.5.2 Individual drugs

Quality-assured single drug formulations are available for all first-line TB drugs. It may be necessary to use them when FDCs cannot be used due to adverse effects or drug interactions.

There are no quality-assured FDCs for second-line TB drugs. The treatment of DR-TB is based on a combination of individual drugs.

8.5.3 Paediatric formulations

Paediatric formulations should be used whenever possible.

However, they are not available for all TB drugs. When the only option is to manipulate the adult formulations:

- Preferably use scored tablets.
- Ensure that tablets/capsules can be split, crushed or opened (e.g. active ingredients may be protected from gastric acidity by an enteric coating).
- If tablets must be crushed (or capsules opened), a fraction of the powder corresponding to the required dose is mixed with food or liquids. Such manipulations should be done immediately before administering the drug. Any remaining powder should be discarded.
- The preparation of extemporaneous formulations using adult formulations is an alternative, however, this can only be considered if there are qualified personnel to ensure preparation in compliance with the appropriate compounding procedures.

Footnotes

(a) Quality assurance:
(b) Supply:
Chapter 9: Treatment of drug-susceptible tuberculosis

9.1 Introduction
9.2 Conventional treatment regimens
9.3 Alternative treatment regimens
9.4 Special situations
9.5 Adjunctive therapy
9.6 Patient monitoring
9.7 Adverse effects
9.8 Treatment adaptation and change of treatment
9.9 Treatment interruptions

Update: October 2022
9.1 Introduction

Drug-susceptible tuberculosis (DS-TB) treatment is indicated:
- When susceptibility to rifampicin and isoniazid is confirmed by drug susceptibility testing (DST), or
- If the probability of resistance to rifampicin and isoniazid is low:
  ▪ while waiting for DST results for rifampicin and/or isoniazid,
  ▪ when susceptibility to rifampicin is confirmed and susceptibility to isoniazid cannot be tested.

The probability of resistance is considered low in the following situations:
- No previous TB treatment;
- No contact with a drug-resistant TB (DR-TB) patient;
- The patient comes from an area of low prevalence of resistance according to drug resistance surveys.

Patients with DS-TB should start a conventional regimen based on first-line drugs (Table 9.1) or, if eligible, an alternative regimen (Table 9.2).
All regimens for DS-TB are standard regimens.

For dosages of fixed-dose combinations see Appendix 13.
For dosages of individual drugs see Appendix 10.
9.2 Conventional treatment regimens

Table 9.1 – Conventional DS-TB regimens according to the infection site

<table>
<thead>
<tr>
<th>Regimen Duration</th>
<th>Eligibility</th>
</tr>
</thead>
<tbody>
<tr>
<td>2(HRZE)/2(HR) 4 months</td>
<td>Children &gt; 3 months and adolescents &lt; 16 years with[^{[1]}]: Pulmonary TB (PTB) - microscopy smear-negative or Xpert result “negative”, “trace”, “very low” and “low” or - clinically diagnosed with TB lesions confined to one lobe and no cavities on chest x-ray (CXR) Extrapulmonary TB (EPTB) non severe, i.e.: - pleural effusion without complications (e.g. no empyema, pneumothorax or fistula) - extra- or intra-thoracic lymph node TB with no airway obstruction</td>
</tr>
<tr>
<td>2(HRZE)/4(HR) 6 months</td>
<td>PTB and EPTB (except miliary TB, TB meningitis and bone and joint TB)[^{[2]}] Adolescents ≥ 16 years and adults Children and adolescents &lt; 16 years not eligible for the 4-month regimen or when the national protocol does not include the 4-month regimen.</td>
</tr>
<tr>
<td>2(HRZE)/10(HR) 12 months</td>
<td>Miliary TB and TB meningitis[^{[3]}] All children, adolescents and adults.</td>
</tr>
<tr>
<td>2(HRZE)/7-10(HR) 9-12 months</td>
<td>Bone and joint TB[^{[4]}] All children, adolescents and adults.</td>
</tr>
</tbody>
</table>

If bacteriological testing and/or CXR are not available, children meeting the following criteria are eligible for the 4-month regimen 2(HRZE)/2(HR):
- Signs and symptoms not requiring hospitalisation\[^{[a]}\].
- Extra-thoracic lymph node TB without involvement of other EP sites.

If after one month of treatment symptoms have completely resolved, continue treatment until the end. If symptoms have not completely resolved, further investigations are needed.
If after 4 months of treatment symptoms have not completely resolved and/or there is no weight gain, further investigation is needed.
The treatment can be extended to 6 months if causes of non-response to treatment (including DR-TB, non-adherence and non-TB disease) are ruled out or unlikely.

Ethambutol can be removed from the 4- and 6-month regimens in non-HIV-infected children living in areas where the prevalence of HIV and/or isoniazid resistance is low with:
- PTB microscopy smear-negative, or
- Extra- or intra-thoracic lymph node TB\[^{[5]}\].

For spinal TB, rest and back support bracing are indicated in addition to drug therapy. For patients with neurological deficit or unstable spine lesion, surgery can also be considered.

Footnotes
(a) Symptoms requiring hospitalisation: signs of severe respiratory disease or distress, severe acute malnutrition, fever > 39 °C, severe pallor, restlessness, irritability or lethargy, etc.
References


9.3 Alternative treatment regimens

Table 9.2 – Alternative DS-TB regimens according to the infection site

<table>
<thead>
<tr>
<th>Regimens</th>
<th>Duration</th>
<th>Eligibility</th>
</tr>
</thead>
</table>
| 2HPZ-Mfx/2HP-Mfx | 4 months | PTB and non-severe EPTB[^1][^2]
|           |          | Adolescents ≥ 12 years and adults meeting all the following criteria: |
|           |          | • Weight ≥ 40 kg |
|           |          | • CD4 ≥ 100 if HIV-infected |
|           |          | • No resistance to fluoroquinolones (FQs) or living in areas where the prevalence of FQs resistance is low. |
| 6HRZ-Eto | 6 months | TB meningitis[^3]
|           |          | Non-HIV infected children and adolescents under 20 years without inhA mutation detected. |

Regimen 2HPZ-Mfx/2HP-Mfx

- This regimen is an alternative to the conventional regimens for PTB and EPTB in eligible patients.
- Implementation requires DST to FQs and supply of rifapentine.
- There are no fixed-dose combinations (FDC) for this regimen which makes treatment adherence more difficult.

Regimen 6HRZ-Eto

- Small studies have shown lower mortality, but more neurological sequelae with the 6HRZ-Eto regimen compared to the 12-month conventional regimen. However, no clinical trials have been conducted to compare the two regimens[^3].
- The advantages of this regimen are short duration and better central nervous system penetration of ethionamide compared to ethambutol.
- Implementation requires supply of ethionamide.
- There are no FDC for this regimen which makes treatment adherence more difficult.
- The daily doses of TB drugs in this regimen are higher than those of other regimens:
  - isoniazid 20 mg/kg daily (max. 400 mg)
  - rifampicin 20 mg/kg daily (max. 600 mg)
  - pyrazinamide 40 mg/kg daily (max. 2 g)
  - ethionamide 20 mg/kg daily (max. 750 mg)

References


9.4 Special situations

9.4.1 Women (pregnant or breastfeeding or of childbearing age)

Pregnant or breastfeeding women

- All first-line TB drugs, except rifabutin and rifapentine, can be used during pregnancy and breastfeeding.\[1\]
- Isoniazid may cause peripheral neuropathy due to vitamin B\(_6\) (pyridoxine) deficiency:
  - Pregnant and breastfeeding women should receive pyridoxine PO (10 mg once daily) throughout the course of TB treatment.
  - Breast-fed neonates or infants should receive pyridoxine PO (5 mg once daily).
- Rifampicin may cause clotting disorders due to increased vitamin K (phytomenadione) metabolism:
  - Women in late pregnancy on rifampicin (or rifabutin) should receive phytomenadione PO (10 mg once daily) for 2 weeks prior to expected date of delivery.
  - Neonates should also receive phytomenadione IM at birth (1 mg single dose) to prevent haemorrhagic disease of the newborn.
- Regimens containing rifapentine, moxifloxacin\[2\] and ethionamide cannot be used to treat DS-TB in pregnant and breastfeeding women.

Women of childbearing age

Women on contraception should use an intra-uterine device or a progestogen-only injectable throughout the courses of TB treatment, as rifamycins reduce the effectiveness of implants and oral contraceptives.

9.4.2 Malnutrition or risk of malnutrition

- For patients with malnutrition, therapeutic feeding should be initiated.
- For children with severe acute malnutrition, a 6-month regimen is preferred over a 4-month regimen until more data on the efficacy of the 4-month regimen in these patients become available.
- For at-risk populations, such as children, pregnant and breastfeeding women and the elderly, nutritional supplementation with a standard food package or ready-to-use food may be considered during the first 2 months of treatment.

9.4.3 Diabetes

TB can impair glycaemic control in patients with diabetes.\[3\] It is necessary to increase blood glucose monitoring in these patients. TB drugs can exacerbate complications of diabetes (e.g. peripheral neuropathy). Avoid prescribing ethambutol in patients with pre-existing diabetic retinopathy.

Rifampicin can reduce the effect of sulfonylureas (e.g. glibenclamide, gliclazide). In contrast, first-line TB drugs have no interactions with metformin.

If diabetes is diagnosed, treat and monitor according to standard protocols.

At the end of TB treatment, it is recommended to schedule a specialist consultation for a complete evaluation and, if necessary, adjust antidiabetic treatment.

9.4.4 Renal insufficiency

In patients with renal insufficiency, creatinine clearance should be calculated. If it is less than 30 ml/minute, doses of certain TB drugs should be adjusted.

For the formula to estimate the creatinine clearance and dose adjustments in renal insufficiency see Appendix 12.

References


9.5 Adjunctive therapy

9.5.1 Pyridoxine prophylaxis

Pyridoxine (vitamin B₆) prophylaxis is indicated for all patients at risk of peripheral neuropathy, i.e. pregnant or breastfeeding women and patients with HIV infection, chronic alcohol use, malnutrition, diabetes, chronic hepatic disease or renal impairment (see Appendix 17).

9.5.2 Corticosteroid therapy

Corticosteroid therapy is indicated for:

- TB meningitis[1] and pericarditis[2];

There is insufficient evidence regarding the use of corticosteroids in other indications[3][4].

Table 9.3 – Corticosteroid treatment

<table>
<thead>
<tr>
<th>Indications</th>
<th>Dosage and duration</th>
</tr>
</thead>
<tbody>
<tr>
<td>TB meningitis[5]</td>
<td>dexamethasone IV/PO</td>
</tr>
<tr>
<td></td>
<td>Child: 0.6 mg/kg once daily for 4 weeks, tapered off over 4 weeks</td>
</tr>
<tr>
<td></td>
<td>Adult: 0.4 mg/kg once daily for 7 days, tapered off over 6 to 8 weeks</td>
</tr>
<tr>
<td>TB pericarditis</td>
<td>prednisolone PO</td>
</tr>
<tr>
<td></td>
<td>Child: 1.5 mg/kg once daily for 4 weeks, tapered off over 6 weeks</td>
</tr>
<tr>
<td></td>
<td>Adult: 60 mg once daily for 4 weeks, tapered off over 6 weeks</td>
</tr>
</tbody>
</table>

References


5. BMJ Best Practice. Extrapulmonary tuberculosis [Accessed 01 March 2023]
9.6 Patient monitoring

Patients should be assessed at baseline, then, regardless of the regimen prescribed, monitored throughout the course of treatment.

Monitoring includes:

- Assessment of treatment response
- Detection of adverse effects and adherence issues.

For the schedule of follow-up examinations see Appendix 14.

Baseline and follow-up findings should be noted in the patient file to enable the detection and interpretation of potential changes.

9.6.1 Clinical visits

Baseline assessment

Assessment includes:

- Symptoms of TB and their severity (cough, fever, night sweats, weight loss, shortness of breath, ability to perform daily activities).
- Vital signs and weight.
- Comorbidities and other risk factors for adverse effects requiring monitoring adaptation.
- Psychological assessment.

Other investigations may be needed depending on the drugs used in the regimen prescribed (Section 9.6.3).

Clinical assessment should be performed by a clinician. Psychological assessment should be performed whenever possible by personnel with appropriate training.

All patients starting treatment should be given the information they need to understand the disease and its treatment (Appendix 21).

Follow-up visits

Each follow-up visit, assessment includes:

- Clinical progress, vital signs and weight. Dosages should be adjusted to the weight if necessary.
- Occurrence of adverse effects.
- Adherence to treatment (Appendix 22).
- Psychological condition.

Frequency of visits depends on the patient’s clinical condition and evolution:

- A visit every other week for the first month, then once a month if there is no particular problem.
- Additional visits may be required in case of comorbidities, severe or multiple adverse effects, pregnancy, etc.

Visits should coincide with bacteriological examinations and other investigations when possible.

The clinician should take into account any information and concerns regarding treatment tolerance and adherence reported by the patient or the team responsible for the patient’s follow-up and support.

9.6.2 Bacteriological tests

To assess treatment response in patients with:

- PTB: bacteriological tests are essential.
- EPTB: evaluation is based on clinical evolution. However, bacteriological tests are required if patients also develop PTB.

Baseline tests

Baseline tests are those performed on specimens collected just prior to treatment initiation. They include:

- Rapid molecular tests (RMTs) for detection of *M. tuberculosis* and rifampicin and isoniazid resistance.
- Smear microscopy to monitor treatment progress.
- Culture and phenotypic DST (pDST) when indicated.

For more information see Chapter 3.

Follow-up tests

- Smear microscopy
Microscopy should be performed every 2 months until treatment completion. If treatment is effective, microscopy at Month 2, 4 and 6 should be negative.

**Notes:**
- Patients with high bacillary load at baseline may have dead bacilli in their sputum for several months.
- As microscopy cannot distinguish dead from live bacilli, a positive result does not necessarily indicate that the treatment has failed.

- Rapid molecular tests
  RMTs cannot be used to monitor treatment progress. However, if microscopy or culture is positive at Month 2 or later, RMTs should be performed to detect the emergence of new drug resistance not present at baseline (**Chapter 3**).

- Culture and pDST
  Culture and pDST should be performed:
  - at Month 2 or later, if RMTs show a new resistance to rifampicin or isoniazid;
  - at Month 4, if microscopy is positive.
  Full pDST (for first- and second-line drugs) should be performed on any positive culture.
  **Note:** bacteriological tests are performed at the end of the month (e.g. Month 2 means the end of the 2nd month of treatment).

Regardless of the above schedule, RMTs, culture and pDST should be performed if the patient’s clinical condition deteriorates.

**End-of-treatment test**

Microscopy should be performed at end of treatment to confirm the end-of-treatment outcome (**Chapter 17**).

### 9.6.3 Other investigations

**Radiography**

- CXR: for children with presumptive PTB, patients with non-bacteriologically confirmed PTB, suspicion of other intra-thoracic TB at baseline, then if indicated (e.g. worsening respiratory symptoms, non-response to TB treatment).
- Bone x-ray: for patients with osteoarticular and spinal TB at baseline, then every 6 months.

**Biological tests**

**Table 9.4 – Blood tests at baseline and during treatment**

<table>
<thead>
<tr>
<th>Tests</th>
<th>Indications</th>
</tr>
</thead>
<tbody>
<tr>
<td>Full blood count**(a)**</td>
<td>HIV-infected patients on rifabutin or isoniazid (AZT), at baseline, then once a month for the first 2 months, then if indicated.</td>
</tr>
<tr>
<td>Liver function tests**(b)**</td>
<td>Patients with pre-existing hepatic disease, at baseline, then once a month.</td>
</tr>
<tr>
<td>Serum creatinine**(c)**</td>
<td>Patients with renal insufficiency at baseline, then if indicated.</td>
</tr>
<tr>
<td>HbA1C and/or blood glucose level</td>
<td>All patients, at baseline, to detect diabetes. If diabetes is detected, monitor according to standard protocols.</td>
</tr>
<tr>
<td>HIV, hepatitis B and C</td>
<td>For patients with undocumented HIV, hepatitis B and C status; HIV test every 6 months in high HIV prevalence areas. Tests can be repeated in case of recent exposure.</td>
</tr>
<tr>
<td>CD4 count and viral load</td>
<td>HIV-infected patients: at baseline, then every 6 months.</td>
</tr>
</tbody>
</table>

**(a)** Haemoglobin, red and white blood cells, platelets.
**(b)** Aspartate aminotransferase (AST) and alanine aminotransferase (ALT). Bilirubin if elevated liver enzymes.
**(c)** For estimation of creatinine clearance see **Appendix 12**.
### 9.7 Adverse effects

Rapid management of adverse effects is essential to increase tolerance and improve outcomes. In the event of minor adverse effects, drugs should not be stopped. Providing support and using ancillary medicines is all that is necessary. In the event of major adverse effects, the regimen may need to be adapted.

#### Table 9.5 – Main adverse effects and likely responsible drugs

<table>
<thead>
<tr>
<th>Adverse effects</th>
<th>Drug(s) likely responsible</th>
<th>Management</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Minor</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nausea, vomiting</td>
<td>Eto, Z</td>
<td>Appendix 17</td>
</tr>
<tr>
<td>Arthralgia</td>
<td>Z</td>
<td>Appendix 17</td>
</tr>
<tr>
<td>Peripheral neuropathy</td>
<td>H, Eto</td>
<td>Appendix 17</td>
</tr>
<tr>
<td>Orange/red urine, tears, etc.</td>
<td>R, P</td>
<td>Patients should be told that this is normal before starting treatment.</td>
</tr>
<tr>
<td><strong>Major</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Skin reactions</td>
<td>E, Z, R, H, P, Mfx, Eto</td>
<td>Appendix 17</td>
</tr>
<tr>
<td>Hepatotoxicity</td>
<td>Z, H, R, P, Eto</td>
<td>Appendix 17</td>
</tr>
<tr>
<td>Optic neuritis</td>
<td>E</td>
<td>Appendix 17</td>
</tr>
<tr>
<td>Haematologic disorders</td>
<td>R, P, H, E</td>
<td>Appendix 17</td>
</tr>
</tbody>
</table>

For more information on individual drugs see [Appendix 10](#).
9.8 Treatment adaptation and change of treatment

9.8.1 Treatment adaptation

The whole treatment or individual drug(s) may be temporarily interrupted by the clinician in case of severe adverse effects (Appendix 17). This is considered as treatment adaptation, as long as it does not meet the definition of “treatment failure” (Chapter 17).

9.8.2 Change of treatment

The clinician should replace the DS-TB treatment with:

- A treatment for isoniazid-resistant TB when RMT or pDST show:
  - the development of isoniazid resistance (Chapter 11) after treatment initiation, or
  - undetected isoniazid resistance at baseline, for any reason.

- A treatment for multidrug-resistant or rifampicin-resistant TB (MDR/RR-TB, see Chapter 10) in the following circumstances:\[1\],
  - Development of rifampicin resistance after treatment initiation.
  - Rifampicin resistance not detected at baseline, for any reason.
  - No bacteriological conversion or bacteriological reversion (Chapter 17).
  - Insufficient clinical response to treatment in patients:
    - with non-bacteriologically confirmed TB (e.g. miliary TB, some forms of EPTB, TB in children).
    - with bacteriologically confirmed TB, when the bacteriological response cannot be assessed, or the result is inconclusive.

The above treatment changes meet the outcome definition of “treatment failure” except when the reason for change is a resistance undetected at baseline\[1\] (Chapter 17).

References

9.9 Treatment interruptions

Treatment interruptions can lead to the emergence of new resistances. Problems of treatment interruption by the patient (e.g. discontinuation of certain drugs, recurrent treatment interruptions) should be detected and addressed (management of adverse effects if necessary and reinforcement of patient support measures). Interruption of the entire treatment for two consecutive months or more meet the definition of “lost to follow-up” (Chapter 17).

Table 9.6 – Management of patients who interrupt treatment

<table>
<thead>
<tr>
<th>Length of treatment before interruption</th>
<th>Length of interruption</th>
<th>Management</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 1 month</td>
<td>&lt; 2 weeks</td>
<td>Continue treatment at the point it was stopped. Doses missed during interruption must be made up to complete the treatment.</td>
</tr>
<tr>
<td>2-7 weeks</td>
<td></td>
<td>Restart treatment or perform RMTs (see below) depending on patient’s clinical evolution.</td>
</tr>
</tbody>
</table>
| ≥ 8 weeks                              |                        | Perform RMTs:  
  • if no resistance, restart treatment.  
  • if resistance, start DR-TB treatment. |
| ≥ 1 month                              | < 2 weeks              | Continue treatment at the point it was stopped. Doses missed during interruption must be made up to complete treatment. |
| ≥ 4 weeks                              |                        | Perform RMTs:  
  • if no resistance, restart treatment.  
  • if resistance, start DR-TB treatment. |

For patients on 6-month regimen who have received adequate treatment for 4 months or more, who return smear negative, are in good clinical condition and with no resistance detected, the decision to re-start a treatment is considered on a case-by-case basis.

When a DST is not feasible (e.g. miliary TB, some forms of EPTB, TB in children), clinical and radiological evaluation should guide the decision to either restart DS-TB treatment or switch to an DR-TB treatment.
Chapter 10: Treatment of multidrug-resistant and rifampicin-resistant tuberculosis

10.1 Introduction
10.2 Treatment regimens in programmatic conditions
10.3 Treatment regimens in operational research conditions
10.4 Special situations
10.5 Adjunctive therapy
10.6 Patient monitoring
10.7 Adverse effects
10.8 Treatment adaptation and change of treatment
10.9 Treatment interruptions
10.10 Surgery
10.11 Treatment failure and palliative care

Update: February 2023
10.1 Introduction

When selecting or building a treatment regimen for multidrug-resistant tuberculosis (MDR-TB) and rifampicin-resistant tuberculosis (RR-TB), the following should be considered:

10.1.1 Standard short regimens and individualized long regimens

Patients should receive a standard short treatment regimen (STR) except if they do not meet the eligibility criteria for STRs, or do not tolerate STRs. In such cases, patients require an individualized long treatment regimen (LTR).

It may be necessary to switch from an STR to an LTR, based on the latest drug-susceptibility test (DST) results and/or clinical evolution during treatment course (e.g. drug intolerance, persistence of a positive culture).

10.1.2 Likely effective drugs

Treatment is based on a combination of “likely effective” TB drugs.

Table 10.1 - Definition of likely effective drugs (adapted from WHO[1])

<table>
<thead>
<tr>
<th>DST</th>
<th>Definition of a likely effective TB drug</th>
</tr>
</thead>
<tbody>
<tr>
<td>Available and reliable</td>
<td>DST indicates susceptibility to the drug.</td>
</tr>
<tr>
<td>Unavailable, unreliable, or result pending</td>
<td>The following criteria should be met:&lt;br&gt;• No resistance detected by DST to a drug with cross-resistance.&lt;br&gt;• No resistance to the drug or to a drug with a cross-resistance to it detected by DST in the presumed source case.&lt;br&gt;• No previous exposure (&gt; 1 month) to the drug or to a drug with a cross-resistance.&lt;br&gt;• The drug has not been widely used in the treatment of TB or drug resistance surveys indicate that drug resistance is rare in the area the patient comes from.</td>
</tr>
</tbody>
</table>

When the criteria of a likely effective drug are not met:
• If the strain of the patient (or the presumed source case) is resistant to clofazimine, bedaquiline can be used but not counted as a likely effective drug until DST demonstrates susceptibility to bedaquiline. The same applies to all drugs with known or potential cross-resistance (e.g. delamanid/pretomanid). For more information on drug resistance and cross-resistance see Chapter 8.
• If a drug has been widely used and there is no reliable DST for this drug (e.g. ethambutol, cycloserine, para-aminosalicylate sodium): it can be used but never counted as a likely effective drug.
• If a drug has been widely used and there is a reliable DST for this drug (e.g. pyrazinamide): it can be used but not counted as a likely effective drug until DST demonstrates susceptibility.

10.1.3 Other considerations

The following should also be considered when choosing or building a treatment regimen:
• Interactions and overlapping toxicities between TB drugs or other drugs the patient may take (see Appendix 10 for individual drugs and Appendix 19 for co-administration of TB drugs and antiretrovirals);
• Comorbidities that can result in increased drug toxicity;
• Absolute contraindications to any drug included in a regimen;
• Pregnancy and breastfeeding (Appendix 11).

References

10.2 Treatment regimens in programmatic conditions

10.2.1 Short treatment regimens

A. 6-month BPaLM regimen

Eligibility

(adapted from WHO[1])

BPaLM is the preferred treatment regimen for all MDR/RR-TB patients meeting the following criteria:
1. Bedaquiline, pretomanid, linezolid and moxifloxacin are likely effective.
2. Age ≥ 14 years.
3. No miliary TB, osteoarticular TB or TB of the central nervous system (CNS), i.e. brain, spinal cord or meninges.

Regimen composition

Box 10.1 - 6-month BPaLM regimen

6Bdq-Pa-Lzd-Mfx

In the BPaLM regimen, the starting dose of linezolid (600 mg once daily) is reduced after 16 weeks to 300 mg once daily.

Note:

In the event of resistance to fluoroquinolones (FQs), this guide recommends a short four-drug regimen under operational research conditions where possible (Section 10.3). An LTR or BPaL (6-Bdq-Pa-Lzd with linezolid given at 600 mg for 6 months) can be used in settings where an STR under operational research conditions is not feasible. Note that linezolid causes frequent toxicity. Management of linezolid adverse effects include temporary or early permanent interruption of the drug. If interruptions are recurrent or linezolid is stopped permanently early in the treatment, patients on BPaL would receive a two-drug regimen for a significant period of time, which is not optimal[1].

B. 9-month bedaquiline-containing regimens[1]

Eligibility

If pretomanid is not available or for patients not eligible for the BPaLM regimen, a 9-month STR should be used in MDR/RR-TB patients meeting the following criteria[8]:
1. Susceptibility to FQs is confirmed by a rapid molecular test (RMT).
2. Other drugs used in the regimen, except isoniazid, are likely effective (Section 10.1.2). High treatment failure rates and amplification of FQ resistance have been observed in some countries[9] in patients with strains presenting resistance to other drugs in the STR.
3. No extensive pulmonary TB (PTB):
   - no bilateral lung cavities or extensive lung damage,
   - no cavities or bilateral disease in patients < 15 years.
4. No severe extrapulmonary TB (EPTB):
   - no miliary TB, osteoarticular TB, TB of the CNS, or pericardial TB,
   - no EPTB other than lymph node TB (peripheral nodes or isolated mediastinal mass without compression) in patients < 15 years.
5. No pregnancy or breastfeeding for the ethionamide-containing regimen.

Regimen composition

Box 10.2 - 9-month bedaquiline-containing regimens
This guide recommends the regimen that includes linezolid (a drug from Group A), rather than the regimen that includes ethionamide.

Regimen 4 to 6Bdq<sub>6</sub>-Lfx-Cfz-Z-E-H<sup>h</sup>-Lzd<sub>2</sub>/5Lfx-Cfz-Z-E or 4 to 6Bdq<sub>6</sub>-Lfx-Cfz-Z-E-H<sup>h</sup>-Eto/5Lfx-Cfz-Z-E

In practice the patient receives:

A 9-month regimen if microscopy negative at Month 4:

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Intensive phase</th>
<th>Continuation phase</th>
</tr>
</thead>
<tbody>
<tr>
<td>Months</td>
<td>M1</td>
<td>M2</td>
</tr>
<tr>
<td>Bdq</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lfx-Cfz-Z-E</td>
<td></td>
<td></td>
</tr>
<tr>
<td>H&lt;sup&gt;h&lt;/sup&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lzd</td>
<td></td>
<td></td>
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</tbody>
</table>

An 11-month regimen if microscopy positive at Month 4:

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Intensive phase</th>
<th>Continuation phase</th>
</tr>
</thead>
<tbody>
<tr>
<td>Months</td>
<td>M1</td>
<td>M2</td>
</tr>
<tr>
<td>Bdq</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lfx-Cfz-Z-E</td>
<td></td>
<td></td>
</tr>
<tr>
<td>H&lt;sup&gt;h&lt;/sup&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lzd</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Regimen 4 to 6Bdq<sub>6</sub>-Lfx-Cfz-Z-E-H<sup>h</sup>-Eto/5Lfx-Cfz-Z-E

- Bdq<sub>6</sub> means that bedaquiline is administered for 6 months (not for 4 months). However, it should be extended to 9 months if the sputum microscopy is positive at Month 4.
- Lzd<sub>2</sub> means that linezolid is administered for 2 months (not for 4 months).
- H<sup>h</sup>-Eto are administered for 4 months or extended to 6 months if the sputum microscopy is positive at Month 4.
- Lfx-Cfz-Z-E are administered for 9 months or extended to 11 months if the sputum microscopy is positive at Month 4.
- Moxifloxacin (Mfx) at standard dose can be used instead of levofloxacin.

In practice the patient receives:

A 9-month regimen if microscopy negative at Month 4:
An 11-month regimen if microscopy positive at Month 4:

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Intensive phase</th>
<th>Continuation phase</th>
</tr>
</thead>
<tbody>
<tr>
<td>Months</td>
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<td>M2</td>
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<tr>
<td>Bdq</td>
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<td></td>
</tr>
<tr>
<td>Lfx-Cfz-Z-E</td>
<td></td>
<td></td>
</tr>
<tr>
<td>$H^b$-Eto</td>
<td></td>
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</tr>
</tbody>
</table>

10.2.2 Long treatment regimens

Eligibility

All MDR/RR-TB patients not eligible for STRs (programmatically or under operational research).

Regimen composition

The regimen should include a minimum number of likely effective drugs.

Box 10.3 - Number of likely effective drugs required in LTRs

At least 4 likely effective TB drugs, including:
- 3 from Group A
- 1 from Group B

If this optimal combination is not feasible:
At least 5 likely effective TB drugs, prioritizing Group A and B drugs and adding Group C drug(s) to bring the total to at least 5 TB drugs.

LTRs may contain more than 5 TB drugs if there is uncertainty of effectiveness in some of the drugs used.

While waiting for full DST results, patients can be treated with:
- An individualized LTR, or
- An empirical LTR according to the known resistance profile.

It may be necessary to switch from an empirical LTR to an individualized LTR, based on the latest DST results and/or clinical evolution during treatment course (e.g. drug intolerance, persistence of a positive culture).

Individualized long regimens

To build an individualized LTR, a stepwise process is recommended.

Table 10.3 - Steps to build an LTR
### Empirical long regimens

**Table 10.4 - Examples of empirical long regimens at treatment initiation**

<table>
<thead>
<tr>
<th>Resistance profiles</th>
<th>Examples</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Group A and B drugs likely effective</strong></td>
<td>18Lfx-Bdq-Lzd-Cfz</td>
</tr>
<tr>
<td></td>
<td>If Bdq is contra-indicated: 18Lfx-Lzd-Cfz-Cs-Dlm</td>
</tr>
<tr>
<td></td>
<td>If Lzd is contra-indicated: 18Lfx-Bdq-Cfz-Cs-Dlm</td>
</tr>
<tr>
<td><strong>FQs not likely effective</strong></td>
<td>18Bdq-Lzd-Cfz-Cs-Dlm-[Mfx]</td>
</tr>
<tr>
<td><strong>Other Group A and B drugs likely effective</strong></td>
<td>18Lzd-Cfz-Cs-Dlm-Ipm/Cln</td>
</tr>
</tbody>
</table>

(a) Moxifloxacin high dose can be used, but not counted if low-level FQ resistance is suspected or found on DST.
Duration of treatment

At least for 18 months, with at least 15 months after culture conversion (for definition see Chapter 17). If well tolerated, all drugs should be taken for the full treatment duration\(^6\)[\(^6\)].

Preliminary evidence suggests that stopping bedaquiline at 6 months is associated with high rates of culture reversion (for definition see Chapter 17) in patients with resistance to several drugs or extensive lung damage\(^8\). No safety issues have been reported with bedaquiline treatment longer than 6 months\(^2\)[\(^2\)][\(^4\)]\(^7\).

Carbapenems are commonly used for a minimum of 2 months after culture conversion. When the number of likely effective drugs included in the regimen is limited, a carbapenem may be required for the entire duration of treatment.

Footnotes

(a) An empirical regimen is a regimen designed to treat most patients in a region whilst waiting the full DST results.

References

   [Link](https://www.who.int/publications/i/item/9789240065116)

   [Link](https://www.who.int/publications/i/item/9789240006997)

   [Link](https://doi.org/10.1183/23120541.00537-2020)

   [Link](http://www.endtb.org/sites/default/files/2018-07/endTB%20interim%20analysis%20%2813%20July%202018%20%29.pdf)

   [Link](https://doi.org/10.1183/13993003.01799-2016)

   [Link](https://doi.org/10.5588/ijtld.17.0840)

10.3 Treatment regimens in operational research conditions

MDR/RR-TB patients can be treated under operational research conditions with short regimens other than the standard STRs. Whatever the rationale or results of operational research, they should be communicated as they may complement those of clinical trials.

10.3.1 Operational research conditions

The requirements for conducting operational research include:
- A study protocol including 12-month post-end-of-treatment follow-up.
- A clinical treatment guide.
- A patient consent process.
- Approval by an ethics review board and Ministry of Health.
- A pharmacovigilance system (core aDSM*).

Study protocol templates are available from the Global Drug-resistant TB Initiative (GDI)* and WHO*.

10.3.2 Treatment regimens under investigation

The following regimens are examples of standardized regimens that can be used under operational research conditions. These regimens have been reviewed by scientific committees and have been tested, or are currently being tested, in clinical trials.

For FQ-susceptible MDR/RR-TB patients
- The five endTB trial experimental regimens[^1]:
  - 9Bdq-Lzd-Mfx-Z
  - 9Bdq-Cfz-Lzd-Lfx-Z
  - 9Bdq-Dlm-Lzd-Lfx-Z
  - 9Dlm-Cfz-Lzd-Lfx-Z
  - 9Dlm-Cfz-Mfx-Z
- The TB-PRACTECAL regimen that includes clofazimine[^2]: 6Bdq-Pa-Lzd-Cfz (BPaLC regimen)

For FQ-resistant MDR/RR-TB patients
- The endTB-Q trial regimen: 6 or 9Bdq-Dlm-Lzd-Cfz[^3]
- The TB-PRACTECAL regimen that includes clofazimine: 6Bdq-Pa-Lzd-Cfz (BPaLC regimen)

Footnotes

References
1. [https://clinicaltrials.gov/ct2/show/NCT02754765](https://clinicaltrials.gov/ct2/show/NCT02754765)
2. [https://clinicaltrials.gov/ct2/show/NCT02589782](https://clinicaltrials.gov/ct2/show/NCT02589782)
10.4 Special situations

10.4.1 Women (pregnant or breastfeeding or of childbearing age)

Pregnant women

Early treatment initiation after diagnosis is recommended. For choosing or building a regimen, see Appendix 11. Pregnancy outcome and any congenital anomalies in the neonate should be documented.

Breastfeeding women

Use of infant formula is recommended as many second-line drugs should be avoided in breastfeeding women (Appendix 11). Mothers must be informed of its benefits and risks and provided with infant formula, clean water, fuel for boiling water and a heating device (stove, saucepan and bottles). They must also receive training on how to prepare and use the formula. When infant formula cannot be used safely, infants must be breastfed.

If the mother is smear-positive, mother-infant contact should be maintained, but kept to a minimum. Appropriate infection prevention and control measures should be taken during contact. Care of the infant should be largely entrusted to family members until the mother becomes smear-negative.

Women of childbearing age

A pregnancy test should be performed before starting treatment and, if necessary, repeated during treatment. A highly effective contraception method (e.g. intra-uterine device or implantable hormonal contraceptive) should be offered prior to starting treatment.

10.4.2 Children and adolescents

Given the severity of MDR/RR-TB, no TB drugs are contra-indicated (except pretomanid while waiting data on appropriate dosing)[1]. Children and adolescents generally tolerate second-line TB drugs well. They should be treated without delay based on the index case resistance profile when DST is not available (e.g. clinically diagnosed TB, EPTB).

Children and adolescents should receive an STR when eligible, however BPaLM and BPaL regimens are not recommended in patients under 14 years.

Children with non-severe TB receiving an LTR can usually be treated for less than 18 months[2]. Some experts suggest that even severe TB could be treated for less than 18 months[3].

10.4.3 Patients with malnutrition or risk of malnutrition

See Chapter 9.

10.4.4 Extrapulmonary tuberculosis

Patients with some forms of EPTB are not eligible for STRs (Section 10.2.1) and should be treated with an LTR as described in Section 10.2.2.

For patients with TB of the CNS, drug penetration into the CNS should be taken into account.

Table 10.5 - Choice of TB drugs for TB of the CNS[4][5]
If the regimen contains a carbapenem, use preferably meropenem in patients with TB meningitis (less risk of seizures than with imipenem/cilastatin).

### 10.4.5 Diabetes

TB can impair glycaemic control in patients with diabetes. It is therefore necessary to increase blood glucose monitoring in these patients.

TB drugs may exacerbate complications of diabetes (e.g. peripheral neuropathy). Avoid prescribing ethambutol or linezolid for patients with pre-existing diabetic retinopathy.

If diabetes is diagnosed, treat and monitor according to standard protocols. At the end of TB treatment, it is recommended to schedule a specialist consultation for a complete evaluation and, if necessary, adjustment of antidiabetic treatment.

### 10.4.6 Renal insufficiency

In patients with renal insufficiency, creatinine clearance should be calculated. If it is less than 30 ml/minute, doses of certain TB drugs should be adjusted.

For the formula to estimate creatinine clearance and dose adjustments in renal insufficiency see Appendix 12.

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**References**


10.5 Adjunctive therapy

10.5.1 Pyridoxine prophylaxis

Pyridoxine (vitamin B₆) is routinely administered to all patients receiving linezolid, cycloserine or teridizone, thionamides or isoniazid high dose to prevent neurotoxic effects (Appendix 17).

10.5.2 Corticosteroid therapy

See Chapter 9.
10.6 Patient monitoring

Patients should be assessed at baseline, then, regardless of the regimen prescribed, monitored throughout the course of treatment. Monitoring includes:

- Assessment of treatment response.
- Detection of adverse effects and adherence issues.

For the schedule of follow-up examinations see Appendix 15.

Baseline and follow-up findings should be noted in the patient file to enable the detection and interpretation of potential changes.

10.6.1 Clinical visits

Baseline assessment

Assessment includes:

- Signs and symptoms of TB and severity (cough, fever, night sweats, weight loss, shortness of breath, ability to perform daily activities).
- Vital signs and weight.
- Comorbidities and other risk factors for adverse effects requiring monitoring schedule adaptation.
- Psychological assessment.

Other investigations may be needed depending on the drugs used in the regimen prescribed (Section 10.6.3).

Clinical assessment should be performed by a clinician. Psychological assessment should be performed whenever possible by personnel with appropriate training.

All patients starting treatment should be given the information they need to understand the disease and its treatment (Appendix 21).

Follow-up visits

Each follow-up visit includes assessing:

- Clinical progress, vital signs and weight. Dosages should be adjusted to the weight if necessary.
- Occurrence of adverse effects.
- Adherence to treatment (Appendix 22).
- Psychological condition.

Frequency depends on the patient’s clinical condition and evolution:

- A visit every week for the first month, every other week for the second month, then once a month if there is no particular problem.
- Additional visits may be required in case of comorbidities, severe or multiple adverse effects, pregnancy, etc.

Visits should coincide with bacteriological examinations and other investigations when possible.

The clinician should take into account any information and concerns regarding treatment tolerance and adherence reported by the patient or the team responsible for the patient’s follow-up and support.

10.6.2 Bacteriological tests

To assess treatment response in patients with:

- PTB: bacteriological tests are essential.
- EPTB: evaluation is based on clinical evolution. However, bacteriological tests are required if patients also develop PTB.

Baseline tests

Baseline tests are those performed on specimens collected just prior to treatment initiation. Baseline tests include:

- RMTs for detection of *M. tuberculosis* and rifampicin, isoniazid and fluoroquinolone resistance.
- Sputum smear microscopy.
- Culture and full phenotypic DST (pDST) or genome sequencing.

For more information see Chapter 3.
If DST results are obtained on a specimen collected more than 2 to 3 weeks prior to treatment initiation, a new specimen should be collected just prior to treatment initiation. The new results are considered as baseline results.

**Follow-up tests**

- Microscopy: once a month until treatment completion. Although less reliable than culture, it provides immediate results which contribute to the assessment of treatment response.
- Culture: once a month until treatment completion. Culture conversion and reversion are useful markers of whether the treatment is effective or not.
- Full pDST (or genome sequencing): if positive culture at Month 4 or later.
- RMTs: Xpert MTB/XDR (or GenoType MTBDRsl if Xpert MTB/XDR is not available) if positive microscopy at Month 4 or later, as it can detect resistance-conferring mutations not present at baseline ([Chapter 3](#)).

**End-of-treatment tests**

Culture and microscopy should be performed at end of treatment, to confirm the end-of-treatment outcome ([Chapter 17](#)).

**Post-treatment tests**

For patients on BPaLM or BPaL regimen, culture and microscopy should be performed 6 and 12-month post-treatment completion, to detect a relapse. For other regimens, post-treatment tests should be performed for operational research purposes only.

### 10.6.3 Other investigations

**Radiography**

At baseline, then every 6 months:

- chest x-ray for patients with PTB,
- bone x-ray for patients with osteoarticular and spinal TB.

**Electrocardiogram**

Some TB drugs cause prolongation of the QT interval, which increases the risk of a potentially life-threatening ventricular arrhythmia, including *torsade de pointes* (TdP)[1].

To monitor the QT interval, electrocardiogram (ECG) should be performed:

- At baseline in all patients taking QT-prolonging TB drugs

Then:

- Once a month in patients:
  - taking < 2 moderate or strong QT-prolonging TB drugs,
  - taking < 3 QT-prolonging drugs (TB and non-TB),
- Once a week for one month, then once a month in patients:
  - taking ≥ 2 moderate or strong QT-prolonging TB drugs,
  - taking ≥ 3 QT-prolonging drugs (TB and non-TB),
  - with other risk factors for QT prolongation or TdP:
    - a history of syncopal episodes, TdP or congenital long QT syndrome;
    - uncompensated heart failure, severe coronary disease, bradycardia;
    - untreated hypothyroidism.

Increased ECG monitoring is required in patients in whom a QT prolongation is detected.

For ECG reading see [Appendix 16](#).
For the management of QT prolongation see [Appendix 17](#).
For the list of QT prolonging drugs see [Appendix 19](#).

**Brief peripheral neuropathy screen**

For patients on linezolid: brief peripheral neuropathy screen (BPNS) at baseline, then once a month to detect peripheral neuropathy ([Appendix 16](#)).

**Visual function tests**

...
For patients on drugs with ocular toxicity: visual acuity and colour vision tests (Ishihara test) at baseline, then once a month to detect the first signs of optic neuritis.

**Audiometry**

For patients on aminoglycosides: at baseline, then once a month to detect hearing loss. Monitoring is particularly important in children, as hearing loss in childhood has negative effects on development.

**Full blood count**

For all patients: haemoglobin, red and white blood cells, and platelets at baseline, then if indicated.
For patients on linezolid: every 2 weeks for the first 2 months, then once a month.
For patients on zidovudine (AZT): once a month for the first 2 months, then if indicated.

**Liver function tests**

For all patients: serum levels of aspartate aminotransferase (AST) and alanine aminotransferase (ALT) at baseline, then once a month.
Bilirubin, if AST and ALT are elevated, or if indicated.
Monitor liver function more frequently in case of increase in AST/ALT or other signs of hepatic disorder or risk factors, such as hepatitis B or C.

**Serum creatinine and potassium level**

For all patients: at baseline, then if indicated (e.g. patients with renal insufficiency).
For patients on aminoglycoside: once a month, or more frequently if indicated.

**Creatinine clearance**

For patients with renal insufficiency: at baseline. If < 30 ml/minute, the dose of certain TB drugs should be adjusted ([Appendix 12](#)).

**Glycated haemoglobin (HbA1c) and/or blood glucose level (BGL)**

For all patients: at baseline to detect diabetes. If diabetes is diagnosed, monitor according to standard protocols.

**HIV, hepatitis B and C**

For all patients with undocumented HIV hepatitis B and C status: at baseline; HIV test every 6 months in high HIV prevalence areas. Tests can be repeated in case of recent exposure.

**CD4 and viral load**

For HIV-infected patients: at baseline, then every 6 months.

**Thyroid-stimulating hormone (TSH)**

For patients on thionamides or PAS: at baseline, then every 3 months.
If hypothyroidism is diagnosed: 4 to 12 weeks after levothyroxine initiation and after each levothyroxine dose adjustment until stable, then every 6 months until the end of TB treatment, or for as long as the patient takes levothyroxine.

**Pregnancy test**

For all adolescents and women of childbearing age: at baseline, then if indicated.

**References**

   https://doi.org/10.1056/NEJMra032426
10.7 Adverse effects

Rapid and aggressive treatment of adverse effects is essential to improve tolerance and treatment outcomes. Some adverse effects should be routinely prevented (e.g. peripheral neuropathy).

Most adverse effects cannot be prevented, but can be managed with symptomatic treatment (e.g. arthropathy due to pyrazinamide).

Some adverse effects cannot be eliminated, but are not serious (e.g. skin discoloration due to clofazimine). Patients need reassurance and support to be able to tolerate them until they subside spontaneously.

Some adverse effects cannot be eliminated, but are not serious (e.g. skin discoloration due to clofazimine). Patients need reassurance and support to be able to tolerate them until they subside spontaneously.

Ascertaining which drug is responsible for a particular adverse effect can be challenging. Temporarily stopping a drug, or reducing the dose, can help identify the responsible drug.

Adverse effects can appear at any time during treatment. Patients should be informed that they are likely to experience adverse effects and should report them immediately to health staff. Treatment supporters and nurses should rapidly report adverse effects to the clinician. Only the managing clinician can modify or stop a TB treatment.

For the management of adverse effects see Appendix 17.
10.8 Treatment adaptation and change of treatment

10.8.1 Treatment adaptation

Treatment adaptation may be done by the clinician in case of severe adverse effects (Appendix 17). The following are considered treatment adaptations:

- For BPaLM and BPaL:
  - permanent interruption of linezolid after Month 4, or
  - interruption < 2 consecutive weeks or < 4 nonconsecutive weeks of individual drug(s) or the whole treatment.
- For 9-month bedaquiline-containing regimens:
  - temporary interruption of individual drug(s) or the whole treatment, or
  - permanent interruption of ethambutol or pyrazinamide during the continuation phase.
- For LTRs:
  - temporary interruption of individual drug(s) or the whole treatment, or
  - change of one drug class in the regimen (no more than one).

In an LTR, at least 4-5 likely effective drugs are needed (Box 10.3). If any of these drugs must be permanently stopped:

- during the first 6 months, the regimen should be modified while maintaining the required number of likely effective drugs.
- after the first 6 months, if the patient clinical status has improved and bacteriological tests are negative, the clinician can decide to continue the treatment if it still includes at least 3 drugs from Group A and/or B.

These modifications are considered as treatment adaptations (not treatment changes, see Section 10.8.2) as they do not meet the definition of “treatment failure” (Chapter 17).

10.8.2 Change of treatment

Treatment change is defined as the switch from an STR to an LTR or from an LTR to a newly designed LTR.

Treatment should be changed by the clinician in the following circumstances[^1]:

- Emergence of a new resistance after treatment initiation.
- Resistance not detected at baseline for any reason.
- No bacteriological conversion or bacteriological reversion (Chapter 17).
- Insufficient clinical response to treatment in patients:
  - with no bacteriologically confirmed TB (e.g. miliary TB, some forms of EPTB, children);
  - with bacteriologically confirmed TB when bacteriological response cannot be assessed, or the result is inconclusive.
- Drug interruption due to severe adverse effects[^2]:
  - For BPaLM and BPaL:
    - permanent interruption of bedaquiline or pretomanid, or
    - permanent interruption of linezolid before the end of Month 4, or
    - interruption ≥ 2 consecutive weeks or ≥ 4 nonconsecutive weeks of the whole treatment.
  - For 9-month bedaquiline-containing regimens:
    - permanent interruption of bedaquiline, levofloxacin/moxifloxacin, linezolid, ethionamide or clofazimine, or
    - permanent interruption of both ethambutol and pyrazinamide.
  - For LTRs: change of at least 2 drug classes in the regimen.

Note: for BPaLM regimen, if moxifloxacin must be interrupted, see the note in Section 10.2.1.

Treatment changes meet the outcome definition of “treatment failure” (Chapter 17) except when the reason for change is a resistance not detected at baseline[^1].

References

https://www.who.int/publications/i/item/9789240065116
10.9 Treatment interruptions

Problems of treatment interruption by the patient (e.g. discontinuation of certain drugs, recurrent treatment interruptions) should be detected and addressed (management of adverse effects if necessary and reinforcement of patient support measures).

Interruptions of individual drug(s) or of the whole treatment may lead to the emergence of new resistances. Moreover, in case of treatment interruption, drugs with a long half-life such as bedaquiline or clofazimine remain in the blood for several months. In practice, it is as if the patient is receiving bedaquiline and/or clofazimine alone, which increases the risk of developing resistance to these drugs.

Patients who have interrupted the whole treatment for 2 months or more meet the definition of patients “lost to follow-up” (Chapter 17). If the patient returns, repeat bacteriological tests (RMTs, culture and full pDST or genome sequencing) to detect potential new resistance; start a new individualized regimen.

For patients who have interrupted the whole treatment for 4 weeks or more but less than 2 months, perform new bacteriological tests as above. Further treatment depends on the results of the RMTs (pending full bacteriological tests results) and the patient’s clinical status: new individualized regimen or continuation of the same regimen with catch-up of doses missed during interruptions to complete treatment.

For patients on BPaLM/BPaL who have interrupted the whole treatment for 2 weeks or more, perform new bacteriological tests as above and start a new individualized regimen.
10.10 Surgery

Surgery is an adjunct to the pharmacological treatment of MDR/RR-TB patients. It can be performed only by trained thoracic surgeons, in specialized surgical units with excellent postoperative care. These units must implement strict infection prevention and control measures because thoracic surgery, mechanical ventilation and post-operative physiotherapy generate large quantities of aerosols.

When access to surgery is limited, it should be considered in priority for patients with resistance to a large number of drugs and localized lung damage.

Surgery can be performed early, when the disease is still localized (e.g. to a lobe). Partial lung resection (lobectomy or wedge resection) can be effective and safe if performed under appropriate conditions.

At the beginning of treatment, there is a window of opportunity during which the bacillary load decreases transiently under the pressure of TB drugs (decrease in mycobacteria in smears and/or culture). This window is the optimal time for surgery. The prognosis is better when resection is performed after culture conversion.

It is recommended to perform culture and DST of the resection material. Depending on the results, modification of treatment may be required.

References


10.11 Treatment failure and palliative care

When a treatment is failing, treatment outcome should be recorded as “failure” (Chapter 17).

A new baseline specimen should be collected, and a new individualized regimen designed according to the principles described in Section 10.2.2.

When the minimum number of likely effective drugs cannot be reached, the use of TB drugs under development available for compassionate use is encouraged (Appendix 18).

When no therapeutic option or new regimen is possible, the patient can continue a TB regimen that is reasonably tolerated, or the regimen can be stopped. The decision to stop treatment should be made after careful evaluation and consultation with the patient, family, and TB treatment team. Palliative and supportive care should be continued.

Palliative and supportive care is an integral part of patient care throughout their illness[1][2]. Some care should be continued after cure if the patient remains with significant respiratory damage. Palliative and supportive include[8]:

- Relief of respiratory symptoms: oxygen for shortness of breath; corticosteroids (prednisolone) for severe respiratory failure; codeine to help control cough.
- Use of all necessary ancillary drugs.
- Nutritional support for undernourished patients.
- Care to improve comfort and prevent complications in debilitated patients; regular position changes in bedridden patients to prevent bedsores; bathing and oral hygiene to improve patient comfort and prevent skin infections.
- Management of anxiety or depression (due to prolonged illness, separation from family, difficult living conditions, etc.); support to family as needed.

Offer home care to families who need help. Reserve inpatient rooms for end-of-life patients if they cannot be cared for at home.

References


Chapter 11: Treatment of mono- and poly-drug resistant tuberculosis (PDR-TB)

11.1 Treatment schemes

11.2 Treatment algorithms for PDR-TB

Update: January 2014
11.1 Treatment schemes

11.1.1 Choice of the treatment scheme

Mono- and poly-drug resistant tuberculosis (PDR-TB) management is based on the PDR treatment schemes presented in Table 11.1.

Table 11.1 - Resistance pattern and recommended treatment schemes
Sus. = susceptible; Res. = resistant.

<table>
<thead>
<tr>
<th>Resistance category</th>
<th>H</th>
<th>R</th>
<th>E</th>
<th>S</th>
<th>Treatment scheme</th>
</tr>
</thead>
<tbody>
<tr>
<td>H-resistance</td>
<td>Res.</td>
<td>Sus.</td>
<td>Sus.</td>
<td>Sus.</td>
<td>PDR Scheme A(^{(a)})</td>
</tr>
<tr>
<td></td>
<td>Res.</td>
<td>Sus.</td>
<td>Sus.</td>
<td>Res.</td>
<td>PDR Scheme A(^{(a)})</td>
</tr>
<tr>
<td></td>
<td>Res.</td>
<td>Sus.</td>
<td>Res.</td>
<td>Sus.</td>
<td>PDR Scheme B</td>
</tr>
<tr>
<td></td>
<td>Res.</td>
<td>Sus.</td>
<td>Res.</td>
<td>Res.</td>
<td>PDR Scheme B</td>
</tr>
<tr>
<td>R-resistance</td>
<td>Sus.</td>
<td>Res.</td>
<td>Sus.</td>
<td>Sus.</td>
<td>PDR Scheme C</td>
</tr>
<tr>
<td></td>
<td>Sus.</td>
<td>Res.</td>
<td>Sus.</td>
<td>Res.</td>
<td>PDR Scheme C</td>
</tr>
<tr>
<td></td>
<td>Sus.</td>
<td>Res.</td>
<td>Res.</td>
<td>Sus.</td>
<td>PDR Scheme C</td>
</tr>
</tbody>
</table>

\(^{(a)}\) Except previously treated patients, for whom PDR Scheme B + ethambutol is preferred.

The treatment schemes for mono/PDR-TB are based on the assumption that a full baseline drug susceptibility testing (DST) is performed before or at the start of treatment with first line anti-TB drugs.

There is little published evidence to determine the best treatment for mono/PDR-TB. The treatment schemes are therefore based on the principles of TB treatment and expert opinion\(^{[1][2][3]}\).

At least 3, ideally 4, likely effective drugs are included in the regimen. DST results at baseline and previous treatment history are used to choose the appropriate scheme.

The use of Xpert MTB/RIF can greatly aid in getting patients on the proper regimens when isoniazid resistance is present and amplification of resistance to rifampicin is a possibility.

Perform second-line DST if patients come from a region of high second-line resistance and if there is a history of second-line anti-TB drug use. Resistance to second-line anti-TB drugs will impact the choice of regimen.

11.1.2 PDR Scheme A for cases with H or HS resistance
For new patients, the treatment regimen is 9 RZE. However, the combination HRZE can be used if more convenient since it can be given as fixed-dose combination.

At Month 2, perform smear, Xpert MTB/RIF, and culture:

<table>
<thead>
<tr>
<th>Xpert available</th>
<th>Xpert RIF+: switch to empiric MDR regimen while waiting for full DST results then, adapt treatment accordingly.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Xpert RIF−: continue PDR Scheme A.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Xpert not available</th>
<th>Culture+: switch to empiric MDR regimen with the inclusion of R while waiting for full DST results.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>• DST is unchanged (H or HS resistance only): stop the MDR regimen, and resume PDR Scheme A;</td>
</tr>
<tr>
<td></td>
<td>• DST has changed: adapt treatment accordingly.</td>
</tr>
<tr>
<td></td>
<td>Culture−: continue PDR Scheme A.</td>
</tr>
</tbody>
</table>

Perform smear and culture every other month. If cultures or smears are positive, switch to MDR regimen while waiting for full DST results then, adapt treatment accordingly.

For previously treated patients, it is safer to use Scheme B plus ethambutol, as DST to this drug should not be relied upon if the patient has already received it.

### 11.1.3 PDR Scheme B for cases with HE or HES resistance

Start patients on 3 Cm (or Km)-Lfx-RZ/7 Lfx-RZ regardless of smear status at the time of diagnosis.

At Month 2, perform smear, Xpert MTB/RIF and culture:

<table>
<thead>
<tr>
<th>Xpert available</th>
<th>Xpert RIF+: switch to empiric MDR regimen while waiting for full DST results then, adapt treatment accordingly.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Xpert RIF−: continue PDR Scheme B.</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Xpert not available</th>
<th>Culture+: switch to empiric MDR regimen with the inclusion of R while waiting for full DST results.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>• DST is unchanged (HE or HES resistance only): stop the MDR regimen, and resume PDR Scheme B;</td>
</tr>
<tr>
<td></td>
<td>• DST has changed: adapt treatment accordingly.</td>
</tr>
<tr>
<td></td>
<td>Culture−: continue PDR Scheme B.</td>
</tr>
</tbody>
</table>

At Month 3, perform smear, Xpert MTB/RIF, and culture. If Xpert shows RIF+ or if the culture is still positive, this regimen is declared “failure”. Switch to MDR treatment.

Even if found susceptible, streptomycin should not be used given the high rates of resistance to this drug in patients with DR-TB and the poor reliability of the DST.

### 11.1.4 PDR Scheme C for cases with R or RS or RE or RES resistance

Start patients on MDR regimen until confirmation that the strain is susceptible to fluoroquinolones and injectable agents.

When DST results confirm resistance to R, RS, RE or RES and susceptibility to H, fluoroquinolones and an injectable agent, there are two options:

1 - Continue the full course of MDR-TB treatment plus isoniazid. This is a reasonable consideration given that DST reliability is not 100%. This is recommended if the suspicion for MDR-TB is high (i.e. a contact of an MDR-TB patient or failure of a first-line regimen).

2 - Start PDR Scheme C: 3 Cm (or Km)-Lfx-HZ (+/- E)/12 Lfx-HZ (+/- E). Ethambutol is added if it is likely to be effective.

Even if found susceptible, streptomycin should not be used given the high rates of resistance to this drug in patients with DR-TB and the poor reliability of the DST.
At Month 2, perform smear and culture:

**Culture+**: start empiric MDR regimen and repeat DST.
- DST is unchanged: resume PDR Scheme C;
- DST has changed: adapt treatment accordingly.

**Culture−**: complete PDR Scheme C.

At Month 3, perform smear and culture. If the culture is still positive, this regimen is declared “failure.” Switch to MDR treatment.

**Note**: if the baseline DST is performed by LPA (Hain® test), only DST for R and H are available. In order to avoid possible resistance amplification, the worst scenario should be assumed:
- If only resistance to H is detected, treat with Scheme B, even new patients while waiting for full DST.
- If only resistance to R is detected, treat as MDR-TB as sensitivity of Hain® test for H resistance is low.

**References**


11.2 Treatment algorithms for PDR-TB

**PDR scheme A**

Note: for previously treated patients it is safer to use Scheme B + ethambutol.

1. Continue the initial regimen (new or previously treated case)
2. At Month 2: Xpert Rif + or culture+
   - **Yes**: Start continuation phase: 7 RZE. Perform smear and culture every other month.
   - **No**: Any smear+/culture+
     - **Yes**: Start empiric MDR-TB treatment while waiting for DST result.
     - **No**: Cured or treatment completed

**PDR scheme B**
PDR SCHEME C
R, RE (+/- S) resistance

Start empiric MDR-TB treatment while waiting for DST result confirming susceptibility to H, FQs and injectable.

Start adapted treatment
3 Cm (or Km) 1fx-HZE
Perform smear and culture

At Month 2:
culture+

Yes
Resume empiric MDR-TB treatment while waiting for DST result.

No
Complete intensive phase

- If DST unchanged: complete continuation phase to a total of 12 1fx-HZE after culture negativisation. Perform smear and culture every other month.
- If DST changed: adapt treatment accordingly.

At Month 3:
smear+ / culture+

Yes
Start continuation phase: 12 1fx-HZE
Perform smear and culture every other month.

No

Any smear+ / culture+

Yes
Failure
Resume MDR-TB treatment.

No
Cured or treatment completed
Chapter 12: Tuberculosis and HIV co-infection

12.1 HIV counselling and testing
12.2 Concomitant treatment of tuberculosis and HIV co-infection
12.3 Interactions and overlapping toxicities between tuberculosis drugs and antiretrovirals
12.4 Prevention of opportunistic infections
12.5 Immune reconstitution inflammatory syndrome
12.6 Patient monitoring

Update: March 2023
12.1 HIV counselling and testing

When HIV status is unknown, HIV counselling and testing is recommended for patients with latent tuberculosis infection (LTBI) and patients with presumed or confirmed active tuberculosis (TB).

The HIV test is performed after counselling, unless the person explicitly declines to be tested.
12.2 Concomitant treatment of tuberculosis and HIV co-infection

12.2.1 Active tuberculosis

For all HIV-infected patients, treatment of active TB should be started first. Then, antiretroviral therapy (ART) should be initiated within 2 weeks of starting treatment of active TB, except for patients with TB meningitis. For patients with TB meningitis, early initiation of ART is associated with an increased risk of serious adverse events. It is therefore recommended to start ART 4 to 8 weeks after the start of TB treatment.

Table 12.1 - First-line ART for patients with active TB and HIV co-infection

<table>
<thead>
<tr>
<th>Patients</th>
<th>First choice</th>
<th>Main alternatives</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neonates</td>
<td>AZT + 3TC + RAL((a))</td>
<td>AZT + 3TC + LPV/r((b))((c))</td>
</tr>
<tr>
<td>Children</td>
<td>ABC + 3TC + DTG((a))</td>
<td>If paediatric DTG not available: ABC + 3TC + LPV/r((b)) ABC + 3TC + RAL((c))</td>
</tr>
<tr>
<td>Adolescents and adults</td>
<td>TDF + 3TC (or FTC) + DTG</td>
<td>TDF + 3TC (or FTC) + EFV ABC + 3TC + DTG AZT + 3TC + EFV</td>
</tr>
</tbody>
</table>

<p>| | | |</p>
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Childbearing-aged and pregnant women</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

(a) Doses of DTG and RAL should be doubled in patients taking rifampicin.
(b) LPV/r paediatric formulation can be administered to children as of the age of 2 weeks.
(c) LPV/r should not be used in children taking bedaquiline. The dose of LPV/r should be adjusted in neonates and children taking rifampicin.
(d) RAL should be used only if LPV/r paediatric formulation is not available.

12.2.2 Latent tuberculosis infection

For patients with LTBI not yet on ART, the initiation of ART should take priority over the initiation of LTBI treatment. See Chapter 16.

References

   https://apps.who.int/iris/rest/bitstreams/1336192/retrieve

   https://apps.who.int/iris/rest/bitstreams/1238289/retrieve
12.3 Interactions and overlapping toxicities between tuberculosis drugs and antiretrovirals

Certain combinations of TB drugs and ARVs are contraindicated or should be avoided or require dose adjustments of TB drugs or ARVs. For more information see Appendix 19.

Note: drug interactions and overlapping toxicities between TB drugs and drugs other than ARVs are common. For example, rifampicin reduces plasma concentrations of fluconazole by 25%. It may be necessary to increase the dose of fluconazole. Conversely, fluconazole increases plasma concentrations of rifabutin. It is necessary to monitor for signs of rifabutin toxicity. If patients are taking drugs other than ARVs, clinicians should be aware of potential interactions and overlapping toxicities.

References

12.4 Prevention of opportunistic infections

During TB treatment, cotrimoxazole preventive therapy (CPT) should be started or continued in order to prevent common and opportunistic infections.
12.5 Immune reconstitution inflammatory syndrome

TB-associated immune reconstitution inflammatory syndrome (TB-IRIS) can occur in a patient on antiretroviral and/or TB treatment. It is characterised by the onset of new or worsening (after initial improvement) signs and symptoms of TB resulting from the restoration of the immune system by ART.

Most common signs and symptoms of TB-IRIS are fever, lymphadenopathy, pulmonary infiltrates, pleural effusion, respiratory distress, neurological signs[1].

TB-IRIS occurs in two circumstances:
- Paradoxical TB-IRIS: the diagnosis of active TB is made, the patient starts TB treatment, followed by ART and then signs and symptoms of TB worsen.
- Unmasking TB-IRIS: TB is not detected, the patient starts ART and then develops signs and symptoms of TB.

TB-IRIS is more common in patients with low CD4 count. It usually occurs within 3 months of starting ART, most often within the first month[2].

The following differential diagnoses should be considered before making the diagnosis of TB-IRIS:
- New onset of opportunistic infection.
- Other infections unmasked after immune reconstitution due to ART.
- Failure of TB treatment due to drug resistance.

TB-IRIS is considered severe in patients with neurological signs, respiratory distress, or if their condition requires hospitalisation or frequent ambulatory care.

Treatment of severe TB-IRIS is based on corticosteroids, except in the case of Kaposi’s sarcoma or cryptococcal meningitis, for which corticosteroids are contraindicated.

Patients on corticosteroids should be monitored to detect any other opportunistic infections.

In patients with non-severe TB-IRIS, treatment is based on non-steroidal anti-inflammatory drugs.

In case of unmasking TB-IRIS, TB treatment should be started immediately.

ART should not be interrupted, except in case of life-threatening IRIS.

Table 12.2 - Symptomatic treatment of TB-IRIS

<table>
<thead>
<tr>
<th>TB-IRIS</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Severe</td>
<td>prednisolone PO</td>
</tr>
<tr>
<td></td>
<td>Child and adult: 1.5 mg/kg once daily (2 weeks) then 0.75 mg/kg once daily (2 weeks)[3]</td>
</tr>
<tr>
<td>Non-severe</td>
<td>ibuprofen PO for the shortest possible duration</td>
</tr>
<tr>
<td></td>
<td>Child over 3 months: 5 to 10 mg/kg 3 to 4 times daily (max. 30 mg/kg daily)</td>
</tr>
<tr>
<td></td>
<td>Child 12 years and over and adult: 200 to 400 mg 3 to 4 times daily (max. 1200 mg daily)</td>
</tr>
</tbody>
</table>

References


12.6 Patient monitoring

For patients on drug susceptible TB treatment see Chapter 9.
For patients on multidrug-resistant or rifampicin-resistant TB treatment see Chapter 10.
For patients on isoniazid-resistant TB treatment see Chapter 11.
Chapter 13: Adherence to tuberculosis treatment

13.1 Introduction

13.2 Treatment delivery model

13.3 Factors that influence adherence

13.4 Patient education and support

Update: January 2022
13.1 Introduction

Good adherence is when the patient follows the treatment as prescribed. Patient understanding, acceptance and motivation to start and complete TB treatment are essential to maximise chances of cure. Good knowledge of drug dosing, length of treatment, required clinical follow-up and common adverse effects help patients to follow the prescribed therapy.

Failure to take tuberculosis (TB) drugs consistently, or in an inappropriate manner, or stopping the treatment too soon, can lead to treatment failure or relapse. It may also contribute to the development of resistance, which can complicate subsequent treatment, thereby decreasing the chances of a successful outcome.
13.2 Treatment delivery model

13.2.1 Self-administered treatment

Self-administered treatment (SAT) is taken autonomously by the patient without daily supervision. The patient is seen at a health facility at regular intervals (e.g. monthly) to receive drugs, support and treatment education. SMS telephone reminders may be considered to reinforce adherence.

13.2.2 Directly observed therapy

Drugs are sometimes provided daily to the patient and the treatment is taken under direct observation (DOT) by a third party.

DOT may be provided:
- In health facilities (facility-based DOT): in this model, DOT is implemented in a centralised setting and treatment is administered by healthcare workers.
- Outside of health facilities (community or home-based DOT): in this model, DOT is implemented in a decentralised setting and is usually provided by supervised, trained and remunerated treatment supporters.

For the roles and responsibilities of treatment supporters see Appendix 20.

- Remotely (video-observed therapy or VOT): VOT uses secure Internet connections via a smart phone or computer application to remotely supervise patients taking their treatment.

DOT is labour-intensive to implement and can be inconvenient for patients. Community and home-based DOT and VOT require fewer resources (personnel and transport) than facility-based DOT and may be more convenient for patients.

Box 13.1 – Recommended treatment delivery models

**Drug-susceptible TB (DS-TB)**
- DOT has not been proven to improve treatment outcomes for DS-TB when compared to SAT in controlled trials[1].
- When there is no factor to complicate adherence, and provided the patient receives appropriate support, treatment should be self-administered.
- There are some situations in which DOT may be preferred:
  - Patients with mental health issues or serious socioeconomic problems (e.g. the homeless) and all patients incapable of taking drugs on their own.
  - Prisoners (risk of drugs being sold or stolen).

**Drug-resistant TB (DR-TB)**
- Due to the lack of fixed-dose combinations (FDC), length of treatment, adverse effects of TB drugs and lack of therapeutic alternatives if treatment fails, patients usually require reinforced support.
- If DOT is considered useful, home-based DOT[2] or VOT are preferred to facility-based DOT. A combination of approaches may be required for some patients.

**Latent TB infection (LTBI)**
- LTBI treatments can be self-administered.
- DOT may be preferred with the 3HP regimen, as it may cause serious hypersensitivity reactions. However, SAT can be considered if the patient well informed and is able to seek rapid medical attention if adverse effects develop.

References


13.3 Factors that influence adherence

Several factors can influence adherence, including barriers related to the patient, the treatment or the therapeutic environment. While it is not always feasible to address all these factors, at the very least it is possible to control the treatment and therapeutic environment-related factors.

13.3.1 Patient-related factors

A discussion should be held with the patient prior to treatment initiation and then during every contact they have with the healthcare team. The objective is to identify and anticipate barriers to treatment adherence. Barriers may include:

- Socioeconomic factors (work and home responsibilities, treatment-related costs, decreased income, etc.).
- Psychological factors (feelings of shame, fear of stigma or marginalisation, uncertainty about the future, conceptions about the disease and its treatment, etc.).
- Physical or mental disability.
- Lack of knowledge about the disease and treatment.
- Perception of the disease and treatment (a patient might abandon treatment due to improvement or absence of improvement, a negative experience with a previous treatment, etc.).

Solutions depend on the context and the patient’s problem, and therefore should be identified on a case-by-case basis.

13.3.2 Treatment-related factors

- Simplicity of treatment improves adherence. The use of FDC simplifies the treatment by reducing the number of tablets. In addition, FDC prevents omission of one or more prescribed TB drugs.
- Adverse effects may lead patients to interrupt their treatment, so these should be detected and managed promptly.

13.3.3 Factors related to the therapeutic environment

- To ensure the widest possible access to treatment, TB diagnosis, monitoring and treatment (including TB drugs and drugs for adverse effects and co-morbidities) should be provided free of charge.
- The relationship between patients and healthcare workers influences if patients have confidence in healthcare workers, they are more likely to follow recommendations and engage with the treatment process. Patients are also more likely to bring questions and concerns to the attention of healthcare workers. The same applies to the relationship with treatment supporters.
- In health facilities, the way in which patients are received is Waiting times for diagnosis or follow-up visits should be reasonable.
- Drug supply management must be rigorous. Shortages can lead to treatment interruption and negatively impact adherence (patients waste time in unnecessary travel and lose confidence in the health facility, ).
- The proximity of drug distribution sites limits the number of patients who abandon due to transportation problems. To anticipate potential problems, give the patients a few extra days of treatment in case they are unable to come to get their drugs on the scheduled
- For the co-management of TB and HIV infection, patients should receive TB and HIV treatment at the same time and in the same place (“one-stop service”). This reduces the number of visits and decreases waiting times, which results in greater patient satisfaction and improved treatment outcomes. Co-management of other co-morbidities (e.g. diabetes, hypertension) should, when possible, use the same approach.
- Hospitalisation should be limited to patients with clinical conditions requiring hospital level care. If hospitalisation is necessary, accommodation (comfort, food, heating, etc.) should be adequate. The duration of stay should be as short as possible and patients should be discharged as soon as their clinical condition allows.
13.4 Patient education and support

Patient education and support require the involvement of the entire healthcare team (clinicians, nurses, treatment supporters, social workers, etc.). In large-scale programmes, the healthcare team sometimes includes trained counsellors who provide information and support.

Treatment education and support may be provided through various channels: organising educational sessions during in-facility or home visits, video and telephone contacts.

Patient education and support are required throughout treatment, as adherence may vary over time and patients may experience phases of treatment acceptance and rejection.

Due to the toxicity and long duration of treatment, patients on DR-TB treatment usually require substantial support.

13.4.1 Patient education

Patient education consists of:
- Helping patients to understand the disease and treatment.
- Enabling patients to acquire and maintain skills that allow them to manage their treatment and disease in their everyday lives.
- Answering patients’ questions throughout the treatment.

For more information see Appendix 21.

13.4.2 Emotional support

Listen to patients and give them encouragement, so that they feel comfortable saying they have forgotten or have made a mistake with their treatment. This is common, and it is important to know so that solutions can be found.

Psychological problems, such as depression and anxiety are frequent, and may have a negative impact on adherence. The healthcare team should be sensitised to their early detection and management.

13.4.3 Social support

Implement social support measures for patients with limited resources. Depending on the situation and specific needs of patients:
- Social workers can help to obtain disability allowances, housing assistance, shelter for the homeless, etc.
- The programme can provide meals or food, vouchers or money for transportation or reimburse the cost, etc.
Chapter 14: Tuberculosis infection control

14.1 Introduction
14.2 Implementation of TB IC strategies
14.3 Administrative controls
14.4 Environmental controls
14.5 Personal protective measures
14.6 Hospital hygiene
14.7 Patients’ homes

Update: January 2014
14.1 Introduction

The largest source of *M. tuberculosis* transmission is the contagious patients with respiratory tuberculosis not yet diagnosed and put on treatment. Therefore, tuberculosis infection control (TB IC) relies, above all, on:

- Early diagnosis (including in clinics and any non-tuberculosis medical wards, whereby active case finding through cough surveillance of all admissions should avoid days or weeks of transmission from unsuspected TB cases);

AND

- Prompt implementation of effective treatment. With effective treatment, contagiousness decreases even after a few days and may be considered nil after 2 to 3 weeks of treatment.[1][2][3][4]. It is essential the treatment is “effective,” as multidrug-resistant TB (MDR-TB) patients that are placed on first-line anti-TB drugs are likely to remain contagious.

However, in health care facilities where TB patients or persons suspected of having TB congregate, additional measures are needed to reduce the risk of transmission between patients, to health care staff and to vulnerable (particularly immunocompromised) patients/visitors.[5].

TB infection control (IC)* consists in different strategies for preventing transmission of TB in health care facilities.

Footnotes

(a) This chapter reviews the basic TB IC strategies. More in depth information can be found from the Tuberculosis Coalition for Technical Assistance which has published a framework and developed a website (http://www.tbcta.org/library) that provides a comprehensive set of examples.

References


14.2 Implementation of TB IC strategies

There is a trio of infection control levels, which include (1) administrative, (2) environmental and (3) personal protective controls. The implementation of these measures requires a dedicated staff and an IC plan.

14.2.1 Infection control practitioner

A person should be clearly identified and designated as responsible for TB IC. This person should have the support and authority to conduct, apply and evaluate TB IC policies. This person in some settings is also known as IC officer.

14.2.2 Infection control committee

The IC practitioner would evaluate the need to create an infection control committee (ICC). The ICC might include doctors, nurses, laboratory technicians, logisticians and administration staff (including representation from the maintenance and housekeeping services). According to the context and degree of risk, experts in IC may be needed.

14.2.3 Infection control plan

All facilities should have a detailed written IC plan that is at least annually updated and distributed to healthcare staff. A simplified version of the plan must be accessible to all healthcare workers including staff not directly involved in TB patients’ management, such as cleaners, kitchen staff, etc.

The first step in developing an IC plan is assessing the health care facility’s risk for TB transmission. This should be performed by the IC practitioner. The plan must be specific to each facility. An example of risk assessment tool is given in Appendix 16.

The IC plan should include the different types of measures—administrative, environmental and personal. Information on specific precautions and procedures for high-risk areas should be detailed.

It is recommended to draw a floor plan of the facility with the different areas, including the patient flow and identifying areas of high risk.

Listed below from highest to lowest level of risk:

<table>
<thead>
<tr>
<th>Highest risk</th>
</tr>
</thead>
<tbody>
<tr>
<td>• Smear-positive inpatient unit</td>
</tr>
<tr>
<td>• Diagnosis department</td>
</tr>
<tr>
<td>• Culture/drug susceptibility test (DST) and sputum smear preparation area (laboratory)</td>
</tr>
<tr>
<td>• Sputum collection area</td>
</tr>
<tr>
<td>• Radiology department</td>
</tr>
<tr>
<td>• Waiting area</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Limited risk</th>
</tr>
</thead>
<tbody>
<tr>
<td>• Children inpatient ward</td>
</tr>
<tr>
<td>• Extrapulmonary TB (EPTB) and smear-negative unit</td>
</tr>
<tr>
<td>• Sputum reception and smear reading area (laboratory)</td>
</tr>
<tr>
<td>• Waste management area</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Lowest risk (non-TB zone)</th>
</tr>
</thead>
<tbody>
<tr>
<td>• Kitchen area</td>
</tr>
<tr>
<td>• Administration</td>
</tr>
</tbody>
</table>
References

   https://apps.who.int/iris/bitstream/handle/10665/44148/9789241598323_eng.pdf?sequence=1

2. Implementing the WHO policy on TB infection control. Tuberculosis Coalition for Technical Assistance.  
14.3 Administrative controls

The administrative controls aim at preventing the exposure to infectious droplet nuclei.

14.3.1 Patients triage

Upon entry into the health facility, a member of the medical staff should identify patients with a cough as soon as possible. Patients with a cough over two weeks should be sent to a separate waiting room if possible.

All patients with cough (including patients with less than two weeks of cough) should receive tissues or face masks, and they should be requested to cover their mouth and nose when they cough.

14.3.2 Patient, visitors and attendants' flow

Inside the TB department, circulation of patients and attendants is controlled:

- Encourage patients/attendants to spend as much time as possible outdoors if weather permits or in areas that are open on three or four sides.
- Have visible signage on entry doors to TB wards that forbid visitors to enter.
- Limit visitation duration, particularly for contagious patients.
- Encourage visits outside the building, especially for contagious patients.
- Have visiting areas well identified with signage.
- Before any visit, the nurse should provide information on transmission risk, including the usage of respirators if carers need to go in high risk areas, such as smear-positive, drug-resistant TB (DR-TB), re-treatment smear-positive inpatient units and areas or clinics were diagnosis of TB is being undertaken.
- Avoid that known or suspect TB patients go through areas where they may infect other patients, and vice versa, that patients without TB go through areas where they are unnecessarily exposed to the bacillus.

14.3.3 Segregation of hospitalized patients

Patients should preferably be treated in ambulatory care. Hospitalisation should be limited and reserved for clinically unwell patients. TB wards must be separated from the others wards in the health structure compound.

Ideally, within the TB department, patients should be placed in single rooms. If this is not possible, cohort isolation must be implemented and different sections should be labelled according to the degree of contagiousness (smear/culture status) and risk of resistance.

The following is one scheme of separation. It does involve the use of some single isolation rooms (all TB inpatient facilities should have some isolation rooms. If none exist, a very high priority is to add some).

- Smear-positive patients with proven or suspected DR-TB, including chronic cases and retreatment cases that are likely to have MDR-TB. MDR-TB cases should have single isolation rooms (place in 2 to 4 person rooms with other MDR-TB patients if there are no single rooms and try to match DST patterns). It is particularly important not to mix MDR-TB patients with extensively drug-resistant TB (XDR-TB) patients.
- Smear-positive patients with fully susceptible TB.
- Smear-negative patients (or patients who have converted), with proven or suspected DRTB (once patients are on effective treatment, they rapidly become non-contagious).
- Less or non-contagious TB: patients with smear-negative pulmonary TB (PTB), EPTB, patients having converted their sputum/culture and most children.
- Patients who are undergoing diagnosis as suspected cases: when possible do not hospitalize patients for diagnosis. If hospitalization is necessary, these patients need isolation rooms. Never put a patient who is not receiving TB medications in a TB ward.

If women and men are to be separated, this scheme requires at least 8 different wards and enough single rooms for suspect cases and MDR-TB patients.

14.3.4 TB IC training
All healthcare personnel should receive initial training on TB transmission, information on high-risk areas in the facility and on protective measures. Continuing education should be offered annually.

The training should also include how staff can teach patients, visitors and attendants about the risk of TB transmission and how to avoid it (cough etiquette, use of masks and respirators).
14.4 Environmental controls

The environmental measures aim at reducing the concentration of infectious droplet nuclei in the air.

14.4.1 Ventilation

Ventilation (replacement of inside air with outside air) is the most effective means for reducing the concentration of *M. tuberculosis* in the air, and as a result, the risk of transmission. The WHO recommends that in areas where TB transmission might occur, a minimum ventilation rate of 12 air changes per hour (ACH)\(^1\) should be achieved. See Appendix 17 for recommendations on ACH measurement. Effective ventilation can be obtained by natural (assisted or not) or mechanical means.

Natural ventilation

Natural ventilation, especially cross-ventilation (windows/doors in opposite sides of the room), has the best cost-effective ratio. It should be done with the windows and outside doors open (as much as weather conditions permit). Inside doors should be closed so that the flow of air is directed outside and not toward the corridors. Create shady spaces so that patients, attendants and visitors can stay outside during the day. Wind-driven roof turbines (whirly birds) or chimneys can also be used to improve natural ventilation, in that they can keep the principle of directing room air towards the exterior. In addition, fans can be used when the natural ventilation flow rate is too low (assisted natural ventilation).

Mechanical ventilation

When natural ventilation cannot reach adequate rates, centralised mechanical ventilation should be considered in some settings, such as within cold climates. Centralised mechanical ventilation relies on the use of mechanical equipment to maintain an air pressure difference between two areas in order to draw air into a room and vent it to the outside. It requires continuous and meticulous maintenance, which renders it costly and difficult to implement and operate.

Advantages and disadvantages of each ventilation technique are presented in Appendix 18.

14.4.2 Architectural considerations

Airborne infection control should be always considered during the planning/construction stages of new health facilities and those being modified. It is important to achieve the following:

- Building layout and design with maximised natural ventilation (assisted or not) and sunlight. Waiting areas should be open on three sides. Design of TB wards should avoid internal hallways with doors from the rooms and wards opening into them. Instead, doors should open to outside hallways that are open to air (this may not be feasible in cold climates).
- Specific areas (open air, sputum collection booth, etc.) should be reserved for procedures with a high risk of *M. tuberculosis* transmission (e.g. sputum collection, sputum induction, etc.).
- Allow patient flow that reduces exposure of patients at risk to patients that are infectious (e.g. separate waiting rooms for different cohorts, one patient per room in a hospital). If designing a new TB ward, incorporate plenty of single rooms or at least small rooms with 2 to 4 beds for easier separation of the different cohorts of patients. General hospitals should also have isolation rooms available for TB suspects and contagious patients.

Rehabilitation of existing structures in order to maximise natural ventilation could be a viable economical option instead of building expensive systems, like centralised mechanical ventilation.

14.4.3 Ultra-violet germicidal irradiation

Ultra-violet germicidal irradiation (UVGI) lamps\(^4\) may be used when adequate ventilation cannot be achieved in high-risk areas. When properly installed, designed, maintained and operated, an UVGI system, in addition to 6-12 ACH ventilation, could be the equivalent of 10-25 ACH\(^7\). For technical information on upper room UVGI, see Appendix 19.

- Main requirements and constraints in UV lamps usage include:
14.4.4 Areas requiring specific measures

Sputum collection areas

These areas must be settled, wherever possible, outside in open air where bacilli will naturally be dispersed by wind rather than in a closed room where the concentration of bacilli will be high.

In cold regions, sputum collection should be performed in very well ventilated indoor rooms (at least 20 ACH) or in well ventilated rooms (at least 12 ACH) equipped with a UVGI system.

Another option for sputum collection areas in cold climate regions is to assign a specific room of small size (1 m²) with one single glass door opening outside. Keep the door largely open for 5 minutes between each patient. The small volume of air in this room facilitates rapid ventilation.

Laboratory

All laboratories should undergo a risk assessment, and IC measures should be adapted accordingly. In any case, limit the access to all TB laboratories.

The use of ventilated workstation (Appendix 7) is strongly recommended for smear preparation (microscopy and test Xpert). In laboratories where culture are carried out, biological safety cabinets type II must be used.

Laboratories must have easy to clean working surfaces (avoid wood) to allow proper disinfection. They should also have large windows to let in sunlight and allow natural ventilation if the laboratory has no mechanical ventilation.

Water-filters should be used to avoid contamination by saprophyte mycobacteria that are sometimes present in the water.

Footnotes

(a) UVGI inactivate bacilli. Natural light dries the droplet but does not inactivate bacilli.

References


14.5 Personal protective measures

Personal protective measures aim at minimising the risk of bacillus transmission by providing barriers to inhaling or exhaling infectious droplet nuclei.

14.5.1 Respirators (or high-filtration masks or anti-inhalation masks)

A respirator is personal protective equipment that prevents inhalation of infectious droplet nuclei by the person who wears it.

Exposed staff

Staff must wear a respirator, regardless if they are the caregiver or not. Respirators should be worn:

- When in contact with contagious patients (suspect or confirmed TB case);
- When collecting sputum samples;
- When collecting and disposing of sputum containers;
- In areas where droplet nuclei could be present (i.e. a room that has been occupied by a TB case, prior to the time required for air cleaning).

Using respirators needs proper training, fit testing and continuous supervision. This also applies to home-based DOT supervisors.

Visitors/attendants

Visitors and attendants must wear a respirator when entering a contagious TB patient's room.

For more information on respirators, see Appendix 27.

14.5.2 Face or surgical masks

Face masks are medical devices that prevent patients from spreading infectious droplets when talking, coughing or sneezing. They should be worn by contagious patients (suspect or confirmed) when they leave their rooms to go to another department or any other enclosed area. They should not be worn when the patient is alone in his/her room and outdoors.

For more information on surgical masks, see Appendix 28.

Using a mask in public areas could be stigmatizing. Patients can use a cloth scarf to achieve the same purpose.
14.6 Hospital hygiene

14.6.1 Hygiene and disinfection

Sputum containers

Patients with pulmonary TB produce sputum that may contain tubercle bacilli.

- In the wards, patients’ sputum containers should be large (about 200-ml), non-sterile, and sealable. They are to be replaced daily and cannot be re-used.
- In the laboratories, containers for sample collection are smaller (25-35 ml), with hemetic screw cap, non-sterile and for single use.

Environmental cleaning

Sterilization or the use of disinfecting chemicals in a TB patient’s room is not necessary. Ordinary cleaning of rooms and objects (linens, dishes, etc.) used by TB patients is sufficient. After the patient is discharged, air the empty room well according to the calculated ACH.

Reusable medical items

Standard operating procedures for reprocessing items should be followed. There are no specific measures for TB services.

Standard precautions

Standard precautions (hand hygiene, gowns, etc.) apply in TB wards, as they do in any other hospital department.

14.6.2 Waste management

Standard operating procedures for handling and the disposal of healthcare waste (including soft, sharp, etc.) should be followed. There are no specific measures for TB services.

Note: used sputum containers should be collected in a leak proof trash bag and incinerated without filling the containers with chlorine solution before incineration (this can produce toxic gases).
14.7 Patients’ homes

In settings where DR-TB (and HIV) is highly prevalent, systematic TB IC evaluations on patients’ homes are recommended.

TB IC at patients’ homes follows the same principles and measures as in healthcare facilities. Administrative, environmental and personal measures should be followed at least until patient’s smear is negative, ideally until culture conversion.

**Administrative measures**

- Assess the risk of TB transmission: gather information on the number of people that live in the house, number of rooms, etc.
- Screen contacts for TB.
- Children under 5 should spend as little time as possible in the same spaces as culture-positive patient (although the risk to the child is greatly reduced once a patient starts an effective regimen). The mother should use a surgical mask while taking care of the child until she becomes smear-negative.
- Offer education on TB transmission, airborne precautions (cough etiquette, masks), clinical symptoms and waste management of sputum containers or tissues (do not empty the container; throw it in the latrines or enclose it hermetically in plastic bags and discard in the normal waste).

**Environmental measures**

- Ideally, the patient should sleep in a separated room, with door closed off to the rest of the house.
- Common spaces should be well ventilated (often done by keeping windows open at all times).
- The patient should be encouraged to spend time outside in a shaded area if weather permits.

**Personal protective measures**

- If smear-positive or not responding to the regimen, the patient should wear surgical masks when in contact with persons in areas poorly ventilated. Once smear-negative, the patient can be considered non-infectious and no longer needs to wear masks.
- Any person attending to the patient in enclosed spaces should wear a respirator. A fit test should be performed, and the person should be educated on the proper use of respirators. Once the patient is smear-negative, respirators are no longer necessary.
Chapter 15: Follow-up of staff exposed to tuberculosis

15.1 Introduction
15.2 Baseline assessment
15.3 BCG vaccination
15.4 Follow-up

Update: October 2022
15.1 Introduction

The following recommendations apply to staff who work in health facilities and are in contact with tuberculosis (TB) patients and/or infectious laboratory specimens.

They provide general guidance, but should be adapted to the context and regulations of each country.
15.2 Baseline assessment

New staff should undergo a baseline assessment. This includes:

- BCG status (BCG scar check)
- Tuberculin skin test (TST) or interferon gamma release assay (IGRA)
- Chest x-ray (CXR)
- HIV test

In addition, the following information should be provided:

- Risk of occupational transmission of *M. tuberculosis*
- Infection prevention and control (IPC) measures to reduce the risk of transmission
- Higher risk of active TB in immunocompromised individuals (e.g. HIV-infected, diabetics) and in pregnant women
- Vigilance required for, and self-reporting of, signs and symptoms suggestive of TB

Immunocompromised staff and pregnant women should not work in TB departments or areas where the risk of exposure to *M. tuberculosis* is high ([Chapter 14](#)).
15.3 BCG vaccination

Recommendations vary between countries, with some requiring staff to be BCG vaccinated if never vaccinated and TST negative.

There is limited evidence regarding the benefits of BCG vaccination in adults who have not previously had BCG vaccination[1]. Vaccination should be considered on a case-by-case basis in the following situations[2]:

- Significant exposure to multidrug-resistant TB (MDR-TB): facilities treating patients with MDR-TB, prisons, or areas with high MDR-TB prevalence.
- While corrective actions are implemented:
  - when transmission of MDR-TB to staff has occurred;
  - when IPC measures are inadequate or poorly applied.

The following information should be provided to staff considered for BCG vaccination:

- Benefits and risks of BCG vaccination.
- Impact of BCG on the interpretation of TST results in diagnosing a potential latent TB infection (LTBI).
- No complete protection conferred by the vaccine: TB may still occur if IPC measures are not used.

BCG vaccine should only be administered if the person:

- Has never had a BCG vaccination.
- Is HIV-negative.
- Is not pregnant.
- Has never had active TB.
- Has a TST negative result.

For more information on BCG vaccine see Appendix 29.

References

   https://doi.org/10.1093/cid/cit790

2. Centers for Disease Control and Prevention Fact Sheets on BCG Vaccine. 
15.4 Follow-up

Follow-up of routinely exposed staff includes:

- An annual clinical evaluation.
- Assessment for TB (including CXR) and HIV, if symptomatic.

For staff who were TST or IGRA negative at baseline, TST may be performed once a year.

Staff working in a TB department and presenting with a recent immunodepression (e.g. HIV infection, immunosuppressive treatment) or a pregnancy, should be transferred to another department or to an area within the TB department where the risk of exposure to *M. tuberculosis* is low (Chapter 14).

LTBI treatment (Chapter 16) should be offered, after exclusion of active TB:

- Once to staff who become TST or IGRA-positive.
- To all HIV-infected staff.
Chapter 16: Treatment of latent tuberculosis infection

16.1 Introduction
16.2 Target populations
16.3 Latent tuberculosis infection treatment regimens
16.4 Latent tuberculosis infection in HIV-infected patients
16.5 Latent tuberculosis infection in household contacts
16.6 Latent tuberculosis infection in other individuals at risk
16.7 Latent tuberculosis infection and multidrug-resistant tuberculosis
16.8 Follow-up for patients treated for latent tuberculosis infection

Update: January 2022
16.1 Introduction

Exposure to *M. tuberculosis* may result in latent tuberculosis infection (LTBI). WHO defines LTBI as a state of persistent immune response to stimulation by *M. tuberculosis* antigens with no evidence of clinically manifest active tuberculosis (TB)\[1\]. This is also referred to as “tuberculosis infection”.

Identification and treatment of LTBI can reduce TB morbidity and mortality, as well as TB transmission.

Tuberculin skin test (TST) or interferon-gamma release assay (IGRA) can be used to detect LTBI (Chapter 3).

The goal of LTBI treatment is to reduce the risk of progression to active TB. It must be initiated only once active TB has been ruled out by appropriate evaluation.

If a patient develops signs and symptoms of active TB while on LTBI treatment, a specimen should be taken for diagnosis and detection of drug resistance (Xpert MTB/RIF, Xpert MTB/XDR, culture and drug susceptibility test, DST) and according to the results, TB treatment should be initiated.

References

16.2 Target populations

TST or IGRA cannot predict which patients with LTBI are likely to develop active TB. Therefore, widespread LTBI testing and treatment are not recommended.

However, in certain populations, the risk of progression to active TB significantly exceeds that of the general population. For these at-risk populations, the benefits of LTBI treatment of preventing active TB and TB transmission outweigh the potential risks.

Populations who benefit most from LTBI treatment include:

- HIV-infected individuals.
- Household contacts of patients with bacteriologically confirmed pulmonary TB (PTB), in particular children under 5 years.
- Other individuals or populations at risk (e.g. health staff, prisoners).
16.3 Latent tuberculosis infection treatment regimens

There are 3 recommended LTBI treatment regimens and 2 alternative treatment regimens[1]. The decision to prescribe one regimen rather than the other should take into consideration:

- Drug-susceptibility of the strain of the presumed source patient, if known.
- Co-morbidities (e.g. HIV infection, pre-existing hepatic disease or neuropathy).
- Risk of drug interactions (especially with antiretrovirals), tolerability, length of treatment and likelihood of adherence.
- Individual characteristics (e.g. age, pregnancy, living conditions, individual preference).
- Epidemiological and programmatic aspects (e.g. HIV prevalence, available drugs, national recommendations).

Table 16.1 - LTBI treatment regimens

<table>
<thead>
<tr>
<th>Recommended regimens</th>
<th>Weekly Isoniazid dose</th>
<th>Daily rifampicin dose</th>
</tr>
</thead>
<tbody>
<tr>
<td>Isoniazid daily for 6 months (6H) or 36 months (36H)</td>
<td>Isoniazid PO once daily:</td>
<td>&lt; 30 kg: 10 mg/kg (7 to 15 mg/kg)</td>
</tr>
<tr>
<td></td>
<td>≥ 30 kg: 5 mg/kg (4 to 6 mg/kg)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>(max. dose 300 mg daily)</td>
<td></td>
</tr>
</tbody>
</table>

OR

| Isoniazid + rifapentine weekly for 3 months (3HP) | Isoniazid PO once weekly: |< 30 kg and ≥ 2 years: 20 to 30 mg/kg |
|                                                   | ≥ 30 kg: 900 mg + |
|                                                   | Rifapentine PO once weekly[2]: |
|                                                   | 10 to 14 kg and ≥ 2 years: 300 mg |
|                                                   | 14.1 to 25 kg and ≥ 2 years: 450 mg |
|                                                   | 25.1 to 32 kg: 600 mg |
|                                                   | 32.1 to 49.9 kg: 750 mg |
|                                                   | ≥ 50 kg: 900 mg max. |

OR

| Isoniazid + rifampicin daily for 3 months (3HR) | Isoniazid PO once daily: |< 30 kg: 10 mg/kg (7 to 15 mg/kg) |
|                                                 | ≥ 30 kg: 5 mg/kg (4 to 6 mg/kg) |
|                                                 | (max. dose 300 mg daily) + |
|                                                 | Rifampicin PO once daily: |
|                                                 | < 30 kg: 15 mg/kg |
|                                                 | ≥ 30 kg: 10 mg/kg |
|                                                 | (max. dose 600 mg daily) |
Isoniazid monotherapy

Isoniazid monotherapy (or isoniazid preventive therapy, IPT) is the treatment currently most often used for LTBI. This treatment has proven to be effective in preventing active TB in both HIV-infected and non-HIV-infected patients. WHO recommends this treatment in all patients regardless of their HIV status, including children of any age and pregnant women. The main disadvantage of isoniazid monotherapy is the length of treatment. Patients are usually healthy and may not be motivated to complete a 6-month therapy. Adverse effects (e.g. peripheral neuropathy, hepatotoxicity) can also lead to treatment interruption. All patients at risk of peripheral neuropathy should receive pyridoxine (vitamin B₆) for the entire duration of treatment to prevent this risk (for doses see Appendix 17). In HIV-infected patients, the treatment may be difficult due to additive adverse effects of antiretrovirals and isoniazid, the extending of the duration of treatment to 36 months in some adolescents and adults (Section 16.4.2) and the high number of tablets to be taken daily. The number of tablets can be reduced using a fixed-dose combination (FDC) of isoniazid/cotrimoxazole/pyridoxine.

Rifapentine-containing regimens

Combination isoniazid-rifapentine once weekly for 3 months (3HP)

This treatment has proven to be effective in preventing active TB in both HIV-infected and non-HIV-infected patients. WHO recommends this treatment in children 2 years and over, adolescents and adults, regardless of their HIV status. It is short, requires few doses, has a high completion rate and the risk of hepatotoxicity is low. The disadvantages of this regimen are the lack of FDC and the development of hypersensitivity reaction in almost 4% of patients (Section 16.8.3).

Combination isoniazid-rifapentine once daily for 1 month (1HP)

This treatment has proven to be effective in preventing active TB in HIV-infected patients. WHO recommends this treatment as an alternative regimen in patients 13 years and over, regardless of their weight and HIV status. The treatment is short, has a high completion rate and the risk of hepatotoxicity is low. However, cutaneous reactions (rash, itching) are common.

Rifapentine containing regimens are not currently recommended for pregnant women. Despite some reassuring data, safety is not definitively established.

Rifampicin-containing regimens

Combination isoniazid-rifampicin once daily for 3 months (3HR)

**Alternative regimens**

<table>
<thead>
<tr>
<th>Isoniazid + rifapentine daily for 1 month (1HP)</th>
<th>Isoniazid PO once daily:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>≥ 13 years: 300 mg +</td>
</tr>
<tr>
<td></td>
<td>rifapentine PO once daily:</td>
</tr>
<tr>
<td></td>
<td>≥ 13 years: 600 mg</td>
</tr>
</tbody>
</table>

OR

<table>
<thead>
<tr>
<th>Rifampicin daily for 4 months (4R)</th>
<th>Rifampicin PO once daily:</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>&lt; 30 kg: 15 mg/kg</td>
</tr>
<tr>
<td></td>
<td>≥ 30 kg: 10 mg/kg (max. dose 600 mg daily)</td>
</tr>
</tbody>
</table>
This treatment has proven to be effective in preventing active TB in both HIV-infected and non-HIV-infected patients. WHO recommends this treatment in all patients regardless of their HIV status, including children of any age and pregnant women. It is short, safe, has a good completion rate and FDC are available for children and adults. Hypersensitivity reaction may occur in approximately 2% of patients.

**Rifampicin monotherapy once daily for 4 months (4R)**

This treatment has proven to be effective in preventing active TB in non-HIV-infected patients of all ages. WHO recommends this regimen as an alternative regimen in all patients regardless of their HIV status, including children of any age and pregnant women. The advantages of this regimen (better safety profile and completion rate compared to 6H) should be weighed against the risk associated with use of rifampicin in monotherapy (development of resistance to rifampicin in patients with undiagnosed active TB).

**Notes:**

For rifamycin-containing regimens:
- Rifapentine and rifampicin have interactions with many drugs, particularly antiretrovirals (Appendix 19) and contraceptives (Chapter 9).
- For pregnant women taking rifampicin, administer phytomenadione (vitamin K) in the last few weeks of pregnancy (Chapter 9).
- Rifapentine and rifampicin are not interchangeable.
- Rifabutin can replace rifampicin if rifampicin cannot be used due to drug interactions.

**References**


16.4 Latent tuberculosis infection in HIV-infected patients

Treatment of LTBI reduces the risk of active TB by 33-64% [1].

For patients not yet on antiretroviral treatment (ART), ART initiation should take priority over initiation of LTBI treatment. Among these patients, there is a high proportion of undiagnosed, asymptomatic TB cases and it is important to use all existing diagnostic means to rule out active TB.

Note: a treatment programme for LTBI should be combined with a screening programme for active TB in HIV-infected patients (Chapter 6).

16.4.1 Children

HIV-exposed children and HIV-infected children and who do not have active TB (for evaluation, see Chapter 5) should receive LTBI treatment:

- After contact with a TB case, including smear-positive, smear-negative and extrapulmonary TB (EPTB), regardless of their age;
- In high TB transmission areas: if aged 12 months and over, regardless of their contact history.

In addition, for children treated for active TB and living in high TB transmission areas, LTBI treatment may also be prescribed immediately after the successful completion of TB treatment to reduce the risk of reinfection.

16.4.2 Adolescents and adults

HIV-infected adolescents and adults who do not have active TB should receive LTBI treatment, regardless of contact history and TB prevalence in the area.

In areas with high TB transmission, HIV-infected adolescents and adults with a LTBI test positive or unknown and who are unlikely to have active TB (no cough, no fever, no weight loss, no night sweats) should receive the treatment for at least 36 months (long-term regimen).

This regimen is more effective in preventing TB in HIV-infected adults with a positive TST than those with a negative TST [2].

If TST is not feasible, or where the national guidelines do not recommend long-term isoniazid monotherapy, HIV-infected adolescents and adults without any TB symptoms should receive another LTBI treatment (6H or a rifapentine- or rifampicin-containing regimen).

Table 16.2 – LTBI treatments for HIV-infected patients

<table>
<thead>
<tr>
<th>Age</th>
<th>Recommended regimens</th>
<th>Alternative regimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Child &lt; 2 years</td>
<td>6H or 3RH</td>
<td>4R</td>
</tr>
<tr>
<td>Child ≥ 2 years</td>
<td>6H or 3HP or 3RH</td>
<td>4R</td>
</tr>
<tr>
<td>Adolescent and adult</td>
<td>6H or 3HP or 3RH or 36H</td>
<td>1HP (if ≥ 13 years) or 4R</td>
</tr>
</tbody>
</table>

Footnotes

(a) HIV-exposed children are children born to HIV-infected women whose HIV status has not been established and/or are still at risk of infection (e.g. still breastfed).

References

16.5 Latent tuberculosis infection in household contacts

A household contact is a person who has shared the same enclosed living space as the index case for one or more nights or for frequent or extended daytime periods during 3 months before the start of the current treatment[1].

16.5.1 Neonates of mothers with active pulmonary tuberculosis

All neonates born to mothers with active PTB should receive treatment for LTBI, after exclusion of active TB, if the mother:

- Has been treated for PTB less than 2 weeks at the time of birth, or
- Has a positive smear microscopy result on a sputum sample collected at birth or close to the time of birth[2].

Xpert MTB/RIF and Xpert MTB/XDR assays should be have been performed to rule out resistance to rifampicin and isoniazid in the mother before starting treatment for LTBI in the neonate.

The recommended regimens are 3HR or 6H. For HIV-exposed neonates receiving nevirapine, only 6H is recommended.

BCG vaccine should be administered just after LTBI treatment completion (not during the treatment).

If a TST is feasible and the regimen chosen is 6H:

- Administer isoniazid for 3 months, then perform a TST.
- If the TST is positive, complete isoniazid monotherapy.
- If the TST is negative, stop isoniazid and administer the BCG vaccine.

Notes:

- A neonate should not be separated from its mother unless severely ill.
- Breastfeeding should continue, and breastfed neonates should receive pyridoxine (vitamin B₆).

16.5.2 Other household contacts

Children under 5 years

It is not mandatory to perform TST or IGRA prior to LTBI treatment.

All children < 5 years in contact with a confirmed PTB case and who do not have active TB (for evaluation, see Chapter 5) should receive LTBI treatment, regardless of their HIV and BCG vaccination status.

If LTBI treatment is contra-indicated or in case of parental refusal, monitor the child closely for one year to enable the early detection of active TB.

Children 5 years and older, adolescents and adults

A TST or IGRA should be performed prior to LTBI treatment. If this is not feasible, LTBI treatment may be considered, weighing benefits and risks.

- Children 5 years and over in contact with a confirmed PTB case and who do not have active TB (for evaluation, see Chapter 5) may receive LTBI treatment, regardless of their HIV status.
- Adolescents and adults in contact with a confirmed PTB case and who do not have active TB (no TB symptoms and no abnormality on CXR) may receive LTBI treatment, regardless of their HIV status.

Table 16.3 - LTBI regimens for household contacts
<table>
<thead>
<tr>
<th>Age</th>
<th>Recommended regimens</th>
<th>Alternative regimens</th>
</tr>
</thead>
<tbody>
<tr>
<td>Child &lt; 2 years</td>
<td>6H or 3RH</td>
<td>4R</td>
</tr>
<tr>
<td>Child ≥ 2 years and &lt; 5 years</td>
<td>6H or 3HP or 3RH</td>
<td>4R</td>
</tr>
<tr>
<td>Child ≥ 5 years, adolescent, adult</td>
<td>6H or 3HP or 3RH</td>
<td>1HP (if ≥ 13 years) or 4R</td>
</tr>
</tbody>
</table>

**References**


   [https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4181157/](https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4181157/)
16.6 Latent tuberculosis infection in other individuals at risk

Routine LTBI testing (TST or IGRA) and treatment after exclusion of active TB:

- Are recommended for patients with silicosis, on dialysis or taking long-term immunosuppressive therapy.
- Can be considered for health staff, populations in congregate living settings (e.g. prisoners, refugees), migrants from countries with a high TB prevalence, homeless people and drug users.

LTBI testing should be performed periodically (e.g. once a year).

Routine LTBI testing and treatment is not recommended for diabetic, malnourished or alcoholic patients, unless they belong to the above-mentioned risk groups.
16.7 Latent tuberculosis infection and multidrug-resistant tuberculosis

Due to limited evidence, routine LTBI treatment for all household contacts of multidrug-resistant TB (MDR-TB) patients cannot be recommended at this time.

However, treatment of LTBI should be considered in certain high-risk household contacts based on an individual risk-benefit assessment. Individual assessment includes:

- High risk of progression to active TB: children under 5 years, individuals with HIV infection or on immunosuppressive therapy.
- Resistance pattern of the source case: the LTBI treatment regimen must be individually tailored as contacts of MDR-TB patients are often infected with the same strain[1].
- Intensity of exposure.
- Contra-indication or risk of adverse drug reactions.

A TST or IGRA should be performed prior to LTBI treatment. If not feasible, LTBI treatment may be considered, weighing benefits and risks.

16.7.1 Household contacts of multidrug-resistant tuberculosis cases eligible for treatment

Evidence is lacking on the choice of treatment to prevent disease in MDR-TB contacts. Few observational studies, primarily using a fluoroquinolone (FQ) for 6 months, reported promising results[2][3]. Randomized clinical trials are ongoing[4][5].

For contacts of FQ-susceptible MDR-TB patients, levofloxacin PO for 6 months can be proposed at the following doses:

<table>
<thead>
<tr>
<th>Weight</th>
<th>5 to 9 kg</th>
<th>10 to 15 kg</th>
<th>16 to 23 kg</th>
<th>24 to 34 kg</th>
<th>35 to 45 kg</th>
<th>&gt; 45 kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Daily dose</td>
<td>150 mg</td>
<td>200 to 300 mg</td>
<td>300 to 400 mg</td>
<td>500 to 750 mg</td>
<td>750 mg</td>
<td>1 g</td>
</tr>
</tbody>
</table>

If active TB develops during LTBI treatment, DST including resistance to FQs is necessary due to the potential risk associated with use of FQs in monotherapy (development of resistance to FQs in patients with undiagnosed active TB). Independent of LTBI treatment, monitor these patients for 2 years for the development of active TB.

16.7.2 Household contacts of multidrug-resistant tuberculosis cases not eligible for treatment

If the contact is not eligible for LTBI treatment, closely monitor for signs and symptoms of active TB every 3 months for the next 2 years.

If active TB develops, initiate TB treatment promptly with a regimen designed according to the DST. If DST is not feasible, a regimen can be designed according to the resistance profile of the source case.

References


https://clinicaltrials.gov/ct2/show/NCT03568383

5. Tuberculosis child multidrug-resistant preventive therapy: TB CHAMP trial.
https://doi.org/10.1186/ISRCTN92634082
16.8 Follow-up for patients treated for latent tuberculosis infection

For the modality of administration of LTBI treatments see Chapter 13.

16.8.1 Baseline assessment of liver function

Before initiating LTBI treatment, look for clinical signs of hepatic disease and specific risks of hepatotoxicity.

For patients with hepatic disease, baseline liver function tests (LFTs), i.e. aspartate aminotransferase (AST), alanine aminotransferase (ALT) and bilirubin should be performed.

The benefit of LTBI treatment should be weighed against the potential risk of aggravation of existing hepatic disease. LTBI treatment is contra-indicated in patients with end-stage hepatic disease or LFTs > 5 times the upper limit of normal (ULN) and should be used with caution in patients with LFTs > 3 times ULN.

Depending on available resources, baseline LFTs can be performed in groups at risk for hepatotoxicity (e.g. patients with HIV infection, women during pregnancy and post-partum period, chronic alcohol consumption, age > 35 years, concomitant use of hepatotoxic drugs, history of hepatic disease).

16.8.2 Follow-up

All patients should be evaluated monthly for signs and symptoms of active TB, adverse effects and adherence.

TST or IGRA should not be repeated.

In patients with pre-existing hepatic disease:

- Baseline LFTS are normal: monitor LFTs monthly.
- Baseline LFTs are elevated or LFTs increase during LTBI treatment: monitor LFTs once a week.

Other patients should be tested if they develop symptoms of hepatotoxicity.

Any problems with adherence should be addressed with the patient.

If signs and symptoms of active TB develop, the patient should undergo full evaluation (Chapter 3, Chapter 4 and Chapter 5).

16.8.3 Management of adverse effects

Hepatotoxicity

Clinical features resemble that of viral hepatitis. Early symptoms include malaise, fatigue, loss of appetite, muscle and joint pain. Nausea, vomiting and abdominal pain are common in severe disease. Jaundice, scleral icterus, dark (tea-coloured) urine and discoloured stool are signs of clinical worsening.

Clinical hepatitis can be fatal, so action should be taken immediately.

- Patient with symptoms of hepatitis:
  Stop all TB drugs and perform LFTs:
  a) AST or ALT or bilirubin ≥ 3 times ULN or severe symptoms: do not re-initiate LTBI treatment.
  b) AST, ALT, and bilirubin < 3 times ULN and mild symptoms (no jaundice): after discussion with the patient on benefits and risk, treatment may be re-initiated. Closely monitor the patient and perform LFTs once a week. Continue treatment as long as LFTs levels remain < 3 ULN and there are no signs of worsening hepatitis.
  c) If LFTs are not available, do not re-initiate LTBI treatment.

- Patient without symptoms of hepatitis, but elevated LFTs:
  a) AST or ALT ≥ 5 times ULN or bilirubin ≥ 3 ULN: stop and do not re-initiate LTBI treatment.
  b) AST and ALT < 5 times ULN and bilirubin < 3 ULN: stop LTBI treatment. Perform LFTs once a week. If LFTs return to normal, after discussion with the patient on benefits and risk, treatment may be re-initiated. Closely monitor the patient and perform LFTs once a week.
Note: 10-20% of patients taking isoniazid alone may have a mild, transient, asymptomatic elevation of LFTs (AST and/or ALT). In most cases, this does not require treatment interruption.

Hypersensitivity reaction

Approximately 2% of patients on 3HR regimen and 4% of patients on 3HP regimen have hypersensitivity reaction, typically after the first 3 to 4 doses. Symptoms may include fever, headache, dizziness, nausea and vomiting, muscle and bone pain, rash, itching, red eyes, angioedema, shortness of breath and, more rarely, hypotension and altered consciousness. In case of hypersensitivity reaction, treatment should be stopped immediately. Symptoms usually resolve within 24 hours after TB drug withdrawal. In case of mild reaction (fever, rash, itching), consider re-initiating the treatment. In this case, the patient should be observed at least 4 hours after each dose is administered to detect first signs of hypersensitivity reaction.

Other adverse effects

See Appendix 17.

References

https://apps.who.int/iris/rest/bitstreams/1272664/retrieve

https://doi.org/10.1164/rccm.200510-1666ST

https://doi.org/10.1093/cid/civ323
Chapter 17: Monitoring and evaluation

17.1 Introduction

17.2 Definitions of treatment outcomes

17.3 Recording tools

17.4 Reporting

17.5 Programme assessment

Update: January 2014
17.1 Introduction

Monitoring and evaluation rely on both quantitative and qualitative information in order to provide information on the following:

- Programme performance (e.g. number of patients started on anti-TB treatment, treatment results, number of patients tested for MDR-TB, etc.);
- Planning for human resources, patient support, diagnostic tests and drug orders, etc.;
- Evaluation of the functioning of the programme (quality of drugs, diagnostics, patient support, etc.).

Recording and reporting are based on a set of standard case and outcome definitions.

Case definitions are presented in Chapter 7.
17.2 Definitions of treatment outcomes

For all forms of TB, outcome definitions have many similarities. These are:
- Outcome assignment is standardized, as to permit comparisons across clinicians, time and sites.
- Outcome assignment relies heavily, but not exclusively, on bacteriologic endpoints (smear or culture*).
- Outcomes are mutually exclusive and exhaustive.

For all forms of TB, definitions exist for:
- Interim outcomes (intended to have an indication on how the programme is functioning before final outcomes are available);
- Final outcomes (cure, completion, failure, treatment interruption, death or not evaluated).

17.2.1 Interim outcomes for drug-susceptible TB and MDR-TB

Given that TB treatment is long (6 to 18 months or more), interim outcomes provide early indicators of programme results. Table 17.1 provides a summary on interim outcomes.

**Table 17.1 - Interim outcomes**

<table>
<thead>
<tr>
<th>TB</th>
<th>Interim outcomes</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Drug-susceptible TB</strong></td>
<td>At 2-3 and 4-5 months:</td>
</tr>
<tr>
<td></td>
<td>• Bacteriological status (smear negative/positive/no information)</td>
</tr>
<tr>
<td></td>
<td>• Final outcomes in patient who had already interrupted or died</td>
</tr>
<tr>
<td><strong>MDR-TB</strong></td>
<td>At 6 months:</td>
</tr>
<tr>
<td></td>
<td>• Bacteriological status (negative/positive/no information) based on smear and culture</td>
</tr>
<tr>
<td></td>
<td>• Final outcomes in patient who had already interrupted or died</td>
</tr>
</tbody>
</table>

17.2.2 Final outcomes for drug-susceptible TB and DR-TB

Table 17.2 provides definitions for the final outcomes.

**Table 17.2 - Summary table of final outcome definitions[1][2]**

---

*a* Bacteriologic endpoints include smear (acid-fast bacilli) and culture results.
<table>
<thead>
<tr>
<th>Outcomes</th>
<th>TB</th>
<th>Definitions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cured</td>
<td>DS TB</td>
<td>Patient initially bacteriologically confirmed (microscopy, culture or molecular test) who completed treatment AND shows no signs of continued active disease AND has at least 2 negative smears or cultures: one at 4-5 months and the other at the end of treatment AND does not meet the definition of failure.</td>
</tr>
<tr>
<td></td>
<td>PDR-TB</td>
<td>Patient initially bacteriologically confirmed (culture or molecular test), who completed treatment AND has been consistently culture-negative with at least 3 results on sputum tested at least one month apart for the final 6 months of treatment AND does not meet the definition of failure.</td>
</tr>
<tr>
<td></td>
<td>MDR-TB</td>
<td>Patient initially bacteriologically confirmed (culture or molecular test), who completed treatment AND with at least 3 negative cultures in the last 8 months of treatment AND does not meet the definition of failure. If there is a lone positive culture or smear reported during that time, and no concomitant clinical evidence of deterioration, a patient may still be considered cured, provided that this positive culture is followed by a minimum of 3 consecutive negative cultures taken at least 30 days apart.</td>
</tr>
<tr>
<td>Completed</td>
<td>All</td>
<td>Patient who completed treatment AND has no signs of continued active disease AND does not meet the bacteriological criteria for cure.</td>
</tr>
<tr>
<td>Failure</td>
<td>DS TB</td>
<td>Patient with signs of continued active disease or deterioration requiring a treatment change: • Any patient with positive smear or culture at 4-5 months of treatment or thereafter. • Any patient with no significant clinical improvement, no significant gain of weight after 4-5 months of treatment and for whom the diagnosis of failure is established by a clinician.</td>
</tr>
<tr>
<td></td>
<td>DR-TB[e][f][g]</td>
<td>Treatment terminated or need for permanent treatment change of at least 2 classes of anti-TB drugs because of one or more of the following: • Lack of monitoring cultures converting to negative by 6 months for MDR-TB (3 months for PDR-TB), and/or • Resistance amplification to rifampicin or isoniazid (PDR-TB) or to Group 2 or Group 3 drugs (MDR-TB), and/or • Bacteriological reversion (at least two positive smears or cultures at least 7 days apart after monitoring smears or cultures have become negative), or • A clinical decision has been made to terminate treatment early due to poor response or adverse events. These latter failures can be indicated separately in order to do sub-analysis.</td>
</tr>
<tr>
<td></td>
<td>All</td>
<td>Patient who interrupted treatment for 2 months or more.</td>
</tr>
<tr>
<td>Death</td>
<td>All</td>
<td>Patient who died on TB treatment or while awaiting TB treatment, irrespective of the cause of death. The cause of death should be recorded.</td>
</tr>
<tr>
<td>Treatment adapted[e][f]</td>
<td>DS TB</td>
<td>Patient initially treated with a standard regimen and for whom the treatment is secondarily adapted according to the results of DST (and not because of a treatment failure).</td>
</tr>
<tr>
<td>Not evaluated</td>
<td>All</td>
<td>Patient whose treatment outcome is unknown (including patients “transferred out” to another treatment centre, for whom the outcome is unknown).</td>
</tr>
</tbody>
</table>
If treatment is continuing at the time of a cohort analysis, an outcome of “still on treatment” may be provisionally assigned.

Footnotes
(a) Molecular techniques are not used to monitor treatment response or to declare failure. These tests may identify dead bacilli for a long time and can even be positive after a patient is truly cured.

References

17.3 Recording tools

Forms used in recording and reporting can be found in the appendices. They are intended to be examples that programmes or country can use to produce their own forms.

17.3.1 Drug-susceptible TB treatment card and drug-susceptible TB register

Drug-susceptible TB treatment card and drug-susceptible TB register (Appendix 23 and Appendix 24) are used for all new patients or previously treated patients treated by standard first-line regimens (with or without confirmation of the drug susceptibility by a DST).

17.3.2 DR-TB treatment card and DR-TB register

DR-TB treatment card (Appendix 25) tracks, in particular, each dose of each drug taken during the full course of treatment. The number of actually observed doses and the number of expected observed doses are reported each month. In addition to the treatment card, it is recommended to keep a medical chart with a full admission note at the time of enrolment and a progress note at each medical encounter.

DR-TB register (Appendix 26) includes data on case definition, bacteriological exams (indicate date of specimen collection and not the date of result), type of treatment and treatment outcome. It is a separate register from the drug-susceptible TB register.

Each DR-TB patient detected should be registered, including patients who refuse treatment.

Transfer of patients from the drug-susceptible TB register to the DR-TB register is done usually while on treatment when DST results are available. Patient’s outcome is reported as ‘treatment adapted’ in the comment row of the drug-susceptible TB register (Appendix 24).

17.3.3 Laboratory request form(s) and register(s)

- Request form for smear microscopy and Xpert assays (Appendix 34);
- Request form for sputum culture, LPA and DST (Appendix 28);
- Sputum smear microscopy register (Appendix 29);
- Xpert MTB/RIF register (Appendix 30).

17.3.4 Drug-O-Gram

The Drug-O-Gram is a summary of the patient’s treatment history. It includes consecutive DST and treatment changes presented in a chronological order and gives a short summary of the patient status (Appendix 31).
17.4 Reporting

The key evaluation tool for all forms of TB is the periodic report. It must be presented in a standardized manner in two parts: case enrolment and treatment outcomes. The data presented in the report comes from the TB register. It is generally completed by quarter for drug-susceptible TB and by semester for DR-TB.

Evaluation of interim and final treatment outcomes is a fundamental stage in the evaluation. This evaluation is done through a cohort analysis. A "cohort" is a group of individuals presenting certain common characteristics and undergoing the same events. In respect to the evaluation of TB patients, a cohort is represented by patients all put under treatment within a given period of time (usually a quarter for drug-susceptible TB and a semester for DR-TB). At the end of treatment, a final outcome is assigned to each patient (Table 17.1).

Notes:

- The number of patients in each group should, in principle, be identical to those registered for the same interval in the case enrolment part of the corresponding periodic report. If it is different, an explanation should be given (e.g., patients "interrupting before treatment" can be excluded from the outcome analysis).
- The outcomes of patients "transferred in" should not be included in the outcomes of the facility to which they were transferred. Their outcome results should be recorded in the facility that initially enrolled the patient in TB treatment.

17.4.1 Case detection and enrolment report for TB

The elements necessary for defining a TB case (treatment history, bacteriological status, anatomical site of the disease, and HIV status) are defined in Chapter 7.

See Quarterly report for case enrolment, Appendix 32.

Main indicators

- **Proportion of confirmed pulmonary TB (PTB)**
  \[
  \text{Proportion of confirmed pulmonary TB (PTB)} = \frac{\text{Number of PTB cases confirmed enrolled}}{\text{Total number of TB cases enrolled for the period}}
  \]
  With the introduction of automated molecular tests and rapid cultures, it is expected that the proportion of confirmed PTB cases will increase as compared to programmes where only smear microscopy is available.

- **Proportion of smear-negative PTB**
  \[
  \text{Proportion of smear-negative PTB} = \frac{\text{Number of smear-negative PTB cases enrolled}}{\text{Total number of TB cases enrolled for the period}}
  \]
  This indicator essentially depends on the following: the quality of microscopy, the number of children under treatment (children are rarely smear-positive), the prevalence of HIV infection within the population (these patients present more smear-negative PTB), and the other diagnostics used (culture, Xpert MTB/RIF, etc).
  The proportion of smear-negative PTB is about 20% when HIV prevalence is low. It is 40 to 60% when HIV prevalence is high. Proportions that differ significantly from these should make one consider the possibility of under- or over-diagnosis of smear-negative forms.

- **Proportion of smear-positive PTB**
  \[
  \text{Proportion of smear-positive PTB} = \frac{\text{Number of smear-positive PTB cases enrolled}}{\text{Total number of TB cases enrolled for the period}}
  \]
  In practice, the proportion of smear-positive PTB should correspond to roughly half of all patients. This proportion is lower, however, in areas where HIV prevalence is high. Proportion of smear-positive PTB is around 60% where HIV prevalence is low, and it is 30 to 40% where HIV prevalence is high. Proportions that differ significantly from these should make one consider the possibility of under- or over-diagnosis of smear-negative pulmonary TB and extra-pulmonary B forms.

- **Proportion of new cases**
  \[
  \text{Proportion of new cases} = \frac{\text{Number of new TB cases enrolled}}{\text{Total number of cases enrolled for the period}}
  \]
  This indicator indirectly reflects the relapse and failure rates and possible parallel treatments outside the programme.

- **Proportion of children**
  \[
  \text{Proportion of children} = \frac{\text{Number of TB patients less than 15 years enrolled}}{\text{Total number of TB cases enrolled for the period}}
  \]
  Children should represent approximately 10 to 15% of the total number of patients. Proportions that differ significantly from these should make one consider the possibility of under- or over-diagnosis of TB in children.
Note: even the best programmes often do not detect more than 60 to 70% of expected new smear-positive cases within a population. In addition, patients might come from outside the target area.

17.4.2 Case detection and enrolment report for DR-TB

See standard DR-TB case detection and enrolment reports in Appendix 33.

Early detection of resistance is intended to ensure that an appropriate treatment is initiated from the start. DST is usually performed for patients at risk of DR-TB. Target groups vary according to local situation, but should at a minimum always include patients who have been previously treated and contacts of confirmed MDR-TB patients.

The indicators for detection aim at measuring the access of TB patients to DST. The frequency of MDR-TB among individuals in different risk groups is also evaluated.

All patients in whom DR-TB is highly suspected or detected should be started on appropriate treatment in the shortest time possible.

A comparison of enrolled patients under treatment to detected DR-TB cases gives an indication of access to care, though some patients started on treatment may have been detected prior to the period of assessment.

The period of assessment is six calendar months. This is usually counted from January to the end of June and July to the end of December. Indicators are measured three months after the end of the six-month period. All data can be extracted from the DR-TB register (Appendix 26), the laboratory register for culture and DST and the Xpert register (Appendix 30).

Each indicator should be calculated for all patients and for each risk group of patients, including: all cases, previously treated cases, failures, household contacts and other local risk groups according to the strategy.

**Case detection indicators**

- **Proportion of TB patients detected with DST result for isoniazid and rifampicin (for each risk group during the period)**
  \[ \text{Proportion} = \frac{\text{Number of TB cases detected with DST result for both isoniazid and rifampicin}}{\text{Total number of TB cases detected}} \]

- **Proportion of TB patients detected with Xpert MTB/RIF result (for each risk group during the period)**
  \[ \text{Proportion} = \frac{\text{Number of TB cases detected with Xpert MTB/RIF result}}{\text{Total number of TB cases detected}} \]

- **Proportion of confirmed MDR-TB cases detected among TB patients tested for isoniazid and rifampicin DST (for each risk group during the period)**
  \[ \text{Proportion} = \frac{\text{Number of TB cases with confirmed resistance to isoniazid and rifampicin}}{\text{Total number of TB cases tested for these 2 drugs}} \]

- **Proportion of Xpert RIF resistant cases detected among patients tested by Xpert MTB/RIF (for each risk group during the period)**
  \[ \text{Proportion} = \frac{\text{Number of Xpert RIF resistant cases}}{\text{Total number of TB cases with Xpert MTB/RIF result}} \]

**Enrolment indicators**

- **Proportion of confirmed MDR-TB cases enrolled on MDR-TB treatment**
  \[ \text{Proportion} = \frac{\text{Number of confirmed MDR-TB cases registered and started on MDR-TB treatment}}{\text{Total number of confirmed MDR-TB cases detected}} \]
  This can also be calculated for rifampicin resistant TB cases.

- **Proportion of confirmed PDR-TB cases enrolled on PDR-TB treatment**
  \[ \text{Proportion} = \frac{\text{Number of confirmed PDR-TB cases registered and started on PDR-TB treatment}}{\text{Total number of confirmed PDR-TB cases detected}} \]
  This calculation does not include rifampicin resistance and unknown isoniazid resistance.
17.4.3 Interim treatment outcomes for drug-susceptible TB and DR-TB

Interim analysis should be completed approximately 3 months after all patients who were registered during a particular interval completed the intensive phase of treatment (three months should allow culture results for all those patients).

**Interim treatment outcomes for drug-susceptible TB**

Interim results at Month 2 or 3 should be evaluated for all patients treated as new or previously treated patients by standard first-line regimens (with or without confirmation of the drug susceptibility by a DST). These results may be disaggregated by treatment history (new, previously treated, and by type of previous treatment).

At the beginning of a programme, when it is not yet possible to do cohort analysis, the conversion rate at Month 2-3 is a proxy indicator of the effectiveness of treatment, and it allows early detection of potential problems. The smear conversion rate of new smear-positive patients is the proportion of new smear-positive patients who are smear-negative at Month 2. The smear conversion rate of previously treated smear-positive patients is the proportion of previously treated smear-positive patients who are smear-negative at Month 3.

**Interim treatment outcomes for DR-TB**

The period of assessment is six calendar months, usually counted from January to end June, July to end December. All patients registered and starting treatment during the period of assessment are included in the calculation. The interim report form should be completed 9 months after the closing day of the cohort. This allows culture information at 6 months of treatment to be included for all patients in the cohort. For instance, interim results of TB patients who started treatment during the first semester of a year (1 January to 30 June), should be calculated at the beginning of April of the following year.

Culture conversion (for confirmed DR-TB cases) and death by six months are used as proxies for final outcomes. Information on treatment interruption by six months is helpful. It is also useful to know how many patients started on second-line drugs for MDR-TB turned out not to be MDR.

All data can be extracted from the DR-TB register (Appendix 26).

At six months:

- **Proportion of death**
  \[ \frac{\text{Number of confirmed MDR-TB cases registered and started on MDR-TB treatment who died of any cause by the end of Month 6}}{\text{Total number of confirmed MDR-TB cases started on treatment for MDR-TB during the period}} \]

- **Proportion of treatment interrupted**
  \[ \frac{\text{Number of confirmed MDR-TB cases started on MDR-TB treatment who interrupted by the end of Month 6}}{\text{Total number of confirmed MDR-TB cases started on treatment for MDR-TB during the period}} \]

- **Proportion with negative culture**
  \[ \frac{\text{Number of bacteriologically confirmed pulmonary MDR-TB cases registered and started on MDR-TB treatment with negative culture at Month 6}}{\text{Total number of bacteriologically confirmed pulmonary MDR-TB cases registered and started on treatment for MDR-TB during the period}} \]

- **Proportion with positive culture**
  \[ \frac{\text{Number of bacteriologically confirmed pulmonary MDR-TB cases registered and started on MDR-TB treatment with positive culture at Month 6}}{\text{Total number of bacteriologically confirmed pulmonary MDR-TB cases registered and started on treatment for MDR-TB during the period}} \]

- **Proportion found not to have MDR-TB**
  \[ \frac{\text{Number of patients started on MDR-TB treatment during the period and later found not to be MDR}}{\text{Total number of patients started on MDR-TB treatment during the period}} \]

17.4.4 Final treatment outcomes for TB

See standard TB treatment outcomes reports (Appendix 32 and Appendix 33).

The final outcome is the most important direct measurement of the effectiveness of a TB programme in terms of patient care. All patients entered on the TB register should be assigned one of six mutually exclusive outcomes at the end of their therapy. All patients should be assigned the first outcome they experience for the treatment being evaluated\(^1\).
Final treatment outcome cohort analysis could be carried out when all patients admitted in a given period of time had a chance to complete their treatment. In practice:

- For drug-susceptible TB (and all patients treated by standard first-line regimens) cohort results are analysed quarterly, one year after inclusion of the last patient of the cohort (e.g. cohort of patients admitted during the first quarter 2014 will be evaluated at the end of the first quarter 2015).
- For DR-TB, evaluation occurs 27 months after inclusion of the last patient in the cohort in order to have the results of cultures performed at 24 months. The period of assessment is six calendar months, usually counted from January to the end of June and July to the end of December. All patients starting treatment during this period are included in the calculation. Indicators are measured 24 months after the end of the semester of assessment. All data can be extracted from the DR-TB register.

Although the timing of the analysis is different for drug-susceptible TB and DR-TB, the indicators are the same.

Indicators should be calculated for patients treated by standard first-line regimens (with or without confirmation of drug-susceptible TB by a DST), and for patients with PDR-TB and MDR-TB.

The most important indicators are:

- **Proportion of cured**
  \[ \text{Proportion of cured} = \frac{\text{Number of confirmed TB cases declared "cured"}}{\text{Total number of confirmed TB cases put under treatment during the period}} \]
  This indicator is calculated for all confirmed drug-susceptible TB cases and DR-TB cases. It is the best indicator of the success of a programme for confirmed TB patients. Though the effectiveness of the treatment for drug-susceptible TB is theoretically above 90%, the proportion of cure is rarely above 70%. For MDR-TB this indicator rarely exceeds 50%.

- **Proportion of treatment completed**
  \[ \text{Proportion of treatment completed} = \frac{\text{Number of patients registered as "treatment completed"}}{\text{Total number of patients put under treatment for the period}} \]
  A high proportion of patients completing treatment is a positive sign for not confirmed PTB and EPTB. For confirmed TB, it indicates insufficient bacteriological verification at the end of treatment, thus, suggesting that a step should therefore be reinforced.

- **Proportion with successful outcome**
  \[ \text{Proportion with successful outcome} = \frac{\text{Number of patients registered as "cured" or "treatment completed"}}{\text{Total number of patients put under treatment during the period}} \]
  This is the best indicator to measure the efficacy of a programme for all forms of TB (confirmed and not confirmed, PTB and EPTB). This indicator rarely exceeds 80% for drug-susceptible TB and 60% for MDR-TB.

- **Proportion of treatment interrupted**
  \[ \text{Proportion of treatment interrupted} = \frac{\text{Number of patients registered as "treatment interrupted"}}{\text{Total number of patients put under treatment during the period}} \]
  Patients who interrupted treatment are at risk of not being cured or of relapsing. Treatment interruption indicates a failure of the programme in supporting the patient to be able to successfully complete treatment.

- **Proportion of death**
  \[ \text{Proportion of death} = \frac{\text{Number of patients registered as "death"}}{\text{Total number of patients put under treatment during the period}} \]
  This ratio usually does not exceed 5% for drug-susceptible TB. Over-mortality may be related to the poor functioning of a programme. It may also be due to a high prevalence of HIV infection among cases or late referrals.

- **Proportion of failure**
  \[ \text{Proportion of failure} = \frac{\text{Number of patients registered as "failures"}}{\text{Total number of patients put under treatment during the period}} \]
  A high failure rate in new cases can be related to poor treatment adherence, high rate of primary resistance or poor quality of anti-TB drugs. The failure rate should not be over 2% in new cases under treatment.

- **Proportion of patients for whom HIV status is known**
  \[ \text{Proportion of patients for whom HIV status is known} = \frac{\text{Number of patients for whom HIV status is known by the end of treatment}}{\text{Total number of patients put under treatment during the period}} \]
  This is one of the indicators that help evaluate the integration of TB and HIV services.

- **TB-HIV co-infection rate**
  \[ \text{TB-HIV co-infection rate} = \frac{\text{Number of HIV-infected TB patients}}{\text{Total number of TB patients put under treatment during the period and for whom HIV status is known at the end of treatment}} \]
  In high HIV-prevalence regions, co-infection rate may exceed 80%. This information is important in assessing other indicators, in particular the proportion of death.
References

   
17.5 Programme assessment

To be complete, evaluation should look at how well the programme functions, particularly with respect to three aspects: organization of care, established procedures and human resources. A set of quality criteria is evaluated for each of these aspects. The criteria may be either qualitative (description) or quantitative (indicators). The following tables can be used as a rough guide.

17.5.1 Organization
<table>
<thead>
<tr>
<th>Criteria</th>
<th>Indicators</th>
<th>Goals</th>
</tr>
</thead>
<tbody>
<tr>
<td>Access to care</td>
<td>• Accessibility of treatment facilities, decentralization, etc.</td>
<td>Easy access to care during the intensive/continuation phases</td>
</tr>
<tr>
<td></td>
<td>• Home-based treatment available when appropriate.</td>
<td></td>
</tr>
<tr>
<td>Patient comfort</td>
<td>• Patient welcome</td>
<td>According to needs</td>
</tr>
<tr>
<td></td>
<td>• Condition of the facility, heating (or cooling), overall organization and cleanliness.</td>
<td>Bed occupancy rate ≤ 100%</td>
</tr>
<tr>
<td></td>
<td>• Food during hospitalization and/or for outpatients (supplemental rations, quantities, organization in charge).</td>
<td></td>
</tr>
<tr>
<td></td>
<td>• Bed occupancy rate of the TB ward.</td>
<td></td>
</tr>
<tr>
<td>Information and therapeutic education</td>
<td>Patient interviews conducted.</td>
<td>Patient understanding of treatment</td>
</tr>
<tr>
<td>Hospital hygiene</td>
<td>• Equipment (respirators, masks, gloves, gowns, autoclaves, cleaning supplies, etc.)</td>
<td>All necessary equipment is available and used.</td>
</tr>
<tr>
<td></td>
<td>• Waste management (sorting, incinerator, etc.)</td>
<td></td>
</tr>
<tr>
<td>Constant supply of lab materials</td>
<td>• Supplied by (government, agency or facility, other)</td>
<td>3-month buffer stock</td>
</tr>
<tr>
<td></td>
<td>• Buffer stock</td>
<td>No shortages</td>
</tr>
<tr>
<td></td>
<td>• Number and duration of shortages</td>
<td></td>
</tr>
<tr>
<td>Constant supply of quality-assured anti-TB drugs</td>
<td>• Stock card maintenance</td>
<td>Stock cards up-to-dated</td>
</tr>
<tr>
<td></td>
<td>• Order frequency, delivery time, buffer stock</td>
<td>One person in charge of the pharmacy</td>
</tr>
<tr>
<td></td>
<td>• Shortage(s)</td>
<td>All adequate</td>
</tr>
<tr>
<td></td>
<td>• Drug sources</td>
<td>No shortages</td>
</tr>
<tr>
<td></td>
<td>• Institution in charge of supply</td>
<td>WHO-prequalified sources (or equivalent)</td>
</tr>
<tr>
<td></td>
<td>• Use of FDCs first-line drugs</td>
<td>Use of FDCs</td>
</tr>
<tr>
<td></td>
<td>• Storage conditions</td>
<td>Appropriate storage conditions</td>
</tr>
<tr>
<td></td>
<td>• Organization of supply for peripheral facilities</td>
<td>Regular supply</td>
</tr>
<tr>
<td>Case detection</td>
<td>• Type of case detection (active or passive)</td>
<td>Know the type, in order to interpret the quantitative results of case detection</td>
</tr>
<tr>
<td></td>
<td>• Contacts screening</td>
<td>• 100%</td>
</tr>
<tr>
<td></td>
<td>• Detection rate of new smear-positive cases</td>
<td>• Depends on the context</td>
</tr>
<tr>
<td></td>
<td>• Percentage of smear-positive patients out of the total number of patients who had a sputum smear.</td>
<td>• &lt; 20%</td>
</tr>
<tr>
<td></td>
<td>• Detection rate of MDR-TB</td>
<td>• Depends on the context</td>
</tr>
<tr>
<td>Diagnosis of smear-negative PTB and EP forms</td>
<td>• Automated molecular test</td>
<td>• Yes</td>
</tr>
<tr>
<td></td>
<td>• Culture or molecular techniques</td>
<td>• Yes</td>
</tr>
<tr>
<td></td>
<td>• X-rays</td>
<td>• Yes</td>
</tr>
<tr>
<td></td>
<td>• Others (e.g. ADA, Pandy, Rivalta, FNAC)</td>
<td>• Yes</td>
</tr>
<tr>
<td></td>
<td>• Algorithms used</td>
<td></td>
</tr>
<tr>
<td>DST</td>
<td>DST possible (methods, quality control)</td>
<td>Detection of DR-TB</td>
</tr>
<tr>
<td>Treatment support</td>
<td>Number of patients receiving treatment support/month</td>
<td>100% of those eligible for support</td>
</tr>
<tr>
<td>Identification of non-adherent patients</td>
<td>• System for identifying and looking for non-adherent patients</td>
<td>• Yes</td>
</tr>
<tr>
<td></td>
<td></td>
<td>• &gt; 90%</td>
</tr>
<tr>
<td>Percentage of patients who resumed treatment among those missing for less than 2 months who had to be looked for</td>
<td></td>
<td></td>
</tr>
<tr>
<td>---</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Integrated TB/HIV care</strong></td>
<td><strong>Access to voluntary counselling and testing (VCT)</strong></td>
<td><strong>Yes</strong></td>
</tr>
<tr>
<td></td>
<td><strong>Access to ART</strong></td>
<td><strong>Yes</strong></td>
</tr>
<tr>
<td></td>
<td><strong>Access to cotrimoxazole prophylaxis</strong></td>
<td><strong>Yes</strong></td>
</tr>
<tr>
<td></td>
<td><strong>HIV treatment integrated in the TB service (or TB treatment in the HIV service)</strong></td>
<td><strong>Yes</strong></td>
</tr>
</tbody>
</table>

### 17.5.2 Procedures
<table>
<thead>
<tr>
<th>Criteria</th>
<th>Indicators</th>
<th>Goals</th>
</tr>
</thead>
<tbody>
<tr>
<td>Registers/records</td>
<td>Description of the documents</td>
<td>Records reliable</td>
</tr>
<tr>
<td></td>
<td>• Consistency between TB registers and treatment cards</td>
<td>• 100%</td>
</tr>
<tr>
<td></td>
<td>• Consistency between TB register and lab registers</td>
<td>• 100%</td>
</tr>
<tr>
<td>Standard case definitions</td>
<td>Percentage of patients with exact case definition out of a randomized sample of patients</td>
<td>100%</td>
</tr>
<tr>
<td>Adequate standard treatment regimens and follow-up</td>
<td>• Percentage of new cases correctly treated (combinations, dosage, duration) out of a randomized sample of patients</td>
<td>&gt; 95%</td>
</tr>
<tr>
<td></td>
<td>• Percentage of patients who did not have bacteriological follow-up according to schedule out of a randomized sample of patients</td>
<td>&lt; 10%</td>
</tr>
<tr>
<td></td>
<td>• Percentage of MDR-TB patients who did not have biochemistry tests according to schedule out of a randomized sample of patients</td>
<td>&lt; 10%</td>
</tr>
<tr>
<td>HIV testing</td>
<td>Percentage of new cases tested for HIV</td>
<td>100%</td>
</tr>
<tr>
<td>ART</td>
<td>Percentage of HIV-positive TB cases started on ART</td>
<td>100%</td>
</tr>
<tr>
<td></td>
<td>ART started within:</td>
<td></td>
</tr>
<tr>
<td></td>
<td>• &lt; 2 weeks; 2 weeks-&lt; 2 months; ≥ 2 months</td>
<td></td>
</tr>
<tr>
<td>Criteria for cure</td>
<td>Percentage of confirmed cases declared cured who actually met the definition of cure out of a randomized sample of patients</td>
<td>&gt; 90%</td>
</tr>
<tr>
<td>Regular monitoring of drug-susceptible TB and DR-TB</td>
<td>• Quarterly report and cohort analysis for drug-susceptible TB</td>
<td>Quantitative data on inclusions and results collected</td>
</tr>
<tr>
<td></td>
<td>• Bi-annual report and cohort analysis for DR-TB</td>
<td>Rapid detection of potential problems</td>
</tr>
<tr>
<td>Adherence monitoring</td>
<td>• Percentage of patients coming in for their appointment out of number of patients expected</td>
<td>&gt; 90% in both the intensive and continuation phases</td>
</tr>
<tr>
<td></td>
<td>• Percentage of doses given under DOT for DR-TB treatment in a randomized sample of patients</td>
<td>100%</td>
</tr>
<tr>
<td>Prevention of <em>M. tuberculosis</em> airborne transmission in TB facilities</td>
<td>• Isolation</td>
<td>Isolation of smear positive patients</td>
</tr>
<tr>
<td></td>
<td>• Building ventilation, lights, UV lamps (hospital wards, outpatient clinics, laboratory); respirators for staff and visitors in contact with contagious</td>
<td>Isolation of DR smear positive patients</td>
</tr>
<tr>
<td></td>
<td>• Appropriate use of means</td>
<td>Appropriate use of means</td>
</tr>
</tbody>
</table>
17.5.3 Human resources

<table>
<thead>
<tr>
<th>Criteria</th>
<th>Indicators</th>
<th>Goals</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staff</td>
<td>• Job descriptions (doctors, nurses, lab technicians, cleaning staff, etc.)</td>
<td>On average:</td>
</tr>
<tr>
<td></td>
<td>• Medical staff-to-patient ratio</td>
<td>• One nurse for 10-15 patients</td>
</tr>
<tr>
<td></td>
<td>Refer to training programme evaluation criteria</td>
<td>• One doctor for 40-50 patients</td>
</tr>
<tr>
<td>Training</td>
<td>Refer to training programme evaluation criteria</td>
<td>Competent staff</td>
</tr>
<tr>
<td>Other contributors</td>
<td>Description: other NGOs, local associations, etc.</td>
<td></td>
</tr>
</tbody>
</table>

A grid for evaluating TB clinic operations can be found in Appendix 35. Each criterion is rated either “satisfactory” or “unsatisfactory”.

patients; masks for contagious patients (if they move about)
- Written prevention plan?
- Person in charge identified?
- Yes
- Yes

<table>
<thead>
<tr>
<th>Standard precautions</th>
<th>Description</th>
<th>Standard precautions followed</th>
</tr>
</thead>
</table>
| Laboratory quality control | • Regular evaluation of laboratory functioning  
• Quarterly EQA of smear microscopy  
• Annual EQA of DST  | • Ensure the quality of laboratory analyses for bacteriological diagnosis  
• Results according to standards  
• Results according to standards |

<table>
<thead>
<tr>
<th>Criteria</th>
<th>Indicators</th>
<th>Goals</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staff</td>
<td>• Job descriptions (doctors, nurses, lab technicians, cleaning staff, etc.)</td>
<td>On average:</td>
</tr>
<tr>
<td></td>
<td>• Medical staff-to-patient ratio</td>
<td>• One nurse for 10-15 patients</td>
</tr>
<tr>
<td></td>
<td>Refer to training programme evaluation criteria</td>
<td>• One doctor for 40-50 patients</td>
</tr>
<tr>
<td>Training</td>
<td>Refer to training programme evaluation criteria</td>
<td>Competent staff</td>
</tr>
<tr>
<td>Other contributors</td>
<td>Description: other NGOs, local associations, etc.</td>
<td></td>
</tr>
</tbody>
</table>
Appendices

Appendix 1. Xpert assays
Appendix 2. Interpretation of Xpert assay results
Appendix 3. Sputum specimen: collection, storage and shipment
Appendix 4. Sputum smear microscopy
Appendix 5. Time required for diagnostic test results
Appendix 6. Ventilated work station (VWS) and bio-safety cabinet (BSC)
Appendix 7. Lymph node fine needle aspiration
Appendix 8. Protein estimation
Appendix 9. Tuberculin skin test
Appendix 10. Drug information sheets and patient instructions for the treatment of tuberculosis

Tuberculosis drug information sheets

- Amikacin (Am)
- Amoxicillin/clavulanic acid ratio 4:1 (Amx/Clv)
- Bedaquiline (Bdq)
- Clofazimine (Cfz)
- Cycloserine (Cs) or terizidone (Trd)
- Delamanid (Dlm)
- Ethambutol (E)
- Ethionamide (Eto) or prothionamide (Pto)
- Imipenem/cilastatin (Ipm/Cln)
- Isoniazid - Standard dose (H)
- Isoniazid - High dose (Hh)
- Levofloxacin (Lfx)
- Linezolid (Lzd)
- Meropenem (Mpm)
- Moxifloxacin (Mfx)
- Para-aminosalicylate sodium (PAS)
- Pretomanid (Pa)
- Pyrazinamide (Z)
- Rifabutin (Rfb)
- Rifampicin (R)
- Rifapentine (P)
- Streptomycin (S)

Patient instructions

Patients on drug-susceptible TB treatment
Patients on drug-resistant TB treatment

Appendix 11. Use of TB drugs in pregnant or breastfeeding women

Appendix 12. Dose adjustments in renal insufficiency

Appendix 13. Daily dose of TB drugs using fixed-dose combinations

Appendix 14. Monitoring of patients on drug-susceptible TB treatment

Appendix 15. Monitoring of patients on drug-resistant TB treatment

Appendix 16. Additional investigations in drug-resistant TB

Appendix 16. Basic TB infection control risk assessment tool

Appendix 17. Management of adverse effects

Gastrointestinal disorders

Abdominal pain

Diarrhoea

Epigastric pain

Hepatotoxicity

Metallic taste

Nausea and vomiting

Neurotoxicity

Depression

Headache

Optic neuritis

Ototoxicity

Peripheral neuropathy

Psychosis

Seizures

Endocrine disorders

Gynecomastia

Hypothyroidism

Dermatological disorders

Alopecia

Fungal infection

Photosensitivity

Skin reactions

Musculoskeletal disorders

Arthralgias

Tendinitis/tendon rupture

Miscellaneous

Electrolyte disorders

Haematologic disorders
Lactic acidosis
Nephrotoxicity
QT prolongation

Appendix 17. Air change per hour (ACH) measurement recommendations
Appendix 18. Compassionate use
Appendix 18. Advantages and disadvantages of ventilation techniques
Appendix 19. Drug interactions and overlapping toxicities
Appendix 19. Upper room ultraviolet germicidal irradiation (UVGI) system
Appendix 20. Treatment supporters
Appendix 21. Patient therapeutic education
Appendix 23. Treatment card for patients on first-line anti-TB therapy
Appendix 24. Tuberculosis register for patients on first-line anti-TB therapy
Appendix 25. Treatment card for patients on second-line anti-TB therapy
Appendix 26. Tuberculosis register for patients on second-line anti-TB therapy
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Appendix 28. Surgical masks
Appendix 29. BCG vaccine
Appendix 29. Sputum smear microscopy register
Appendix 30. Xpert MTB/RIF register
Appendix 31. Drug-o-gram
Appendix 32. Quaterly report
Appendix 33. Report on detection and enrolment of TB cases with rifampicin and multidrug-resistance
Appendix 34. Request form for smear microscopy and Xpert assays
Appendix 34. Report of final outcomes of drug-resistant tuberculosis
Appendix 35. Request form for culture, pDST, LPA, genome sequencing
Appendix 35. Check-list for the evaluation of a TB service
Appendix 1. Xpert assays

Update: October 2022

1.1 Specimen processing

Staff members present during specimen preparation should wear a respirator (FFP2 or N95) to prevent the inhalation of bacilli. A biosafety cabinet (Appendix 6) should be used to protect staff from aerosols when the specimen is to be centrifuged or cut/ground.

1.1.1 Sputum specimens

See Xpert MTB/RIF package insert:

See Xpert MTB/XDR package insert:

1.1.2 Lymph node and other tissue specimens

If the Xpert assay is performed on a biopsy (adapted from WHO)[1]:

- Cut the tissue specimen in small pieces in a sterile mortar (or grinder).
- Add 2 ml of sterile phosphate buffer saline (PBS).
- Grind solution of tissue and PBS to obtain a homogeneous mixture.
- Transfer 0.7 ml of mixture into a centrifuge tube using a transfer pipette. Avoid transferring clumps that are not well homogenized.
- Add 1.4 ml of Xpert Sample Reagent (XSR) using a transfer pipette.
- Shake vigorously 10 to 20 times or vortex for at least 10 seconds.
- Keep at room temperature for 10 minutes.
- Shake vigorously 10 to 20 times or vortex for at least 10 seconds.
- Keep at room temperature for 5 minutes.
- Transfer 2 ml of the mixture to the Xpert cartridge using a transfer pipette.
- Load the cartridge into the Xpert instrument as per the manufacturer's instructions.

If the Xpert assay is performed on a lymph node specimen obtained by fine needle aspiration (FNA):

- Flush the needle and syringe into a sterile container containing 1 ml of sterile 0.9% sodium chloride or sterile PBS.
- Transfer 0.7 ml of mixture into a centrifuge tube using a transfer pipette.
- Add 1.4 ml of XSR using a transfer pipette.
- For the next steps, continue as above.

For lymph node fine needle aspiration technique see Appendix 7.

1.1.3 Cerebrospinal fluid specimens

Adapted from WHO[1]

The processing method for cerebrospinal fluid (CSF) depends on the volume available for testing.
Note: a volume of CSF less than 0.1 ml is insufficient for testing.

### 1.1.4 Stool specimens[2]

Stool specimens can be used within 3 hours if kept at room temperature.

- Add 0.8 to 1 g of stool into the 8 ml XSR bottle.
- Shake vigorously for 30 seconds.
- Keep at room temperature for 10 minutes.
- Shake vigorously for 30 seconds.
- Sediment at room temperature for 10 minutes.
- Without disturbing the sediment, transfer 2 ml of the supernatant into the Xpert cartridge.
- Load the cartridge into the Xpert instrument as per the manufacturer's instructions.

### 1.1.5 Urine specimens[3]

Urine specimens can be used within 3 hours if kept at room temperature.

- Centrifuge 4 ml of urine at 3,000g for 5 minutes.
- Pour the supernatant and add 2 ml of XSR to the sediment.
- Shake vigorously.
- Transfer 2 ml of the mixture into the Xpert cartridge.
- Load the cartridge into the Xpert instrument as per the manufacturer's instructions.

### 1.2 Diagnostic accuracy of Xpert in specimens other than sputum

<table>
<thead>
<tr>
<th>Volume of CSF</th>
<th>Procedure</th>
</tr>
</thead>
</table>
| 0.1 to 1 ml   | - Add XSR to the CSF to obtain a final volume of 2 ml.  
                - Transfer 2 ml of the mixture into the Xpert cartridge.  
                - Load the cartridge into the Xpert instrument as per the manufacturer's instructions. |
| 1 to 5 ml     | - Add an equal volume of XSR to the CSF.  
                - Add 2 ml of the mixture directly into the Xpert cartridge.  
                - Load the cartridge into the Xpert instrument as per the manufacturer's instructions. |
| > 5 ml        | - Centrifuge the CSF at 3,000g for 15 minutes.  
                - Pour the supernatant and add XSR to the sediment to obtain a final volume of 2 ml.  
                - Transfer 2 ml of the mixture into the Xpert cartridge.  
                - Load the cartridge into the Xpert instrument as per the manufacturer's instructions. |
### 1.3 Logistic requirements

All Xpert assays are performed with the same instrument.
The 10-colour module can read all Xpert cartridges.
The 6-colour module can read Xpert MTB/RIF and Xpert MTB/RIF Ultra cartridges.

#### 1.3.1 Power supply

The instrument requires a constant and stable power supply.
If power cuts are short (less than 10 minutes), use a 1500VA "on line" UPS.
If power cuts are long, the system must be able to sustain a full cycle (approximately 45 minutes). Use a battery charger, a stationary battery, and a voltage stabilizer.

#### 1.3.2 Storage and operating temperatures

Storage of cartridges and reagents: between 2 and 28 °C for 12 months from date of manufacture.
Operating temperature for the Xpert instrument: between 15 and 30 °C. According to climate conditions, air conditioning may be required.

#### 1.3.3 Calibration

<table>
<thead>
<tr>
<th>Specimens</th>
<th>Performances of Xpert MTB/RIF compared to culture[^4]</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lymph node biopsy or aspirate</td>
<td>Biopsy: sensitivity: 82%; specificity: 79%</td>
</tr>
<tr>
<td></td>
<td>Aspirate: sensitivity: 89%; specificity: 86%</td>
</tr>
<tr>
<td>CSF</td>
<td>Sensitivity: 70%; specificity: 97%</td>
</tr>
<tr>
<td>Pleural fluid</td>
<td>Sensitivity: 50%; specificity: 99%</td>
</tr>
<tr>
<td>Pericardial fluid</td>
<td>Sensitivity: 67.6%; specificity: 99.4%</td>
</tr>
<tr>
<td>Nasopharyngeal aspirate (children with suspected PTB)</td>
<td>Sensitivity: 46%; specificity: 100%</td>
</tr>
<tr>
<td>Gastric aspirate (children with suspected PTB)</td>
<td>Sensitivity: 73%; specificity: 98%</td>
</tr>
<tr>
<td>Stool (children with suspected PTB)</td>
<td>Compared to respiratory specimens' culture:</td>
</tr>
<tr>
<td></td>
<td>- No HIV infection: sensitivity: 61%; specificity: 98%</td>
</tr>
<tr>
<td></td>
<td>- HIV infection: sensitivity: 70%; specificity: 98%</td>
</tr>
<tr>
<td>Urine (suspected genitourinary TB)</td>
<td>Sensitivity: 85%; specificity: 97%</td>
</tr>
<tr>
<td>Urine (HIV patients with suspected disseminated TB)</td>
<td>Sensitivity: 40%; specificity: 98%[^5]</td>
</tr>
<tr>
<td>Synovial fluid</td>
<td>Sensitivity: 97%; specificity: 94%</td>
</tr>
<tr>
<td>Peritoneal fluid</td>
<td>Sensitivity: 59%; specificity: 97%</td>
</tr>
<tr>
<td></td>
<td>Adult: sensitivity: 56%; specificity: 94%</td>
</tr>
</tbody>
</table>

[^4]: the performances of Xpert MTB/XDR in non-sputum specimens are considered similar to those of Xpert MTB/RIF as the tests are based on similar technologies.

[^5]:

[^6]:
The Xpert modules require annual calibration performed by an authorized service provider or carried out by swapping out the modules. A detailed contract with the supplier should guarantee regular maintenance, calibration, repair, and replacement as and when needed.

### 1.3.4 Required space

The dimensions of the Xpert IV instrument (4 modules enabling the processing of 4 specimens at the same time) are:

- Width: 29.8 cm; height 35.6 cm; depth 31.1 cm; weight: 12 kg.

The instrument is designed for indoor use only. Provide at least 5 cm of clearance on each side to ensure adequate ventilation. Do not place the instrument close to the vents of other instruments or air-handling units.

The dimensions of the kits containing cartridges and reagents are:

- Xpert MTB/RIF kit 50 tests: 31 cm x 28 cm x 20 cm
- Xpert MTB/XDR kit 10 tests: 24 cm x 16 cm x 7 cm

### 1.3.5 Waste disposal

Same procedure as for sputum containers.

Xpert assays generate large volumes of waste.

---

**References**


Appendix 2. Interpretation of Xpert assay results

Update: October 2022

2.1 Xpert MTB/RIF and Xpert MTB/RIF Ultra

MTB: *M. tuberculosis*; RIF: rifampicin

<table>
<thead>
<tr>
<th>Results</th>
<th>Interpretation and decisions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Invalid/Error/No result</td>
<td>Perform a 2\textsuperscript{nd} test on a new specimen.</td>
</tr>
</tbody>
</table>
| MTB not detected                              | • Child with suspected PTB: perform a 2\textsuperscript{nd} test on a new (respiratory or stool) specimen.  
   • Adult: re-evaluate clinically, perform x-ray if indicated, perform a 2\textsuperscript{nd} test and/or a culture on a new specimen. |
| MTB detected                                  | • Treat for DS-TB.                                                                           |
| No RIF resistance detected                    | • Perform Xpert MTB/XDR, LPA or pDST to detect H resistance\textsuperscript{(h)}; adjust treatment according to DST. |
| MTB detected                                  | Evaluate risk factors for rifampicin resistance (RR):                                        |
| RIF resistance detected                       | • High risk of RR\textsuperscript{(b)}: treat for MDR/RR-TB.                                 |
|                                               | • Low risk of RR\textsuperscript{(c)}: perform a 2\textsuperscript{nd} test on a new specimen\textsuperscript{(b)}. If 2\textsuperscript{nd} test shows: |
|                                               |   • R susceptibility: treat for DS-TB.                                                         |
|                                               |   • R resistance: treat for MDR/RR-TB.                                                         |
| For patients with MDR/RR-TB, perform:         |                                                                                              |
|                                               | • Xpert MTB/XDR or LPA and pDST or genome sequencing for resistance to other TB drugs.         |
|                                               | • Culture and pDST for treatment monitoring.                                                   |
|                                               | • If discordant results with pDST (R resistance with Xpert, R susceptibility with pDST): treat for MDR/RR-TB\textsuperscript{[f]}. |
| MTB detected                                  | Xpert MTB/RIF:                                                                              |
| RIF resistance indeterminate                  | • Perform a 2\textsuperscript{nd} test on a new specimen. If still “indeterminate”, treat for DS-TB while investigating RR. |
|                                               | • Perform pDST or other gDST to confirm or rule out RR.                                      |
|                                               | • Perform Xpert MTB/XDR, LPA or pDST to detect H resistance\textsuperscript{(a)}.               |
|                                               | Xpert MTB/RIF Ultra:                                                                        |
|                                               | • Send an extraction of the raw results (gxx file) to a reference laboratory for identification of possible mutations (interpretation of melting curves). |
|                                               | • If not feasible or still “indeterminate”: proceed as for Xpert MTB/RIF.                     |
| MTB detected “trace” RIF resistance indeterminate (Xpert Ultra) | HIV-infected patients, children and EP specimens: a “trace” result should be considered as positive. |
|                                               | • Adults with history of TB in the previous 5 years: a “trace” result cannot be interpreted, culture should be performed. |
|                                               | • No interpretation of RR is possible.                                                        |
|                                               | • If suspected resistance to R or other TB drugs: perform pDST or other gDST. Adjust treatment according to DST. |
|                                               | • Do not test the specimen with Xpert MTB/XDR as the Xpert MTB/XDR has a higher detection limit than Ultra. |
(a) For all patients if possible, and at least those with high risk of H resistance (patients with previous TB treatment with H, or contact with a TB case resistant to H, or from an area with a prevalence of resistance to H ≥ 3%).

(b) Patients with previous TB treatment with R, or contact with a TB case resistant to R, or from an area of high prevalence of resistance to R.

(c) Patients with no previous TB treatment with R, or contact with a TB case resistant to R, and from an area of low prevalence of resistance to R.

(d) A 2nd test is necessary because in a population with a prevalence of resistance to rifampicin < 5%, the positive predictive value of one test is < 80%, i.e. > 20% of rifampicin resistant results are false positive.

### 2.2 Xpert MTB/XDR

**MTB**: *M. tuberculosis*; **RIF**: rifampicin; **INH**: isoniazid; **FLQ**: fluoroquinolones; **ETH**: ethionamide; **AMK**: amikacin; **KAN**: kanamycin; **CAP**: capreomycin

<table>
<thead>
<tr>
<th>Results</th>
<th>Interpretation and decisions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Invalid/Error/No result</td>
<td>Perform a 2nd test on a new specimen.</td>
</tr>
<tr>
<td>MTB detected</td>
<td>After a positive Xpert MTB/RIF, an “MTB detected” result is expected because Xpert MTB/XDR and Xpert MTB/RIF have similar detection limit.</td>
</tr>
</tbody>
</table>
| MTB not detected No resistance detected | • After a positive Xpert MTB/RIF: perform a 2nd test on a new specimen. If the 2nd test is negative, it can be performed on culture isolates.  
• After a “trace” result with Ultra, a negative result is expected because the Xpert MTB/XDR has a higher detection limit than Ultra. |
| MTB detected No resistance detected | • Treat according to the result of Xpert MTB/RIF or Ultra.  
Resistance cannot be ruled out because other resistance-conferring mutations are not detected by Xpert MTB/XDR (e.g. only 30% of Eto resistance conferring mutations are detected).  
• Perform pDST for resistance to other TB drugs and monitor treatment. |
| MTB detected Low INH resistance detected | Evaluate risk factors of resistance for each drug:  
• High risk of resistance: consider as resistant to the drug.  
  ▪ If low-level H resistance detected (inhA mutation and no katG mutation): H\(^h\) can be used, but not counted as a likely effective drug.  
  ▪ If low-level resistance to FQs detected: Mfx\(^h\) can be used, but not counted as a likely effective drug\(^[2]\).  
  ▪ Resistance to Eto can be detected (inhA mutation). However, a negative result does not rule out resistance.  
  ▪ Perform pDST for resistance to other TB drugs and monitor treatment.  
• Low risk of resistance: perform a 2nd test on a new specimen\(^[5]\). If the 2nd test shows:  
  ▪ Drug susceptibility: treat with the drug.  
  ▪ Drug resistance: consider as resistant (see above for “High risk of resistance to the drug”). |
| MTB detected AMK, KAN and/or CAP resistance detected | Perform a 2nd test on a new specimen. If still “indeterminate”: treat with likely effective drug(s) while investigating resistance with pDST or other gDST (second-line LPA, genome sequencing). |
No "indeterminate" result is given for Eto.

References


Appendix 3. Sputum specimen: collection, storage and shipment

3.1 Sputum collection techniques

Regardless the collection technique used, staff member present during sputum collection should wear a respirator to prevent bacilli inhalation.

3.1.1 Sputum obtained spontaneously

Two specimens are to be collected. When possible, specimens should be collected outside in the open air and far away from other people.

The first sample is collected on the spot, at the consultation, when the patient is identified as suspected TB case. If the patient has recently eaten, ask him/her to rinse his/her mouth with water in order to avoid the presence of food in the sample.

The second sample is collected the day after, in the early morning, right after the patient wakes up and before eating. The second sample may be collected at home then the patient brings it to the health facility.

Alternatively, two sputum specimens can be collected one hour apart (frontloaded microscopy).

Collection technique:
- The patient must be given a labelled sputum container (or a Falcon® tube, if the sample is to be shipped by air).
- Have the patient take a deep breath, hold for a few seconds, exhale, repeat two or three times, then cough: sputum is material brought up from the lungs after a productive cough. One or two minutes of chest clapping are of benefit.
- Collect at least 3 ml and close the container hermetically.

The quality of sample determines the reliability of the result. Always check that the sample contains solid or purulent material and not only saliva. Take a new sample if unsatisfactory.

If the sample is collected at home, make sure that the patient has understood the technique, including closing the container hermetically after collecting the sputum.

3.1.2 Sputum induction

Sputum induction is sometimes used in children when sputa cannot be spontaneously expectorated, and only in order to perform cultures or Xpert MTB/RIF.

Sputum induction must be performed under close medical supervision. The child should be observed for respiratory distress during, and for 15 minutes after, the procedure. Bronchospasm may occur. Salbutamol spray and oxygen must be ready at hand.

Equipment
- Gloves and respirator
- Suction catheter (6, 7, 8F)
- Sputum container
- 50 ml syringe, needle
- Mask and tubing for nebulizer
- Holding chamber with child’s mask (to be sterilized between each patient)
- Sterile hypertonic solution of 5% sodium chloride (to be kept refrigerated)
- Sterile solution of 0.9% sodium chloride (for the specimen)
- Salbutamol spray
- Oxygen

Procedure

The child should fast for at least 2 hours before the procedure.
- Prior to nebulization:
3.1.3 Gastric aspiration

Gastric aspiration is sometimes used in children when sputa cannot be spontaneously expectorated nor induced using hypertonic saline, and only in order to perform cultures or Xpert MTB/RIF.

**Equipment**

- Gloves and respirator
- Suction catheter (6, 7, 8F)
- Sputum container
- 50 ml syringe
- Sterile water

**Procedure**

- Prior to inserting the suction catheter:
  - Explain the procedure to the child and/or the person accompanying him/her (this person must wear a respirator);
  - Place the child in a half-sitting or sitting position in the adult’s arms.
- Insert a nasogastric tube and check that it is correctly placed.
- First suction to collect the gastric fluid and place it in the sputum container, then rinse the stomach with 30 ml of sterile water and suction again. Add the suctioned fluid to the first sample.
- Start culture within 4 hours of collecting the sample. If there will be more than four hours’ delay, neutralize with 100 mg of sodium bicarbonate.

3.2 Sputum specimen storage

When examinations are not performed on the site of collection:

**Specimen for smear microscopy**

Smears should be performed within three-four days of collection and in the meanwhile stored refrigerated (2 to 8 °C) and protected from light.
Contamination does not affect microscopy but heat make specimen liquefy, with selection of mucopurulent part of the sample more difficult.

**Specimen for culture in liquid medium**
Keep the specimen refrigerated (2 to 8 °C), protected from light. Do not use cetylpyrodinium chloride (CPC) as it is not compatible with MGIT.

The specimen should be processed as soon as possible.

**Specimen for culture on Lowenstein-Jensen medium (LJ)**

- Specimens that can be cultured in less than 3 days after collection:
  Keep refrigerated (2 to 8 °C) and protected from light until transport OR immediately transport to the laboratory for processing.

- Specimens that will be cultured more than 3 days after collection:
  Use Falcon tubes and add 1% CPC to preserve the specimen for up to 2 weeks. Specimens with CPC should not be refrigerated, as the CPC will crystallize and be ineffective.
  Samples with CPC can be inoculated on LJ. For inoculation on agar, they require prior neutralization by neutralizing buffer (Difco®).
  CPC can be used for specimens tested by Xpert MTB/RIF.

### 3.3 Sputum specimen shipment

**To a local laboratory**

- Without CPC transport medium: between 2 and 8 °C and protected from light;
- With CPC transport medium: should not be refrigerated because at low temperatures the CPC will crystallize and ruin the sample.
  Specimens should be kept at room temperature, protected from heat and light.

**By air to a reference laboratory for culture**

Samples are collected and shipped in 50 ml Falcon® conical tubes with screw caps. The tubes are labelled UN 3373, corresponding to Category B infectious substances. If transport times are less than 12 hours, even specimens without CPC can be transported at room temperature.

Samples are triple-packaged, in accordance with IATA packing instruction 650:
1. Primary container holding the sputum sample: tube tightly closed and placed into a latex glove;
2. Secondary container intended to protect the primary container: leak-proof box with enough absorbent material to absorb the entire sample, should the primary container break;
3. Outer packaging intended to protect the secondary container, with UN 3373 labelling.

Information to be provided:
- Primary container: label with the patient’s name or identification number and the sample collection date and location;
- Outer package: indicate the name of the receiving laboratory, the complete address (name, street, postal code, locality, country), and telephone number.
- All samples must be accompanied by the corresponding laboratory test request form (including clinical information).

**Notes:**
- Procedures for shipping bacterial strains obtained after culture are different, more complicated, and rarely feasible in practice.
  Cultures are classified as Category A infectious substances (UN 2814).
- For a detailed description of the shipment procedures, see MSF Medical catalogue, volume 4.
Appendix 4. Sputum smear microscopy

Update: January 2022

4.1 Sputum smear preparation

Staff members present during sputum smear preparation should wear a respirator to prevent the inhalation of bacilli. Sputum smears should be prepared promptly after sputum collection.

**Equipment**

- Gloves
- Respirator (FFP2 or N95)
- New, clean glass slides (never re-use sputum smear slides)
- Wooden applicator sticks

**Technique**

- Label one end of the slide with the date of sputum collection and laboratory serial number.
- Select a mucopurulent or blood-stained portion of the sputum specimen.
- Use an applicator stick to transfer to the slide.
- Smear the specimen over an area of 1.5 to 2 cm x 2 to 3 cm. Make it thin enough to be able to read through it.
- Allow the smear to air dry for 15 minutes. Do not dry the smear in direct sunlight or over a flame.
- Fix the smear by passing the underside of the slide through a flame for 2 to 3 seconds. Repeat 3 or 4 times.
- Allow to cool before staining.

4.2 Ziehl-Neelsen staining

**Equipment**

- Gloves
- Distilled or filtered water
- 0.3% carbol fuchsin
- 3% acid-alcohol
- 0.3% methylene blue
- Binocular microscope with oil immersion objective (100x magnification)

**Technique**

- Flood the slide with 0.3% carbol fuchsin (after filtering the carbol fuchsin).
- Gently heat the underside of the slide. Begin timing as soon as steam appears. Let it steam for 5 minutes. Do not let the stain boil or dry.
- Gently rinse the slide until the water runs clear, then drain off excess water.
- Flood the slide with 3% acid-alcohol for 2 to 3 minutes, then drain. Repeat this operation if the slide is not completely decolourised.
- Gently rinse the slide, then drain off excess water.
- Flood the slide with 0.3% methylene blue for one minute, then drain.
- Gently rinse the slide until the water runs clear, then drain off excess water.
- Allow to air dry. Do not wipe or blot.

**Reading**

- The slides should be examined by an experienced technician. Technicians must be given sufficient time to accurately read slides.
- Before reading the slide, apply a drop of immersion oil to the left edge of the stained smear. Do not touch the slide with the immersion oil applicator (risk of AFB transfer into the oil bottle and onto another slide).
- Examine at least one length (100 high power fields, HPF) before giving a negative result (this should take at least 5 minutes).
• AFB are red, straight or slightly curved rods. They may be found singly or in small groups. The background stains blue.

Reporting

Table 4.1 - Grading AFB scale (WHO-IUATLD)[1]

<table>
<thead>
<tr>
<th>Number of AFB (1000x magnification: one length = 100 HPF)</th>
<th>Reporting</th>
</tr>
</thead>
<tbody>
<tr>
<td>Zero AFB/one length</td>
<td>No AFB</td>
</tr>
<tr>
<td>1-9 AFB/one length or 100 HPF</td>
<td>Report exact number of AFB</td>
</tr>
<tr>
<td>10-99 AFB/one length or 100 HPF</td>
<td>1+</td>
</tr>
<tr>
<td>1-10 AFB/one HPF in at least 50 fields</td>
<td>2+</td>
</tr>
<tr>
<td>&gt; 10 AFB/one HPF in at least 20 fields</td>
<td>3+</td>
</tr>
</tbody>
</table>

Note: 1-9 AFB in 100 HPF is a positive result. Note that 1-9 AFB in 100 HPF is reported as “scanty” followed by the exact number of AFB seen in 100 HPF (e.g. “scanty 3” means there are 3 AFB in 100 HPF). Do not confuse “scanty 3” (3 AFB in 100 HPF) with AFB 3+ (more than 10 AFB per HPF).

4.3 Auramine O or auramine/rhodamine staining

Equipment

• Gloves
• Distilled or filtered (not chlorinated) water
• 0.1% auramine O or auramine/rhodamine solution
• 0.5% acid alcohol
• 0.5% potassium permanganate or 0.3% methylene blue
• Fluorescence microscope (or a LED device that can be attached to a standard light microscope)

Technique

• Flood the slide with auramine O or auramine/rhodamine solution for 15 minutes. Ensure that the staining solution remains on the smear.
• Gently rinse, then drain off excess water. Do not use chlorinated water to avoid disturbing the fluorescence reading.
• Flood the slide with 0.5% acid-alcohol for one minute, then drain.
• Gently rinse, then drain off excess water.
• Flood the slide with 0.5% potassium permanganate solution or 0.3% methylene blue for one minute, then drain.
• Gently rinse, then drain off excess water.
• Allow to air dry. Do not wipe or blot.

Note: to control the quality of the colouration include at least one known positive smear in the batch.

Reading

• The slides should be examined by an experienced technician (artefacts are frequent). Technicians must be given sufficient time to read slides.
• Use a 20x objective to screen the smear.
• Examine one length before giving a negative result.
• Always read the positive control smear first. If the positive control is not positive do not continue with the patient smears, but re-stain the batch.
AFB are bright yellow, straight or slightly curved rods. They may be found singly or in small groups. The background is dark. Non-specific debris stains pale yellow.

**Reporting**

**Table 4.2 - Grading AFB scale (WHO-IUATLD)**

<table>
<thead>
<tr>
<th>Number of AFB (200–250x magnification: one length = 300 HPF)</th>
<th>Reporting</th>
</tr>
</thead>
<tbody>
<tr>
<td>Zero AFB/one length</td>
<td>No AFB</td>
</tr>
<tr>
<td>1-29 AFB/one length</td>
<td>Report exact number of AFB</td>
</tr>
<tr>
<td>30-299 AFB/one length</td>
<td>1+</td>
</tr>
<tr>
<td>10-100 AFB/one field on average</td>
<td>2+</td>
</tr>
<tr>
<td>&gt;100 AFB/one field on average</td>
<td>3+</td>
</tr>
</tbody>
</table>

**Notes:**

- 1-29 AFB per length is a positive result. Note that 1-29 AFB per length is reported as “scanty” followed by the exact number of AFB seen per length (e.g. “scanty 3” means there are 3 AFB per length). Do not confuse “scanty 3” (3 AFB per length) with AFB 3+ (more than 100 AFB per field).
- The fluorescence stain remains stable when sheltered from light for only 3 days. Quality control should be done within this time.

**References**

# Appendix 5. Time required for diagnostic test results

**Update: October 2022**

<table>
<thead>
<tr>
<th>Test Type</th>
<th>Estimated time for results</th>
<th>Additional time for DST</th>
</tr>
</thead>
<tbody>
<tr>
<td>Xpert MTB/RIF</td>
<td>110 minutes</td>
<td>–</td>
</tr>
<tr>
<td>Xpert MTB/RIF Ultra</td>
<td>&lt; 80 minutes</td>
<td>–</td>
</tr>
<tr>
<td>Xpert MTB/XDR</td>
<td>&lt; 90 minutes</td>
<td>–</td>
</tr>
<tr>
<td>Truenat</td>
<td>35 minutes (Truenat MTB)</td>
<td>1 hour (Truenat MTB-RIF Dx)</td>
</tr>
<tr>
<td>Culture liquid medium (MGIT (a))</td>
<td>8 days (smear+)</td>
<td>2 weeks</td>
</tr>
<tr>
<td></td>
<td>16 days (smear−)</td>
<td></td>
</tr>
<tr>
<td>Culture solid medium LJ standard medium</td>
<td>16 days (smear+)</td>
<td>6 weeks</td>
</tr>
<tr>
<td></td>
<td>29 days (smear−)</td>
<td></td>
</tr>
<tr>
<td>Culture microcolonies (TLA (b), MODS (c))</td>
<td>14 days</td>
<td>–</td>
</tr>
<tr>
<td>Smear microscopy</td>
<td>2 hours</td>
<td>–</td>
</tr>
<tr>
<td>LPA GenoType MTBDRplus (V2.0)</td>
<td>1 to 2 days (direct testing on smear+)</td>
<td>–</td>
</tr>
<tr>
<td></td>
<td>21 days (indirect testing)</td>
<td></td>
</tr>
<tr>
<td>LPA GenoTypeMTBDRs/ (V2.0)</td>
<td>1 to 2 days (direct testing)</td>
<td>–</td>
</tr>
<tr>
<td></td>
<td>21 days (indirect testing)</td>
<td></td>
</tr>
<tr>
<td>LF-LAM</td>
<td>25 minutes</td>
<td>–</td>
</tr>
<tr>
<td>tNGS</td>
<td>1 to 3 days (direct testing on smear+)</td>
<td>–</td>
</tr>
<tr>
<td></td>
<td>21 days (indirect testing)</td>
<td></td>
</tr>
<tr>
<td>WGS</td>
<td>21 days (indirect testing)</td>
<td>–</td>
</tr>
</tbody>
</table>

(a) Mycobacteria growth indicator tube.
(b) Thin-layer agar.
(c) Microscopic observation of drug susceptibility.

**Note:** to provide negative results, cultures need to be incubated for 6 to 7 weeks on liquid media and 8 weeks on solid media.
Appendix 6. Ventilated work station (VWS) and bio-safety cabinet (BSC)

6.1 Ventilated workstation (VWS)

The VWS provides a safe work environment while preparing sputum smears for AFB staining and Xpert MTB/RIF. It is used when adequate natural ventilation cannot be achieved. Designed to be placed over a bench, it is constituted by a rectangular box ducted to the outside, where the duct is connected to an extraction fan.

VWS are used without filter and do not provide product protection. They should never be used for preparing cultures.

6.2 Class II BSC

A Class II BSC protects not only the operator and the environment, but also the material being manipulated inside the cabinet. The room air and the air circulating within the cabinet are drawn by a downward flowing current through a grate then, through a HEPA filter, which protects both the operator and the product. The air exiting the cabinet is filtered through a HEPA filter to protect the environment.

Class II BSCs are required for performing cultures.

Notes:
- Class I BSCs cannot be used for performing cultures and VWS are sufficient for preparing samples for microscopy and tests Xpert MTB/RIF. Therefore, their use is not recommended in this manual.
- Class III BSCs are generally not used for TB.
Appendix 7. Lymph node fine needle aspiration

FNAC is used to obtain material from lymph nodes. The material is expressed onto slides and prepared for examination. Two smears will be prepared with Giemsa stain to look for caseum, granuloma, giants cells, and epithelioid cells or histocytes and 1 or 2 will be prepared with Ziehl-Neelsen (ZN) stain to look for acid-fast bacilli (AFB).

Equipment

- Needle 23G (in very few cases, it would be possible to use 19G)
- 10 ml syringe
- 2 slides for Giemsa + one or 2 slides for ZN stain
- 10% povidone iodine, sterile gauze, gloves

Technique

- Disinfect the area.
- With the needle attached to the syringe, insert the needle deep into the lymph node.
- After the needle has entered the mass, pull back on the syringe plunger to create a vacuum.
- Rapidly move the needle in a to-and-fro fashion to allow material entering the needle.
- When blood or material appears in the needle hub the aspiration should be stopped. Try to aspirate as much as possible of materials, the amount of materials that has been aspirated would have effect on the specificity and sensitivity of diagnosis.
- Release the negative pressure before to take out the needle from the lymph node. Do not continue sucking while taking out the needle, this will avoid aspiration of materiel into the barrel of the syringe and avoid mixing the sample with the possible peripheral blood in the skin.

Slide preparation

Slide should be identified prior to the aspiration and prepared immediately after the aspiration.

- Detach the needle from the syringe immediately after the aspiration.
- Fill the syringe with air (needle is still detached).

Prepare the smear as follow:

- Giemsa
  - Reattach the needle to the syringe and carefully release one small drop of sample onto one end of the slide by pushing down the plunger of the syringe (if the drop is placed in the middle of slide it would be difficult to make smear afterwards).
  - Put another slide over the sample.
  - Slide the two slides against each other, in opposite directions, to spread the sample out completely between them. Do not press the slides together forcefully, to avoid crushing the cells.
  - Allow to air dry.
  - Fix the smears by methanol when they are completely dry.
  - Proceed to Giemsa staining.
- Ziehl-Neelsen
  - Place a small drop of ganglion aspirate on the slide.
  - Make a smear that is neither to thin or too thick.
  - Allow to air dry.
  - Fix the smear by flame when it is completely dry.
  - Proceed to ZN staining.

Reading after Giemsa staining

On each slide, one or several of the following aspects can be found:

- Caseation necrosis (caseum): a uniform, acellular, pinkish substance.
- Granuloma: cluster of epithelioid cells and lymphocytes scattered through out smear with or without caseous necrosis.
- Epithelioid cells: elongated, often semi-lunar cells with a fine granular nuclear chromatin surrounded by pink cytoplasm.
- Giant cells: huge multinuclear cells.
Notes:
- It would be better to look for granuloma and necrosis with the 10x and 40x power of microscope then to look for epithelioid cells and giant cells with 100x power.
- Observation of smear requires a competent reader with skills in cytology. Slides have to be sent to a referral cytopathology laboratory for quality control or confirmation.
- The quality of the specimen and the preparation are essential. The smear is to be done by skilled technicians.

Footnotes
(a) The golden standard of diagnosis for TB on tissue samples is hematoxylin-eosin stain, but Giemsa stain can be used as an alternative in remote areas with limited equipment.
Appendix 8. Protein estimation

Update: January 2022

8.1 Pandy test

Pandy test is used to detect an increase of protein in the cerebrospinal fluid (CSF).
The normal range of protein in CSF is 0.20 to 0.45 g/litre.
The Pandy test is positive when protein is superior to 0.45 g/litre.

Equipment

- Disposable gloves
- Pandy reagent
- Pasteur pipettes
- Conical centrifuge glass tube or test tube
- 1 ml pipettes

Preparation of 500 ml of Pandy reagent

Pandy is a saturated phenol solution.
- Weigh 30 g of phenol and transfer it into a 1000 ml bottle.
- Add 500 ml of distilled water and shake vigorously.
- Leave to stand for one 24 hours.
- Check that some phenol remains undissolved:
  - If so, filter: the solution is ready.
  - If all the phenol has dissolved, add a further 10 g of phenol and wait another 24 hours before filtering.

Pandy reagent is a highly corrosive and toxic solution:
- Label the bottle and mark it corrosive and poisonous.
- Wash hands after preparation.

Technique

- Place 1 ml of Pandy reagent in a centrifuge tube.
- Add 3 drops of CSF, drop by drop.
- After each drop, look for a white cloud in the tube.
- To facilitate the reading, place a black surface behind the tube.

Results

- Presence of a white precipitate: Pandy test
- Absence of a white precipitate: Pandy test

8.2 Rivalta test

The Rivalta test is used to detect an increase of protein in the body fluid (pleural fluid, ascites).
The test is positive when the proteins are superior to 30 g/litre.

Equipment

- Disposable gloves
- Rivalta reagent
- Pasteur pipettes
- Conical centrifuge glass tube or test tube
- 5 ml pipette
Preparation of 100 ml of Rivalta reagent

- Place 50 ml of distilled water in a 100 ml measuring cylinder.
- With a 5 ml pipette, add 3 ml of glacial acetic acid and make up to the 100 ml mark with the remaining 50 ml of distilled water.
- Transfer the solution into a bottle.

Technique

- Place 2 ml of Rivalta reagent in a centrifuge tube.
- Add 3 drops of pleural fluid/ascites, drop by drop.
- After each drop, look for a white cloud in the tube.
- To facilitate the reading, place a dark surface behind the tube.

Results

- Presence of a white precipitate: Rivalta test positive.
- Absence of a white precipitate: Rivalta test
Appendix 9. Tuberculin skin test

Update: January 2022

9.1 Introduction

A delayed hypersensitivity reaction occurs after an intradermal injection of tuberculin (tuberculin skin test, TST) in persons infected by *M. tuberculosis* or vaccinated with BCG.

The test is performed by injecting 5 international units of tuberculin (purified protein derivative, PPD) intradermally on the ventral surface of the forearm (side of forearm exposed with palm facing up).

The test, which should be performed by a trained healthcare worker, requires 2 visits. The reading is done on the second visit, 48 to 72 hours after the tuberculin injection.

If the patient does not return within 72 hours, another TST should be performed.

The result is determined by the diameter of the reaction and individual characteristics of the person being tested (Table 9.1). It should be recorded in millimetres, not as “positive” or “negative”.

The reaction is the area of induration (swelling that can be felt) around the injection site.

Using a ruler, the diameter of induration is measured transversely. The erythema (redness) around the indurated area is not the reaction and should not be measured.

A reaction that appears several minutes, hours or even 24 hours after injection, but disappears on the day after its appearance, is of no significance.

There is no correlation between the diameter of the induration and:

- likelihood of active TB,
- risk of developing active TB,
- protection against TB disease in vaccinated people.

9.2 Positive TST

A positive TST signifies that a *M. tuberculosis* infection has occurred.

However, TST cannot differentiate between active and latent infection.

A positive test supports the diagnosis of latent TB when other diagnostic tools have been used to rule out active TB.

In children, a positive TST may be one element among many to establish the diagnosis of active TB.

Table 9.1 - Positive TST results
A highly positive (induration diameter > 20 mm) or phlyctenular reaction should be considered as an argument in favour of active TB but is not enough to decide on treatment.

Some persons may have a positive TST result even if they have not been infected with *M. tuberculosis*. Causes of false positive results include:

- Errors in tuberculin administration
- Previous BCG vaccination
- Infection with non-tuberculosis mycobacteria
- Low specificity of TST

BCG is given at birth so previous BCG vaccination has limited impact on the interpretation of TST results, except in small children. The average diameter of the TST reaction 1 year after BCG vaccination is 10 mm, with extremes ranging from 4 to 20 mm. The reaction becomes weaker over time and disappears 5 to 10 years post-vaccination.

### 9.3 Negative TST

Usually, a negative TST result signifies that no *M. tuberculosis* infection has occurred. However, a negative TST result does not rule out TB infection.

Causes of false negative results include:

- Errors in tuberculin administration
- Recent viral illness or live virus vaccination (e.g. measles)
- Severe TB disease (e.g. TB meningitis or miliary TB)
- Recent (< 12 weeks) or very old (many years) TB infection
- Immuno depletion or a weak immune response (e.g. the very elderly, children < 5 years, malnutrition, patients taking corticosteroids or immunosuppressants)
- Persons with diseases that result in anergy (e.g. AIDS, haemopathy, sarcoidosis)
- Natural extinction of post-vaccination reaction from the 5th year following BCG

### Footnotes

[https://apps.who.int/iris/rest/bitstreams/1272664/retrieve](https://apps.who.int/iris/rest/bitstreams/1272664/retrieve)
Appendix 10. Drug information sheets and patient instructions for the treatment of tuberculosis

Update: November 2022

- **Tuberculosis drug information sheets**
  - Amikacin (Am)
  - Amoxicillin/clavulanic acid ratio 4:1 (Amx/Clv)
  - Bedaquiline (Bdq)
  - Clofazimine (Cfz)
  - Cycloserine (Cs) or terizidone (Trd)
  - Delamanid (Dlm)
  - Ethambutol (E)
  - Ethionamide (Eto) or prothionamide (Pto)
  - Imipenem/cilastatin (Ipm/Cln)
  - Isoniazid - Standard dose (H)
  - Isoniazid - High dose (H\(^2\))
  - Levofloxacin (Lfx)
  - Linezolid (Lzd)
  - Meropenem (Mpm)
  - Moxifloxacin (Mfx)
  - Para-aminosalicylate sodium (PAS)
  - Pretomanid (Pa)
  - Pyrazinamide (Z)
  - Rifabutin (Rfb)
  - Rifampicin (R)
  - Rifapentine (P)
  - Streptomycin (S)
- **Patient instructions**
  - Patients on drug-susceptible TB treatment
  - Patients on drug-resistant TB treatment
Tuberculosis drug information sheets

Update: November 2022

- Amikacin (Am)
- Amoxicillin/clavulanic acid ratio 4:1 (Amx/Clv)
- Bedaquiline (Bdq)
- Clofazimine (Cfz)
- Cycloserine (Cs) or terizidone (Trd)
- Delamanid (Dlm)
- Ethambutol (E)
- Ethionamide (Eto) or prothionamide (Pto)
- Imipenem/cilastatin (Ipm/Cln)
- Isoniazid - Standard dose (H)
- Isoniazid - High dose (H²)
- Levofloxacin (Lfx)
- Linezolid (Lzd)
- Meropenem (Mpm)
- Moxifloxacin (Mfx)
- Para-aminosalicylate sodium (PAS)
- Pretomanid (Pa)
- Pyrazinamide (Z)
- Rifabutin (Rfb)
- Rifampicin (R)
- Rifapentine (P)
- Streptomycin (S)
Amikacin (Am)

Update: January 2022

Forms, strengths and route of administration

- 500 mg amikacin base in 2 ml ampoule (250 mg/ml), for IM injection

Dosage

- Child and adult: 15 to 20 mg/kg once daily
- Patient 60 years and over: 15 mg/kg 3 times a week
- Maximum dose: 1000 mg daily
- Renal insufficiency: 12 to 15 mg/kg 2 or 3 times a week
| Weight (kg) | Daily dose (mg) | Daily dose (ml) - IM injection<sup>(a)</sup> (500 mg in 2 ml = 250 mg/ml) |
|------------|----------------|
| 5          | 75-100         | 0.4 ml         |
| 6          | 90-120         | 0.4 ml         |
| 7          | 105-140        | 0.6 ml         |
| 8          | 120-160        | 0.6 ml         |
| 9          | 135-180        | 0.6 ml         |
| 10         | 150-200        | 0.8 ml         |
| 11         | 165-220        | 0.8 ml         |
| 12         | 180-240        | 0.8 ml         |
| 13         | 195-260        | 1 ml           |
| 14         | 210-280        | 1 ml           |
| 15         | 225-300        | 1 ml           |
| 16         | 240-320        | 1.2 ml         |
| 17         | 255-340        | 1.2 ml         |
| 18         | 270-360        | 1.2 ml         |
| 19         | 285-380        | 1.5 ml         |
| 20         | 300-400        | 1.5 ml         |
| 21         | 315-420        | 1.5 ml         |
| 22         | 330-440        | 1.5 ml         |
| 23         | 345-460        | 1.5 ml         |
| 24         | 360-480        | 1.5 ml         |
| 25         | 375-500        | 2 ml           |
| 26         | 390-520        | 2 ml           |
| 27         | 405-540        | 2 ml           |
| 28         | 420-560        | 2 ml           |
| 29         | 435-580        | 2 ml           |
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to aminoglycosides.
- Administer with caution to patients 60 years and over or patients with pre-existing renal, vestibular, auditory or severe hepatic impairment.
- May cause:
  - nephrotoxicity, ototoxicity, electrolyte disturbances; rarely, hypersensitivity reactions;
  - local pain after injection.
- For the management of adverse effects see Appendix 17.
- Avoid or monitor combination with other ototoxic and/or nephrotoxic drugs (furosemide, amphotericin B, tenofovir, etc.).
- Pregnancy: CONTRA-INDICATED
- Breastfeeding: no contra-indication

Monitoring

- Symptomatic monitoring.
- Audiometry, serum creatinine and electrolytes (K, Ca, Mg).

Patient instructions

- Maintain a good fluid intake to limit renal problems.

Remarks

- Use a different site for each injection (absorption may be delayed if the same site is used repeatedly).

Storage

- Below 25 °C
Solution may darken from colourless to a pale yellow, but this does not indicate a loss of potency.

<table>
<thead>
<tr>
<th>30-35</th>
<th>625</th>
<th>2.5 ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>36-45</td>
<td>750</td>
<td>3 ml</td>
</tr>
<tr>
<td>46-55</td>
<td>875</td>
<td>3.5 ml</td>
</tr>
<tr>
<td>56-70</td>
<td>1000</td>
<td>4 ml</td>
</tr>
<tr>
<td>&gt; 70</td>
<td>1000</td>
<td>4 ml</td>
</tr>
</tbody>
</table>

(a) For doses less than 1 ml, use a 1 ml syringe graduated in 0.01 ml.
Amoxicillin/clavulanic acid ratio 4:1 (Amx/Clv)

Update: January 2022

Forms and strengths

- 500 mg amoxicillin/125 mg clavulanic acid tablet
- 250 mg amoxicillin/62.5 mg clavulanic acid per 5 ml, powder for oral suspension

Dosage (expressed in clavulanic acid)

- Child under 30 kg: 3 mg (0.25 ml)/kg of clavulanic acid 3 times daily, 60 minutes before each dose of meropenem
- Adolescent ≥ 15 years and ≥ 30 kg and adult: 125 mg of clavulanic acid 2 times daily, 60 minutes before each dose of carbapenem
- Maximum dose: 250 mg daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>500 mg/125 mg tablet</th>
<th>250 mg/62.5 mg per 5 ml oral suspension</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>50</td>
<td>–</td>
<td>1.3 ml x 3</td>
</tr>
<tr>
<td>6</td>
<td>60</td>
<td>–</td>
<td>1.5 ml x 3</td>
</tr>
<tr>
<td>7</td>
<td>70</td>
<td>–</td>
<td>2 ml x 3</td>
</tr>
<tr>
<td>8</td>
<td>80</td>
<td>–</td>
<td>2 ml x 3</td>
</tr>
<tr>
<td>9</td>
<td>90</td>
<td>–</td>
<td>2.5 ml x 3</td>
</tr>
<tr>
<td>10</td>
<td>100</td>
<td>–</td>
<td>2.5 ml x 3</td>
</tr>
<tr>
<td>11</td>
<td>110</td>
<td>–</td>
<td>3 ml x 3</td>
</tr>
<tr>
<td>12</td>
<td>120</td>
<td>–</td>
<td>3 ml x 3</td>
</tr>
<tr>
<td>13</td>
<td>130</td>
<td>–</td>
<td>3.5 ml x 3</td>
</tr>
<tr>
<td>14</td>
<td>140</td>
<td>–</td>
<td>3.5 ml x 3</td>
</tr>
<tr>
<td>15</td>
<td>150</td>
<td>–</td>
<td>4 ml x 3</td>
</tr>
<tr>
<td>16</td>
<td>160</td>
<td>–</td>
<td>4.5 ml x 3</td>
</tr>
<tr>
<td>17</td>
<td>170</td>
<td>–</td>
<td>4.5 ml x 3</td>
</tr>
<tr>
<td>18</td>
<td>180</td>
<td>–</td>
<td>5 ml x 3</td>
</tr>
<tr>
<td>19</td>
<td>190</td>
<td>–</td>
<td>5 ml x 3</td>
</tr>
<tr>
<td>20</td>
<td>200</td>
<td>–</td>
<td>5.5 ml x 3</td>
</tr>
<tr>
<td>21</td>
<td>210</td>
<td>–</td>
<td>5.5 ml x 3</td>
</tr>
<tr>
<td>22</td>
<td>220</td>
<td>–</td>
<td>6 ml x 3</td>
</tr>
<tr>
<td>23</td>
<td>230</td>
<td>–</td>
<td>6 ml x 3</td>
</tr>
<tr>
<td>24</td>
<td>240</td>
<td>–</td>
<td>6.5 ml x 3</td>
</tr>
<tr>
<td>25</td>
<td>250</td>
<td>–</td>
<td>6.5 ml x 3</td>
</tr>
<tr>
<td>26</td>
<td>250</td>
<td>–</td>
<td>6.5 ml x 3</td>
</tr>
<tr>
<td>27</td>
<td>250</td>
<td>–</td>
<td>6.5 ml x 3</td>
</tr>
<tr>
<td>28</td>
<td>250</td>
<td>–</td>
<td>6.5 ml x 3</td>
</tr>
<tr>
<td>29</td>
<td>250</td>
<td>–</td>
<td>6.5 ml x 3</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to penicillin-allergic patients and patients with history of hepatic disorders during a previous treatment with amoxicillin/clavulanic acid.
- Administer with caution to patients allergic to betalactams (cross-hypersensitivity may occur) and to patients with hepatic impairment.
- May cause: gastrointestinal disturbances (mainly diarrhoea), hypersensitivity reactions, hepatotoxicity.
- For the management of adverse effects see Appendix 17.
- Pregnancy: no contra-indication
- Breastfeeding: no contra-indication

Monitoring

- Symptomatic monitoring

Patient instructions

- Take with food.

Storage

🌞 – ☀ – Below 25 °C
- Powder for oral suspension: between 15 °C and 25 °C
- Once reconstituted, the oral suspension must be kept refrigerated (between 2 °C and 8 °C) and may be used for up to 7 days.
Bedaquiline (Bdq)

Update: January 2023

Forms and strengths

- 100 mg tablet
- 20 mg dispersible tablet

Dosage

- Child up to 15 kg: according to weight and age
- Child 16 to 29 kg: 200 mg once daily for 2 weeks, then 100 mg 3 times a week
- Child 30 kg and over and adult: 400 mg once daily for 2 weeks, then 200 mg 3 times a week

When administered 3 times a week, keep an interval of 48 hours between doses (Monday, Wednesday, Friday = M/W/F).

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Weeks 1 and 2</th>
<th>Subsequent weeks</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Dose (mg)</td>
<td>Once daily</td>
</tr>
<tr>
<td></td>
<td>100 mg tablet</td>
<td>20 mg dispersible tablet</td>
</tr>
<tr>
<td>5-6</td>
<td>30-60</td>
<td>&lt; 3 months: 1½ tab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>≥ 3 months: 3 tab</td>
</tr>
<tr>
<td>7-9</td>
<td>30-80</td>
<td>&lt; 3 months: 1½ tab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>≥ 3 months: 3 tab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>≥ 6 months: 4 tab</td>
</tr>
<tr>
<td>10-15</td>
<td>60-120</td>
<td>&lt; 6 months: 3 tab</td>
</tr>
<tr>
<td></td>
<td></td>
<td>≥ 6 months: 6 tab</td>
</tr>
<tr>
<td>16-29</td>
<td>200 tab</td>
<td>–</td>
</tr>
<tr>
<td>≥ 30</td>
<td>400 tab</td>
<td>–</td>
</tr>
</tbody>
</table>

- Alternatively, for children 16 to 29 kg: 10 dispersible tablets of 20 mg (200 mg) once daily on Weeks 1 and 2, then 5 dispersible tablets of 20 mg (100 mg) 3 times a week.

- If 20 mg dispersible tablets are not available, 100 mg tablets can be crushed and suspended in 10 ml of water or fruit juice to obtain a solution containing 10 mg of bedaquiline per ml, then administered as follows:
Contra-indications, adverse effects, precautions

- Do not administer (or discontinue) to patients with severe hepatic impairment, QTcF > 500 ms or clinically significant ventricular arrhythmia.
- Avoid or use with caution and under close monitoring in patients with:
  - history of syncopal episodes, * torsades de pointes, congenital QT prolongation;*
  - uncompensated heart failure, severe coronary artery disease, bradycardia;
  - electrolyte disturbances (correct first K, Ca, Mg), hypothyroidism (provide thyroxine);
  - severe renal impairment, end-stage renal disease (optimal dosing not established).
- May cause:
  - hepatotoxicity, moderate QT prolongation;
  - nausea, vomiting, arthralgia, headache, increased amylase level.
- For the management of adverse effects see Appendix 17.
- Avoid or use with caution and under close monitoring in patients taking CYP450 inducers/inhibitors, some ARVs, or other QT prolonging drugs (Appendix 19).

**Pregnancy:** use if benefits outweigh the risks (safety not established).

**Breastfeeding:** avoid breastfeeding during treatment (safety not established).

**Monitoring**

- Symptomatic monitoring.
- Liver function, ECG, electrolytes (K, Ca, Mg).

**Patient instructions**

- Take with food.
- 100 mg tablets can be crushed and mixed with water or fruit juice.
- 20 mg tablets should be dispersed in water, juice, milk, yogurt, porridge, etc.
- Avoid alcohol during treatment.

**Remarks**

- For patients over 14 years who receive the regimen BPaL or BPaLM, bedaquiline can be given daily instead of 3 times a week: 200 mg once daily for the first 8 weeks then, 100 mg once daily.

**Storage**

🌡️ – Below 25 °C
Clofazimine (Cfz)

Update: August 2022

Forms and strengths

- 50 mg and 100 mg soft capsules or tablets

Dosage

- Child under 10 kg: doses are administered 3 times a week (Monday, Wednesday, Friday = M/W/F)
- Child 10 to 29 kg: 2 to 5 mg/kg once daily
- Child 30 kg and over and adult: 100 mg once daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>100 mg capsule(a)</th>
<th>50 mg capsule(a)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>–</td>
<td>–</td>
<td>1 caps (M/W/F)</td>
</tr>
<tr>
<td>6</td>
<td>–</td>
<td>–</td>
<td>1 caps (M/W/F)</td>
</tr>
<tr>
<td>7</td>
<td>–</td>
<td>–</td>
<td>1 caps (M/W/F)</td>
</tr>
<tr>
<td>8</td>
<td>–</td>
<td>–</td>
<td>1 caps (M/W/F)</td>
</tr>
<tr>
<td>9</td>
<td>–</td>
<td>–</td>
<td>1 caps (M/W/F)</td>
</tr>
<tr>
<td>10</td>
<td>20-50</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>11</td>
<td>22-55</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>12</td>
<td>24-60</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>13</td>
<td>26-65</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>14</td>
<td>28-70</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>15</td>
<td>30-75</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>16</td>
<td>32-80</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>17</td>
<td>34-85</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>18</td>
<td>36-90</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>19</td>
<td>38-95</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>20</td>
<td>40-100</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>21</td>
<td>42-105</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>22</td>
<td>44-110</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>23</td>
<td>46-115</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>24</td>
<td>48-120</td>
<td>1 caps</td>
<td>–</td>
</tr>
<tr>
<td>25</td>
<td>50-125</td>
<td>1 caps</td>
<td>–</td>
</tr>
<tr>
<td>26</td>
<td>52-130</td>
<td>1 caps</td>
<td>–</td>
</tr>
<tr>
<td>27</td>
<td>54-135</td>
<td>1 caps</td>
<td>–</td>
</tr>
<tr>
<td>28</td>
<td>56-140</td>
<td>1 caps</td>
<td>–</td>
</tr>
<tr>
<td>29</td>
<td>58-145</td>
<td>1 caps</td>
<td>–</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with history of allergy to clofazimine.
- Administer with caution to patients with severe hepatic impairment.
- May cause:
  - orange-brown discoloration of skin and body fluids;
  - strong QT prolongation;
  - gastrointestinal intolerance (nausea, vomiting, abdominal pain);
  - severe abdomen pain, bowel obstruction, intestinal bleeding;
  - eye and skin dryness and irritation, hypersensitivity reactions, photosensitivity.
- For the management of adverse effects see Appendix 17.
- Avoid or use with caution and under close monitoring in patients taking other QT prolonging drugs (Appendix 19).
- Pregnancy: use only if benefits outweigh the risks (safety is not established).
- Breast-feeding: avoid breastfeeding during treatment (safety not established). If used, may cause breast milk discolouration and reversible skin discolouration in breastfed infants.

Monitoring

- Symptomatic monitoring.
- ECG.

Patient instructions

- Take with food to improve gastrointestinal tolerance.
- Protect your skin from sun.
- Harmless orange-brown discoloration of the skin and body fluids (urine, sweat, saliva, sputum, tears, breast milk, etc.). It is reversible but may take months to disappear after stopping treatment.

Storage

°C – Below 25 °C
Cycloserine (Cs) or terizidone (Trd)

Update: January 2022

Forms and strengths

- 250 mg and 125 mg capsules

Dosage

- Child under 30 kg: 7.5 to 10 mg/kg 2 times daily (or 15 to 20 mg/kg once daily if tolerated)
- Child 30 kg and over and adult: 5 to 7.5 mg/kg 2 times daily (or 10 to 15 mg/kg once daily if tolerated)
- Maximum dose: 1000 mg daily
- Renal insufficiency: 250 mg once daily or 500 mg 3 times a week
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>250 mg capsule</th>
<th>125 mg capsule</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>75-100</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>6</td>
<td>90-120</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>7</td>
<td>105-140</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>8</td>
<td>120-160</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>9</td>
<td>135-180</td>
<td>–</td>
<td>1 caps</td>
</tr>
<tr>
<td>10</td>
<td>150-200</td>
<td>–</td>
<td>1 caps x 2</td>
</tr>
<tr>
<td>11</td>
<td>165-220</td>
<td>–</td>
<td>1 caps x 2</td>
</tr>
<tr>
<td>12</td>
<td>180-240</td>
<td>–</td>
<td>1 caps x 2</td>
</tr>
<tr>
<td>13</td>
<td>195-260</td>
<td>–</td>
<td>1 caps x 2</td>
</tr>
<tr>
<td>14</td>
<td>210-280</td>
<td>–</td>
<td>1 caps x 2</td>
</tr>
<tr>
<td>15</td>
<td>225-300</td>
<td>–</td>
<td>1 caps x 2</td>
</tr>
<tr>
<td>16</td>
<td>240-320</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>17</td>
<td>255-340</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>18</td>
<td>270-360</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>19</td>
<td>285-380</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>20</td>
<td>300-400</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>21</td>
<td>315-420</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>22</td>
<td>330-440</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>23</td>
<td>345-460</td>
<td>–</td>
<td>1 caps (morning) + 2 caps (evening)</td>
</tr>
<tr>
<td>24</td>
<td>360-480</td>
<td>1 caps x 2</td>
<td>–</td>
</tr>
<tr>
<td>25</td>
<td>375-500</td>
<td>1 caps x 2</td>
<td>–</td>
</tr>
<tr>
<td>26</td>
<td>390-520</td>
<td>1 caps x 2</td>
<td>–</td>
</tr>
<tr>
<td>27</td>
<td>405-540</td>
<td>1 caps x 2</td>
<td>–</td>
</tr>
<tr>
<td>28</td>
<td>420-560</td>
<td>1 caps x 2</td>
<td>–</td>
</tr>
<tr>
<td>29</td>
<td>435-580</td>
<td>1 caps x 2</td>
<td>–</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Avoid in patients with epilepsy, depression, psychosis, severe anxiety, history of neurological or psychiatric disorders, chronic alcohol use. However, if essential to the regimen, it can be administered under close monitoring.
- May cause:
  - neurotoxicity: seizure, headache, lethargy, confusion, mood change, drowsiness, anxiety, psychosis, depression, suicidal ideation, peripheral neuropathy; rarely, vestibular toxicity;
  - hypersensitivity reactions.
- For the management of adverse effects see Appendix 17.
- Avoid or monitor combination with isoniazid and thionamides (increased risk of neurotoxicity).
- Administer concomitantly pyridoxine (vitamin B₆); child: 1 to 2 mg/kg (usual range: 10 to 50 mg) once daily; adult: 100 mg once daily.
- Pregnancy: use if the benefits outweigh the risks. Administer pyridoxine to the mother (as above).
- Breastfeeding: no contra-indication. Administer pyridoxine to the mother (as above) and the breast-fed neonate or infant (1 to 2 mg/kg once daily).

Monitoring

- Symptomatic monitoring.

Patient instructions

- Take capsules with water before or after meals.
- Avoid alcohol during treatment.

Remarks

- To increase tolerance, start with a low dose (e.g. 250 mg daily in adults), then increase over 1 to 2 weeks to achieve the requested dose.

Storage

°C – Below 25 °C
Delamanid (Dlm)

Update: September 2022

Forms and strengths

- 50 mg tablet
- 25 mg dispersible tablet

Dosage

- Child under 10 kg: according to weight and age
- Child 10 to 15 kg: 25 mg 2 times daily
- Child 16 to 29 kg: 50 mg morning and 25 mg evening
- Child 30 to 45 kg and under 15 years: 50 mg 2 times daily
- Child 46 kg and over and adult: 100 mg 2 times daily

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>50 mg tablet</th>
<th>25 mg dispersible tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-9</td>
<td>25-50</td>
<td>–</td>
<td>&lt; 3 months: 1 tab</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>≥ 3 months: 1 tab x 2</td>
</tr>
<tr>
<td>10-15</td>
<td>50</td>
<td>–</td>
<td>1 tab x 2</td>
</tr>
<tr>
<td>16-29</td>
<td>75</td>
<td>–</td>
<td>2 tab (morning) + 1 tab (evening)</td>
</tr>
<tr>
<td>30-45</td>
<td>100-200</td>
<td>&lt; 15 years: 1 tab x 2</td>
<td>≥ 15 years: 2 tab x 2</td>
</tr>
</tbody>
</table>

If 25 mg dispersible tablets are not available, 50 mg tablets can be crushed and suspended in 10 ml of water or fruit juice to obtain a solution of 5 mg of delamanid per ml, administered as follows:

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>50 mg tablet in 10 ml (5 mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-9</td>
<td>25-50</td>
<td>&lt; 3 months: 5 ml</td>
</tr>
<tr>
<td></td>
<td></td>
<td>≥ 3 months: 5 ml x 2</td>
</tr>
<tr>
<td>10-15</td>
<td>50</td>
<td>5 ml x 2</td>
</tr>
<tr>
<td>16-29</td>
<td>75</td>
<td>10 ml (morning) + 5 ml (evening)</td>
</tr>
</tbody>
</table>

Contra-indications, adverse effects, precautions

- Do not administer (or discontinue) to patients with QTcF > 500 ms or albumin level < 2.8 g/dl.
- Avoid or use with caution and under close monitoring in patients with:
  - history of syncopal episodes or torsades de pointes, congenital QT prolongation, cardiac disease;
  - electrolyte disturbances (correct first K, Ca, Mg);
  - severe renal or hepatic impairment.
- Use with caution and under close monitoring in patients taking QT-prolonging drugs (Appendix 19).
• May cause: nausea, vomiting, dizziness, insomnia, mild QT prolongation.
• For the management of adverse effects see Appendix 17.

**Pregnancy:** use if benefits outweigh the risks (safety not established).

**Breastfeeding:** avoid breastfeeding during treatment (safety not established).

**Monitoring**

• Symptomatic monitoring.
• ECG.

**Patient instructions**

• Take with food.
• 50 mg tablets should be swallowed whole if possible.
• 25 mg tablets should be dispersed in water or fruit juice.

**Storage**

🌡 – Below 25 °C
Ethambutol (E)

Update: January 2022

Forms and strengths

- 100 mg and 400 mg tablets
- 100 mg dispersible tablet, to be dispersed in 10 ml water

Dosage

- Child and adult: 15 to 25 mg/kg once daily
- Maximum dose: 1200 mg daily
- Renal insufficiency: 15 to 25 mg/kg 3 times a week
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>400 mg tablet</th>
<th>100 mg tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>75-125</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>6</td>
<td>90-150</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>7</td>
<td>105-175</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>8</td>
<td>120-200</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>9</td>
<td>135-225</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>10</td>
<td>150-250</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>11</td>
<td>165-275</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>12</td>
<td>180-300</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>13</td>
<td>195-325</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>14</td>
<td>210-350</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>15</td>
<td>225-375</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>16</td>
<td>240-400</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>17</td>
<td>255-425</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>18</td>
<td>270-450</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>19</td>
<td>285-475</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>20</td>
<td>300-500</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>21</td>
<td>315-525</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>22</td>
<td>330-550</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>23</td>
<td>345-575</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>24</td>
<td>360-600</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>25</td>
<td>375-625</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>26</td>
<td>390-650</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>27</td>
<td>405-675</td>
<td>1½ tab</td>
<td>–</td>
</tr>
<tr>
<td>28</td>
<td>420-700</td>
<td>1½ tab</td>
<td>–</td>
</tr>
<tr>
<td>29</td>
<td>435-725</td>
<td>1½ tab</td>
<td>–</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with severe renal impairment or pre-existing optic neuritis (e.g. diabetic retinopathy).
- May cause: dose-related retrobulbar optic neuritis, exacerbated in renal impairment.
- The dosage must be carefully adjusted to the weight, especially for children under 5 years, as it is more difficult to detect visual changes at this age.
- For the management of adverse effects see Appendix 17.
- Pregnancy: no contra-indication
- Breastfeeding: no contra-indication

Monitoring

- Symptomatic monitoring.

Patient instructions

- Take with or without food.
- 100 mg dispersible tablets should be dispersed in 10 ml water.

Remarks

- For adults on drug-susceptible TB treatment, ethambutol is given as part of a fixed-dose combination.
- Ethambutol is also used in the treatment of drug-resistant TB treatment for longer duration. For treatment > 2 months, daily doses should be closer to 15 mg/kg and visual acuity and colour discrimination should be monitored.

Storage

- Below 25 °C
Ethionamide (Eto) or prothionamide (Pto)

Update: January 2022

Forms and strengths

- 250 mg tablet (ethionamide or prothionamide)
- 125 mg dispersible tablet (ethionamide), to be dispersed in 10 ml water

Dosage

- Child and adult: 15 to 20 mg/kg once daily
- Maximum dose: 1000 mg daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>250 mg tablet</th>
<th>125 mg dispersible tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>75-100</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>6</td>
<td>90-120</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>7</td>
<td>105-140</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>8</td>
<td>120-160</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>9</td>
<td>135-180</td>
<td>–</td>
<td>1½ tab</td>
</tr>
<tr>
<td>10</td>
<td>150-200</td>
<td>–</td>
<td>1½ tab</td>
</tr>
<tr>
<td>11</td>
<td>165-220</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>12</td>
<td>180-240</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>13</td>
<td>195-260</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>14</td>
<td>210-280</td>
<td>–</td>
<td>2½ tab</td>
</tr>
<tr>
<td>15</td>
<td>225-300</td>
<td>–</td>
<td>2½ tab</td>
</tr>
<tr>
<td>16</td>
<td>240-320</td>
<td>–</td>
<td>2½ tab</td>
</tr>
<tr>
<td>17</td>
<td>255-340</td>
<td>–</td>
<td>2½ tab</td>
</tr>
<tr>
<td>18</td>
<td>270-360</td>
<td>–</td>
<td>2½ tab</td>
</tr>
<tr>
<td>19</td>
<td>285-380</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>20</td>
<td>300-400</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>21</td>
<td>315-420</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>22</td>
<td>330-440</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>23</td>
<td>345-460</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>24</td>
<td>360-480</td>
<td>–</td>
<td>3 tab</td>
</tr>
<tr>
<td>25</td>
<td>375-500</td>
<td>2 tab</td>
<td>–</td>
</tr>
<tr>
<td>26</td>
<td>390-520</td>
<td>2 tab</td>
<td>–</td>
</tr>
<tr>
<td>27</td>
<td>405-540</td>
<td>2 tab</td>
<td>–</td>
</tr>
<tr>
<td>28</td>
<td>420-560</td>
<td>2 tab</td>
<td>–</td>
</tr>
<tr>
<td>29</td>
<td>435-580</td>
<td>2 tab</td>
<td>–</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>----</td>
<td>----</td>
<td>----</td>
<td>----</td>
</tr>
<tr>
<td>30-35</td>
<td>500</td>
<td>2 tab</td>
<td></td>
</tr>
<tr>
<td>36-45</td>
<td>500</td>
<td>2 tab</td>
<td></td>
</tr>
<tr>
<td>46-55</td>
<td>750</td>
<td>3 tab</td>
<td></td>
</tr>
<tr>
<td>56-70</td>
<td>750</td>
<td>3 tab</td>
<td></td>
</tr>
<tr>
<td>&gt; 70</td>
<td>1000</td>
<td>4 tab</td>
<td></td>
</tr>
</tbody>
</table>

Contra-indications, adverse effects, precautions

- Do not administer to patients with severe hepatic impairment.
- Administer with caution to patients with hepatic disease, diabetes or depression.
- May cause:
  - frequently: gastrointestinal disturbances (abdominal or epigastric pain, diarrhoea, metallic taste, nausea and vomiting, stomatitis, etc.);
  - occasionally: endocrine disorders (gynecomastia, hypothyroidism), alopecia, depression, anxiety, psychosis, hypoglycaemia, vestibular disorders, hepatotoxicity, peripheral neuropathy, optic neuritis, hypersensitivity reactions, seizures
- For the management of adverse effects see Appendix 17.
- Monitor combination with: cycloserine or terizidone (increased risk of seizures) and para-aminosalicylic acid (increased risk of gastrointestinal disturbances and hypothyroidism).
- Administer concomitantly pyridoxine (vitamin $B_6$); child: 1 to 2 mg/kg (usual range: 10 to 50 mg) once daily; adult: 100 mg once daily.
- **Pregnancy:** CONTRA-INDICATED
- **Breastfeeding:** administer pyridoxine to the mother (as above). Observe the breast-fed neonate or infant for adverse effects and supplement it with pyridoxine (1 to 2 mg/kg once daily).

Monitoring

- Symptomatic monitoring.
- Liver function and thyroid function.

Patient instructions

- Take with food and/or at bedtime to limit gastrointestinal disturbances.
- 125 mg tablets should be dispersed in 10 ml water.
- Avoid alcohol during treatment.

Remarks

- To improve tolerance, start with a low dose (e.g. 250 mg daily in adults), then increase over 1 to 2 weeks to achieve the requested dose.
- For the 6HRZEt regimen for drug-susceptible TB meningitis, the dose is 20 mg/kg once daily (max. 750 mg daily).

Storage

☀ – Below 25 °C
Imipenem/cilastatin (Ipm/Cln)

Update: January 2022

Forms, strengths and route of administration

- Powder for injection, in vial of 500 mg imipenem monohydrate/500 mg cilastatin sodium, to be reconstituted with 20 ml of 0.9% sodium chloride (25 mg imipenem/ml).
- Each dose is to be diluted in 100 ml of 0.9% sodium chloride and to be administered by IV infusion:
  - over 30 minutes for doses ≤ 500 mg/500 mg
  - over 60 minutes for doses > 500 mg/500 mg
- Use a deep line, preferably an implantable venous access device (Port-a-Cath).

Dosage (expressed in imipenem)

- Adolescent 15 years and over (and ≥ 30 kg) and adult: 1000 mg (2 vials) 2 times daily with 10 hours minimum between infusions
- Maximum dose: 2000 mg daily
- Renal insufficiency: 750 mg every 12 hours for CrCl 20-40 ml/minute; 500 mg every 12 hours for CrCl < 20 ml/minute

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>Daily dose (ml) - IV infusion (500 mg/500 mg per vial)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-29</td>
<td></td>
<td>Do not used in patients &lt; 15 years and &lt; 30 kg</td>
</tr>
<tr>
<td>30-33</td>
<td>2000</td>
<td>2 vials (40 ml) in 100 ml of 0.9% NaCl x 2</td>
</tr>
<tr>
<td>34-40</td>
<td>2000</td>
<td>2 vials (40 ml) in 100 ml of 0.9% NaCl x 2</td>
</tr>
<tr>
<td>41-45</td>
<td>2000</td>
<td>2 vials (40 ml) in 100 ml of 0.9% NaCl x 2</td>
</tr>
<tr>
<td>46-50</td>
<td>2000</td>
<td>2 vials (40 ml) in 100 ml of 0.9% NaCl x 2</td>
</tr>
<tr>
<td>51-70</td>
<td>2000</td>
<td>2 vials (40 ml) in 100 ml of 0.9% NaCl x 2</td>
</tr>
<tr>
<td>&gt; 70</td>
<td>2000</td>
<td>2 vials (40 ml) in 100 ml of 0.9% NaCl x 2</td>
</tr>
</tbody>
</table>

Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to carbapenems.
- Administer with caution to patients allergic to other betalactams (cross-hypersensitivity may occur).
- May cause:
  - nausea, vomiting (the infusion rate may be slowed down in case of nausea), diarrhoea;
  - neurotoxicity: confusional state, seizures (most frequently in patients with history of seizures or renal impairment);
  - hypersensitivity reactions;
  - local reactions (phlebitis/thrombophlebitis).
- For the management of adverse effects see Appendix 17.
- Avoid or monitor combination with: valproic acid (decreased plasma concentration of valproic acid and risk of seizures), oral or injectable ganciclovir (risk of seizures).
- Pregnancy and breastfeeding: avoid unless the benefits outweigh the risks.
**Monitoring**

- Symptomatic monitoring.

**Remarks**

- Administer clavulanic acid 60 minutes before each dose of imipenem/cilastatin.
- Do not mix with Ringer lactate (incompatibility) but may be administered via Y-site.
- Do not mix with other drugs in the infusion bag.

**Storage**

- Below 25 °C
- Once reconstituted, solution:
  - remains stable 4 hours at room temperature or 24 hours between 2 to 8 °C;
  - may darken from colourless to yellow (this does not indicate a loss of potency);
  - should be discarded if it becomes brown.
Isoniazid - Standard dose (H)

Update: January 2022

Forms and strengths

- 300 mg and 100 mg tablets
- 100 mg and 50 mg dispersible tablets, to be dispersed in 10 ml water

Dosage

- Child under 30 kg: 10 mg/kg (7 to 15 mg/kg) once daily
- Child 30 kg and over and adult: 5 mg/kg (4 to 6 mg/kg) once daily
- Maximum dose: 300 mg daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>300 mg tablet</th>
<th>100 mg tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>35-75</td>
<td>–</td>
<td>½ tab</td>
</tr>
<tr>
<td>6</td>
<td>42-90</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>7</td>
<td>49-105</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>8</td>
<td>56-120</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>9</td>
<td>63-135</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>10</td>
<td>70-150</td>
<td>–</td>
<td>1½ tab</td>
</tr>
<tr>
<td>11</td>
<td>77-165</td>
<td>–</td>
<td>1½ tab</td>
</tr>
<tr>
<td>12</td>
<td>84-180</td>
<td>–</td>
<td>1½ tab</td>
</tr>
<tr>
<td>13</td>
<td>91-195</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>14</td>
<td>98-210</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>15</td>
<td>105-225</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>16</td>
<td>112-240</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>17</td>
<td>119-255</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>18</td>
<td>126-270</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>19</td>
<td>133-285</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>20</td>
<td>140-300</td>
<td>–</td>
<td>2 tab</td>
</tr>
<tr>
<td>21</td>
<td>147-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>22</td>
<td>154-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>23</td>
<td>161-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>24</td>
<td>168-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>25</td>
<td>175-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>26</td>
<td>182-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>27</td>
<td>189-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>28</td>
<td>196-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
<tr>
<td>29</td>
<td>203-300</td>
<td>1 tab</td>
<td>–</td>
</tr>
</tbody>
</table>
Alternatively, 50 mg dispersible tablets may be used instead of ½ tablets of 100 mg.

**Contra-indications, adverse effects, precaution**

- Do not administer to patients with severe hepatic impairment.
- May cause:
  - peripheral neuropathy;
  - hepatotoxicity;
  - hypersensitivity reactions, arthragias, optic neuritis, psychotic reactions, seizures and depression.
- Monitor closely:
  - pregnant and breastfeeding women, patients with renal impairment or diabetes; malnourished or HIV-infected patients (increased risk of neuropathy);
  - alcoholic patients (increased risk of neuropathy and hepatotoxicity);
  - patients with chronic hepatic disease or taking rifampicin or ≥ 35 years (increased risk of hepatotoxicity);
  - patients taking anticonvulsants, benzodiazepines (risk of toxicity), warfarin (risk of bleeding). Dose adjustment may be required.
- For the management of adverse effects see Appendix 17.
- Administer concomitantly pyridoxine (vitamin B6) to patients at risk of peripheral neuropathy (child: 5 to 10 mg once daily; adult: 10 mg once daily).
- **Pregnancy and breastfeeding**: no contra-indication. Administer pyridoxine to the mother (as above) and the breast-fed neonate or infant (5 mg once daily).

**Monitoring**

- Symptomatic monitoring.
- Liver function in patients with hepatic disease.

**Patient instructions**

- Take without food.
- 100 mg dispersible tablet should be dispersed in 10 ml water.
- Avoid alcohol during treatment.

**Remarks**

- For patients on drug-susceptible TB treatment, isoniazid is given as part of a fixed-dose combination.
- For the 6HRZ-Eto regimen for drug-susceptible TB meningitis, the dose of isoniazid is 20 mg/kg once daily (max. 400 mg daily).
- Isoniazid is also used in the treatment of latent TB infection and multidrug-resistant TB treatment (at high dose - H3).

**Storage**

![Solar symbol] - [Cold symbol] - Below 25 °C
Isoniazid - High dose (Hh)

Update: November 2022

Forms and strengths

- 300 mg and 100 mg tablets
- 100 mg and 50 mg dispersible tablets, to be dispersed in 10 ml water

Dosage

- Child under 30 kg: 15 to 20 mg/kg once daily
- Child 30 kg and over and adult: 10 to 15 mg/kg once daily
- Maximum dose: 600 mg daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>300 mg tablet</th>
<th>100 mg tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>75-100</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>6</td>
<td>90-120</td>
<td>–</td>
<td>1 tab</td>
</tr>
<tr>
<td>7</td>
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<td>29</td>
<td>435-580</td>
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<td>4½ tab</td>
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</tbody>
</table>
Alternatively, 50 mg dispersible tablets may be used instead of ½ tablets of 100 mg.

**Contra-indications, adverse effects, precautions**

- Do not administer to patients with severe hepatic impairment.
- May cause:
  - peripheral neuropathy;
  - hepatotoxicity;
  - hypersensitivity reactions, arthragias, optic neuritis, psychotic reactions, seizures and depression.
- Monitor closely:
  - pregnant and breastfeeding women, patients with renal impairment or diabetes; malnourished or HIV-infected patients (increased risk of neuropathy);
  - alcoholic patients (increased risk of neuropathy and hepatotoxicity);
  - patients with chronic hepatic disease or taking rifampicin or ≥ 35 years (increased risk of hepatotoxicity);
  - patients taking anticonvulsants, benzodiazepines (risk of toxicity), warfarin (risk of bleeding). Dose adjustment may be required.
- For the management of adverse effects see Appendix 17.
- Administer concomitantly pyridoxine (vitamin B₆): child: 1 to 2 mg/kg (usual range: 10 to 50 mg) once daily; adult: 100 mg once daily.
- **Pregnancy and breastfeeding:** no contra-indication. Administer pyridoxine to the mother (as above). Observe the breast-fed neonate or infant for adverse effects and supplement it with pyridoxine (1 to 2 mg/kg once daily).

**Monitoring**

- Symptomatic monitoring.
- Liver function.

**Patient instructions**

- Take without food.
- Dispersible tablets should be dispersed in 10 ml water.
- Avoid alcohol during treatment.

**Storage**

![Storage](below 25 °C)
Levofloxacin (Lfx)

Update: September 2022

Forms and strengths

- 250 mg and 500 mg tablets
- 100 mg dispersible tablet, to be dispersed in 10 ml water

Dosage

- Child under 30 kg: 15 to 20 mg/kg once daily
- Child 30 kg and over and adult: 750 to 1000 mg once daily
- Maximum dose: 1500 mg daily
- Renal insufficiency: 750 to 1000 mg 3 times a week
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>500 mg tablet</th>
<th>250 mg tablet</th>
<th>100 mg dispersible tablet</th>
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</thead>
<tbody>
<tr>
<td>5</td>
<td>75-100</td>
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<td>29</td>
<td>435-580</td>
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<td>2 tab</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity or tendon damage during a previous treatment with a fluoroquinolone.
- Administer with caution to patients:
  - over 60 years or on corticosteroid treatment (increased risk of tendon damage);
  - with diabetes or history of mental disorders or seizures.
- May cause:
  - tendinitis, tendon rupture, mild QT prolongation;
  - gastrointestinal disturbances (abdominal or epigastric pain, diarrhoea);
  - neurological disorders (headache, psychosis, seizures, etc.);
  - photosensitivity;
  - hypersensitivity reactions, hypo/hyperglycaemia;
  - rarely: crystalluria, peripheral neuropathy, ototoxicity.
- For the management of adverse effects see Appendix 17.
- Avoid or use with caution and under close monitoring in patients taking other QT prolonging drugs (Appendix 19) or warfarin.
- Do not administer simultaneously with: antacids containing magnesium/aluminium, calcium, iron and zinc salts (administer 2 hours apart).

Pregnancy: use if benefits outweigh the risks (safety not established).
Breastfeeding: avoid breastfeeding during treatment (no absolute contra-indication).

Monitoring

- Symptomatic monitoring.

Patient instructions

- Take 2 hours apart from milk-based product, antacids, calcium, iron and zinc salts.
- 100 mg tablets should be dispersed in 10 ml water.
- Maintain a good fluid intake.
- Protect your skin from sun.

Storage

- Below 25 °C
Linezolid (Lzd)

Update: October 2022

Forms and strengths
- 600 mg tablet (breakable and non-breakable)
- 150 mg dispersible tablet
- 100 mg/5 ml, granules for oral suspension

Dosage
- Child under 15 kg: 15 mg/kg once daily
- Child 15 to 45 kg: 10 to 12 mg/kg once daily
- Patient 46 kg and over: 600 mg once daily
- Maximum dose: 600 mg daily

Note: in the BPaLM therapeutic regimen for patients 15 years and over, the dose of linezolid is 600 mg once daily for 16 weeks then 300 mg once daily up to the end of treatment.

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>600 mg tablet</th>
<th>150 mg dispersible tablet</th>
<th>100 mg per 5 ml oral suspension</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>75</td>
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<td></td>
<td>3 ml</td>
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<td>6</td>
<td>90</td>
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<td></td>
<td>4 ml</td>
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<td>7</td>
<td>105</td>
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<td></td>
<td>5 ml</td>
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<td>8-9</td>
<td>120-135</td>
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<td>6 ml</td>
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<td>10-15</td>
<td>150-180</td>
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<td>16-23</td>
<td>160-276</td>
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<td>1½ tab</td>
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<td>24-29</td>
<td>240-348</td>
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<td>30-35</td>
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<td>36-45</td>
<td>450</td>
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<td>3 tab</td>
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<td>46-55</td>
<td>600</td>
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<td>56-70</td>
<td>600</td>
<td>1 tab</td>
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<td>600</td>
<td>1 tab</td>
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</tbody>
</table>

Alternatively, for children 5 to 6 kg, if oral suspension is not available: one half of a 150 mg dispersible tablet (75 mg) once daily.

If 150 mg dispersible tablets are not available, 600 mg tablets can be crushed and suspended in 10 ml of water or fruit juice to obtain a solution of 60 mg of linezolid per ml, administered as follows:
### Weight (mg) | Daily dose (mg) | 600 mg tablet in 10 ml (60 mg/ml)
--- | --- | ---
5 | 75 | 1.25 ml
6 | 90 | 1.5 ml
7-9 | 105-135 | 2 ml
10-15 | 150-180 | 2.5 ml

**Contra-indications, adverse effects, precautions**

- Administer with caution to patients with haematological disorders or hypertension.
- May cause:
  - anaemia, neutropenia and/or thrombocytopenia;
  - lactic acidosis;
  - peripheral neuropathy (can be irreversible); rarely, optic neuritis;
  - abdominal pain, diarrhoea, nausea.
- For the management of adverse effects see Appendix 17.
- Avoid or monitor combination with serotonergic drugs such as tricyclic antidepressants (e.g. amitriptyline) or selective serotonin reuptake inhibitors (e.g. fluoxetine, paroxetine): risk of serotonin syndrome.
- Administer concomitantly pyridoxine (vitamin B₆); child: 1 to 2 mg/kg (usual range: 10 to 50 mg) once daily; adult: 100 mg once daily.
- **Pregnancy**: use if the benefits outweigh the risks. Administer pyridoxine to the mother (as above).
- **Breastfeeding**: avoid breastfeeding during treatment (safety not established).

**Monitoring**

- Symptomatic monitoring.
- Full blood count.
- Visual acuity and colour discrimination.

**Patient instructions**

- Take with or without food.

**Storage**

> ☀ – ☁ – Below 25 °C

Once reconstituted, the oral suspension may be kept at room temperature for 21 days, protected from light.
Meropenem (Mpm)

Update: January 2022

Forms, strengths and route of administration

- Powder for injection, in 500 mg vial, to be reconstituted with 10 ml of water for injection (50 mg meropenem/ml).
- Each dose is to be diluted in 5 ml/kg of 0.9% sodium chloride in children under 20 kg and in 100 ml of 0.9% sodium chloride in children 20 kg and over and adults and to be administered by IV infusion over 15 to 30 minutes.
- Use a deep line, preferably an implantable venous access device (Port-a-Cath).

Dosage

- Child under 30 kg: 20 to 40 mg/kg every 8 hours
- Child 30 kg and over and adult: 1500 to 2000 mg 2 times daily with 10 hours minimum between infusions
- Maximum dose: 6000 mg daily
- Renal insufficiency: 750 mg every 12 hours for CrCl 20-40 ml/minute; 500 mg every 12 hours for CrCl < 20 ml/minute
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>Daily dose (ml) – IV infusion (500 mg per vial)</th>
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</thead>
<tbody>
<tr>
<td>5</td>
<td>300</td>
<td>2 ml in 25 ml of 0.9% NaCl x 3</td>
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<td>6</td>
<td>300</td>
<td>2 ml in 30 ml of 0.9% NaCl x 3</td>
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<tr>
<td>7</td>
<td>600</td>
<td>4 ml in 35 ml of 0.9% NaCl x 3</td>
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<td>8</td>
<td>600</td>
<td>4 ml in 40 ml of 0.9% NaCl x 3</td>
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<td>9</td>
<td>600</td>
<td>4 ml in 45 ml of 0.9% NaCl x 3</td>
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<tr>
<td>10</td>
<td>900</td>
<td>6 ml in 50 ml of 0.9% NaCl x 3</td>
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<td>11</td>
<td>900</td>
<td>6 ml in 55 ml of 0.9% NaCl x 3</td>
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<td>12</td>
<td>900</td>
<td>6 ml in 60 ml of 0.9% NaCl x 3</td>
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<td>13</td>
<td>900</td>
<td>6 ml in 65 ml of 0.9% NaCl x 3</td>
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<td>14</td>
<td>900</td>
<td>6 ml in 70 ml of 0.9% NaCl x 3</td>
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<td>15</td>
<td>900</td>
<td>6 ml in 75 ml of 0.9% NaCl x 3</td>
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<td>16</td>
<td>1200</td>
<td>8 ml in 80 ml of 0.9% NaCl x 3</td>
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<td>17</td>
<td>1200</td>
<td>8 ml in 85 ml of 0.9% NaCl x 3</td>
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<td>18</td>
<td>1200</td>
<td>8 ml in 90 ml of 0.9% NaCl x 3</td>
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<td>19</td>
<td>1200</td>
<td>8 ml in 95 ml of 0.9% NaCl x 3</td>
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<td>20</td>
<td>1200</td>
<td>8 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>21</td>
<td>1200</td>
<td>8 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>22</td>
<td>1200</td>
<td>8 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>23</td>
<td>1200</td>
<td>8 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>24</td>
<td>1650</td>
<td>11 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>25</td>
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<td>11 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>1650</td>
<td>11 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>1650</td>
<td>11 ml in 100 ml of 0.9% NaCl x 3</td>
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<td>29</td>
<td>1650</td>
<td>11 ml in 100 ml of 0.9% NaCl x 3</td>
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</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to carbapenems.
- Administer with caution to patients allergic to cephalosporins (cross-hypersensitivity may occur).
- May cause:
  - nausea, vomiting (the infusion rate may be slowed down in case of nausea), diarrhoea;
  - neurotoxicity: confusional state, seizures (rarely compared to imipenem/cilastatin, most frequently in patients with history of seizures or renal impairment);
  - hypersensitivity reactions;
  - local reactions (phlebitis/thrombophlebitis).
- For the management of adverse effects see Appendix 17.
- Avoid or monitor combination with valproic acid (decreased concentration of valproic acid and risk of seizures).
- Pregnancy and breastfeeding: avoid unless the benefits outweigh the risks.

Monitoring

- Symptomatic monitoring.

Remarks

- Administer clavulanic acid 60 minutes before each dose of meropenem.
- Do not mix with other drugs in the infusion bag.

Storage

- Below 25 °C

Once reconstituted, solution should be used immediately (within 1 hour of preparation)
Moxifloxacin (Mfx)

Update: September 2022

Forms and strengths

- 400 mg tablet
- 100 mg dispersible tablet, to be dispersed in 10 ml water

Dosage (standard dose)

- Child under 30 kg: 10 to 15 mg/kg once daily
- Child 30 kg and over and adult: 400 mg once daily
- Maximum dose: 400 mg daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>400 mg tablet</th>
<th>100 mg dispersible tablet</th>
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<td>5</td>
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</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity or tendon damage during a previous treatment with a fluoroquinolone.
- Administer with caution to patients:
  - over 60 years or on corticosteroid treatment (increased risk of tendon damage);
  - with diabetes or history of mental disorders or seizures.
- May cause:
  - tendinitis, tendon rupture, moderate QT prolongation;
  - gastrointestinal disturbances (abdominal or epigastric pain, diarrhoea);
  - neurological disorders (headache, psychosis, seizures, etc.);
  - photosensitivity;
  - hypersensitivity reactions, hypo/hyperglycaemia;
  - rarely: crystalluria, peripheral neuropathy, ototoxicity.
- For the management of adverse effects see Appendix 17.
- Avoid or use with caution and under close monitoring in patients taking other QT prolonging drugs (Appendix 19) or warfarin.
- Do not administer simultaneously with: antacids containing magnesium/aluminium, calcium, iron and zinc salts (administer 2 hours apart).
- Pregnancy:
  - DR-TB: use if benefits outweigh the risks (safety not established).
  - DS-TB: do not use.
- Breastfeeding:
  - DR-TB: avoid breastfeeding during treatment (no absolute contra-indication).

Monitoring

- Symptomatic monitoring

Patient instructions

- Take 2 hours apart from milk-based product, antacids, calcium, iron and zinc salts.
- 100 mg tablets should be dispersed in 10 ml water.
- Maintain a good fluid intake.
- Protect your skin from sun.

Remarks

- Higher dose moxifloxacin \( (Mfx^h) \), i.e. 600 to 800 mg once daily in patients over 30 kg may be used in the presence of certain mutations conferring low level fluoroquinolone resistance. \( Mfx^h \) may cause strong QT prolongation.

Storage

\[ \text{☀ – ☑ – Below 25 °C} \]
Para-aminosalicylate sodium (PAS)

Update: October 2022

Forms and strengths

- Powder for oral solution, 5.52 g sachet of para-aminosalicylate sodium (equivalent to 4 g PAS acid), to be dissolved in 100 ml water.

Dosage (expressed in PAS acid)

- Child under 30 kg: 100 to 150 mg/kg 2 times daily
- Child 30 kg and over and adult: 4 g 2 times daily (max. 12 g daily)
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>Oral solution or sachet PAS sodium</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>1000-1500</td>
<td>19 ml x 2</td>
</tr>
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<td>6</td>
<td>1200-1800</td>
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</tr>
<tr>
<td>7</td>
<td>1400-2100</td>
<td>25 ml x 2</td>
</tr>
<tr>
<td>8</td>
<td>1600-2400</td>
<td>25 ml x 2</td>
</tr>
<tr>
<td>9</td>
<td>1800-2700</td>
<td>25 ml x 2</td>
</tr>
<tr>
<td>10</td>
<td>2000-3000</td>
<td>50 ml x 2</td>
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<td>11</td>
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</tr>
<tr>
<td>12</td>
<td>2400-3600</td>
<td>50 ml x 2</td>
</tr>
<tr>
<td>13</td>
<td>2600-3900</td>
<td>50 ml x 2</td>
</tr>
<tr>
<td>14</td>
<td>2800-4200</td>
<td>50 ml x 2</td>
</tr>
<tr>
<td>15</td>
<td>3000-4500</td>
<td>50 ml x 2</td>
</tr>
<tr>
<td>16</td>
<td>3200-4800</td>
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<td>17</td>
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<td>21</td>
<td>4200-6300</td>
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<td>23</td>
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<td>26</td>
<td>5200-7800</td>
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<td>27</td>
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</tr>
<tr>
<td>28</td>
<td>5600-8000</td>
<td>80 ml x 2</td>
</tr>
<tr>
<td>29</td>
<td>5800-8000</td>
<td>80 ml x 2</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Avoid in patients with severe renal disease.
- Avoid or use with caution in patients with hepatic impairment or gastric ulcer.
- May cause:
  - frequent gastrointestinal disturbances (nausea, vomiting, gastritis, diarrhoea);
  - hypothyroidism, hepatotoxicity, hypersensitivity reactions.
- Monitor combination with ethionamide/prothionamide (increased risk of gastrointestinal disturbances and hypothyroidism).
- For the management of adverse effects see Appendix 17.
- Pregnancy: use only if benefits outweigh the risks (safety not established).
- Breastfeeding: avoid breastfeeding during treatment (safety not established).

Monitoring

- Symptomatic monitoring.
- Liver and thyroid function.

Patient instructions

- Mix the powder with 100 ml water.
- Take with food to limit gastrointestinal disturbances.

Remarks

- To increase gastrointestinal tolerance, start with a low dose, e. g. for an adult: 2 g 2 times daily for 1 to 2 weeks, then 4 g 2 times daily.

Storage

- Below 25 °C
Pretomanid (Pa)

Update: October 2022

Forms and strengths

- 200 mg tablet

Dosage

- Adolescent 15 years and over and adult: 200 mg once daily, in combination with:
  - bedaquiline, linezolid and moxifloxacin (BPaLM)
  - bedaquiline, linezolid and clofazimine (BPaLC)
  - bedaquiline and linezolid (BPaL)
- Maximum dose: 200 mg daily

<table>
<thead>
<tr>
<th>Age</th>
<th>Daily dose (mg)</th>
<th>200 mg tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 15 years</td>
<td>Do not administer</td>
<td>–</td>
</tr>
<tr>
<td>≥ 15 years</td>
<td>200</td>
<td>1</td>
</tr>
</tbody>
</table>

Contra-indications, adverse effects, precautions

- Do not administer if one of the drugs included in the regimen is contraindicated.
- The contribution of pretomanid to the adverse effects of pretomanid-containing regimens is not determined.
- For adverse effects of companion drugs see individual drug information sheets.
- Pregnancy: use if benefits outweigh the risks (safety not established).
- Breastfeeding: avoid breastfeeding during treatment (safety not established).

Monitoring

- Symptomatic monitoring.
- For monitoring of companion drugs see individual drug information sheets.

Patient instructions

- Take with food.

Storage

- Below 25 °C
Pyrazinamide (Z)

Update: January 2022

Forms and strengths

- 400 mg tablet
- 150 mg dispersible tablet, to be dispersed in 10 ml water

Dosage

- Child under 30 kg: 35 mg/kg (30 to 40 mg/kg) once daily
- Child 30 kg and over and adult: 25 mg/kg (20 to 30 mg/kg) once daily
- Maximum dose: 2000 mg daily
- Renal insufficiency: 25 mg/kg 3 times a week
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>400 mg tablet</th>
<th>150 mg dispersible tablet</th>
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<td>450-600</td>
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<td>3 tab</td>
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<td>16</td>
<td>480-640</td>
<td></td>
<td>4 tab</td>
</tr>
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<td>17</td>
<td>510-680</td>
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<td>18</td>
<td>540-720</td>
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<td>4 tab</td>
</tr>
<tr>
<td>19</td>
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</tr>
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<td>720-960</td>
<td>2½ tab</td>
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</tr>
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<td>27</td>
<td>810-1080</td>
<td>2½ tab</td>
<td>–</td>
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<tr>
<td>28</td>
<td>840-1120</td>
<td>2½ tab</td>
<td>–</td>
</tr>
<tr>
<td>29</td>
<td>870-1160</td>
<td>2½ tab</td>
<td>–</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to pyrazinamide, severe hepatic impairment or severe gout.
- May cause: gout and arthralgias, hepatotoxicity, gastrointestinal disturbances (epigastric pain, nausea and vomiting), hypersensitivity reactions; rarely, photosensitivity.
- For the management of adverse effects see Appendix 17.
- **Pregnancy:** no contra-indication
- **Breastfeeding:** no contra-indication

Monitoring

- Symptomatic monitoring.
- Liver function in patients with hepatic impairment or under drug-resistant TB treatment.

Patient instructions

- Take with or without food.
- 150 mg tablets should be dispersed in 10 ml water.
- Protect your skin from sun.

Remarks

- For patients on drug-susceptible TB treatment, pyrazinamide is given as part of a fixed-dose combination.
- For the 6HRZ-Eto regimen for drug-susceptible TB meningitis, the dose of pyrazinamide is 40 mg/kg once daily (max. 2000 mg daily).

Storage

- Below 25 °C
Rifabutin (Rfb)

Update: January 2022

Forms and strengths

- 150 mg capsule

Dosage

- Child and adult: 5 to 10 mg/kg once daily
- Maximum dose: 300 mg daily
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>150 mg capsule</th>
</tr>
</thead>
<tbody>
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<td>70-140</td>
<td></td>
</tr>
<tr>
<td>15</td>
<td>75-150</td>
<td>1 caps</td>
</tr>
<tr>
<td>16</td>
<td>80-160</td>
<td>1 caps</td>
</tr>
<tr>
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<td>85-170</td>
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<td>1 caps</td>
</tr>
<tr>
<td>19</td>
<td>95-190</td>
<td>1 caps</td>
</tr>
<tr>
<td>20</td>
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</tr>
<tr>
<td>21</td>
<td>105-210</td>
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</tr>
<tr>
<td>22</td>
<td>110-220</td>
<td>1 caps</td>
</tr>
<tr>
<td>23</td>
<td>115-230</td>
<td>1 caps</td>
</tr>
<tr>
<td>24</td>
<td>120-240</td>
<td>1 caps</td>
</tr>
<tr>
<td>25</td>
<td>125-250</td>
<td>1 caps</td>
</tr>
<tr>
<td>26</td>
<td>130-260</td>
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<td>1 caps</td>
</tr>
<tr>
<td>29</td>
<td>145-290</td>
<td>1 caps</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to rifamycins or history of severe haematological disorders (thrombocytopenia, purpura) during a previous treatment with a rifamycin.
- Administer with caution to patients with severe renal impairment or hepatic or haematological disorders.
- May cause:
  - gastrointestinal disturbances, hepatotoxicity;
  - haematological disorders (leukopenia, anaemia, thrombocytopenia), hypersensitivity reactions;
  - reversible uveitis.
- For the management of adverse effects see Appendix 17.
- Reduce the dose of rifabutin:
  - in patients taking boosted protease inhibitors (Appendix 19);
  - if rifabutin toxicity is suspected in patients taking clarithromycin, fluconazole or itraconazole.
- Rifabutin reduces the effect of many drugs (macrolides, some antiretrovirals, some hormones, warfarin, etc.):
  - in patients taking antiretrovirals see Appendix 19;
  - in women using contraception, use injectable medroxyprogesterone or an intrauterine device;
  - for the other drugs, adjust dosage if necessary.
- **Pregnancy and breastfeeding:** avoid (safety not established). If used in late pregnancy, administer phytomenadione (vitamin K₁) to the mother and the neonate.

Monitoring

- Symptomatic monitoring.
- Liver function in patients with hepatic disease.
- Full blood count.

Patient instructions

- Take with or without food.
- Harmless orange-red discoloration of the urine, faeces, sweat, saliva, sputum, tears and other body fluids.

Storage

![Lightning bolt symbol] - [Cold temperature] - Below 25 °C
Rifampicin (R)

Update: January 2022

Forms and strengths

- 300 mg capsule and 150 mg tablet

Dosage

- Child under 30 kg: 15 mg/kg (10 to 20 mg/kg) once daily
- Child 30 kg and over and adult: 10 mg/kg (8 to 12 mg/kg) once daily
- Maximum dose: 600 mg daily
- Hepatic impairment: 8 mg/kg once daily max.
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>300 mg capsule</th>
<th>150 mg tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>5</td>
<td>50-100</td>
<td>–</td>
<td>½ tab</td>
</tr>
<tr>
<td>6</td>
<td>60-120</td>
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</tr>
<tr>
<td>29</td>
<td>290-580</td>
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<td>–</td>
</tr>
</tbody>
</table>
Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to rifamycins or history of severe haematological disorders (thrombocytopenia, purpura) during a previous treatment with a rifamycin.
- Avoid or administer with caution to patients with hepatic disorders.
- May cause:
  - hepatotoxicity;
  - influenza-like symptoms, thrombocytopenia, hypersensitivity reactions.

- For the management of adverse effects see Appendix 17.
- Rifampicin reduces the effect of many drugs (antimicrobials, some antiretrovirals, some hormones, antidiabetics, corticosteroids, phenytoin, direct-acting antivirals for chronic hepatitis C, warfarin, etc.):
  - in patients taking antiretrovirals see Appendix 19;
  - in women using contraception, use injectable medroxyprogesterone or an intrauterine device;
  - in the event of concomitant fluconazole administration, administer each drug 12 hours apart (rifampicin in the morning, fluconazole in the evening);
  - for the other drugs, adjust dosage if necessary.

- **Pregnancy and breastfeeding:** no contra-indication. If used in late pregnancy, administer phytomenadione (vitamin K₁) to the mother and the neonate.

Monitoring

- Symptomatic monitoring.
- Liver function in patients with hepatic disease.

Patient instructions

- Take without food (or with a small amount of food to increase gastrointestinal tolerance).
- Harmless orange-red discoloration of the urine, faeces, sweat, saliva, sputum, tears and other body fluids.

Remarks

- For patients on drug-susceptible TB treatment, rifampicin is given as part of a fixed-dose combination.
- For the 6HRZ-Eto regimen for drug-susceptible TB meningitis, the dose of rifampicin is 20 mg/kg once daily (max. 600 mg daily).
- Rifampicin is also used in the treatment of latent TB infection.

Storage

🌞 – 🥵 – Below 25 °C
Rifapentine (P)

Update: October 2022

Forms and strengths

- 300 mg and 150 mg coated tablets

Dosage

- Child 12 years and over and adult 40 kg and over: 1200 mg once daily

<table>
<thead>
<tr>
<th>Age</th>
<th>Daily dose (mg)</th>
<th>300 mg tablet</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 12 years</td>
<td>Do not administer</td>
<td>–</td>
</tr>
<tr>
<td>≥ 12 years</td>
<td>1200</td>
<td>4</td>
</tr>
</tbody>
</table>

Contra-indications, adverse effects, precautions

- Do not administer to patients with hypersensitivity to rifamycins or history of severe haematological disorders (thrombocytopenia, purpura) during a previous treatment with rifamycins.
- Avoid or administer with caution to patients with hepatic disorders.
- May cause:
  - hepatotoxicity;
  - influenza-like symptoms, thrombocytopenia, hypersensitivity reactions.
- For the management of adverse effects see Appendix 17.
- Rifapentine reduces the effect of many drugs (antimicrobials, some antiretrovirals, some hormones, antidiabetics, corticosteroids, phenytoin, direct-acting antivirals for chronic hepatitis C, warfarin, etc.):
  - in patients taking antiretrovirals see Appendix 19.
  - in women using contraception, use injectable medroxyprogesterone or an intrauterine device;
  - in the event of concomitant fluconazole administration, administer each drug 12 hours apart (rifampicin in the morning, fluconazole in the evening);
  - for the other drugs, adjust dosage if necessary.
- **Pregnancy and breastfeeding**: not recommended (safety not established).

Monitoring

- Symptomatic monitoring.
- Liver function in patients with hepatic disease.

Patient instructions

- Take with food.
- Harmless orange-red discoloration of the urine, faeces, sweat, saliva, sputum, tears and other body fluids.

Remarks

- While rifampicin should be taken on an empty stomach, rifapentine is better absorbed if taken with food.
- Also comes in fixed dose combination containing 300 mg of rifapentine/300 mg of isoniazid which can be used in the treatment regimen 2HPZ-Mfx/2HP-Mfx for drug-susceptible TB.
- Rifapentine is also used in the treatment of latent TB infection in children, adolescents, and adults.

Storage
Streptomycin (S)

Update: January 2022

Forms, strengths and route of administration

- Powder for injection, in vial of 1 g streptomycin base, to be dissolved in 4 ml of water for injection, for IM injection
  DO NOT ADMINISTER BY IV INJECTION.

Dosage

- Adolescent 30 kg and over and adult: 12 to 18 mg/kg once daily
- Adult 60 years and over: 15 mg/kg 3 times a week
- Maximum dose: 1000 mg daily
- Renal insufficiency: 12 to 15 mg/kg 2 or 3 times a week

The daily doses take into account the displacement volume (see note below).

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Daily dose (mg)</th>
<th>Daily dose (ml) - IM injection (1 g in 4 ml of water for injection; final volume 4.83 ml; 207 mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>5-29</td>
<td></td>
<td>Not used in patients &lt; 30 kg</td>
</tr>
<tr>
<td>30-33</td>
<td>500</td>
<td>2.4 ml</td>
</tr>
<tr>
<td>34-40</td>
<td>600</td>
<td>2.8 ml</td>
</tr>
<tr>
<td>41-45</td>
<td>700</td>
<td>3.4 ml</td>
</tr>
<tr>
<td>46-50</td>
<td>800</td>
<td>4.0 ml</td>
</tr>
<tr>
<td>51-70</td>
<td>900</td>
<td>4.4 ml</td>
</tr>
<tr>
<td>&gt; 70</td>
<td>1000</td>
<td>Entire volume</td>
</tr>
</tbody>
</table>

Note: displacement volume
Powders for injection are usually formulated such that after reconstitution the final content of the vial corresponds to an adult dose. Errors may occur when only part of the reconstituted solution is to be administered and no allowance is made for the displacement volume. The risk of error increases the greater the weight of the powder and the smaller the volume of solvent used.

Contra-indications, adverse effects, precautions

- Do not administer to children or adolescents under 30 kg and patients with allergy to aminoglycosides.
- Administer with caution to patients 60 years and over or patients with pre-existing renal, vestibular, auditory or severe hepatic impairment.
- May cause:
  - ototoxicity, nephrotoxicity, electrolyte disturbances; rarely, hypersensitivity reactions;
  - local pain after injection.
- For the management of adverse effects see Appendix 17.
- Avoid or monitor combination with other ototoxic and/or nephrotoxic drugs (furosemide, amphotericin B, tenofovir, etc.)
• **Pregnancy**: CONTRA-INDICATED
• **Breastfeeding**: no contra-indication

**Monitoring**

• Symptomatic monitoring.
• Audiometry, serum creatinine and electrolytes (K, Ca, Mg).

**Patient instructions**

• Maintain a good fluid intake to limit renal problems.

**Remarks**

• Use a different site for each injection (absorption may be delayed if the same site is used repeatedly).

**Storage**

°C – Below 25 °C
Patient instructions

Update: January 2022

- Patients on drug-susceptible TB treatment
- Patients on drug-resistant TB treatment
Patients on drug-susceptible TB treatment

TB drugs are usually well tolerated. However, inform patients that they should immediately seek medical attention in the event of:

- Skin rash
- Yellowing of the skin or eyes or dark urine
- Numbness or tingling of fingers or toes
- Decreased urination
- Palpitations
- Blurred vision, reduced visual acuity, blind spot, green-red colour blindness, eye pain, sensitivity to light
- Pain, burning, swelling of a tendon or muscle
- Pain or swelling in the joints
Patients on drug-resistant TB treatment

Inform patients that they should immediately seek medical attention in the event of:

- Skin rash
- Yellowing of the skin or eyes or dark urine
- Numbness or tingling of fingers or toes
- Decreased urination
- Palpitations
- Dizziness or hearing loss
- Blurred vision, reduced visual acuity, blind spot, green-red colour blindness, eye pain, sensitivity to light
- Muscle cramps, spasms, or weakness
- Pain, burning, swelling of a tendon or muscle
- Pain or swelling in the joints
- Personality changes (depression, aggressive behaviour, anxiety)
- Severe abdominal upset or severe nausea, vomiting, black or bloody stools
- Unusual bleeding
Appendix 11. Use of TB drugs in pregnant or breastfeeding women

Update: October 2022
<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Evidence and recommendations</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Fqs</strong></td>
<td>For DR-TB: commonly used in pregnant women despite limited data. Associated with low birth weight in one observational study[^1]. As Fqs reduce mortality from DR-TB, the benefits often outweigh the risks. Avoid breastfeeding if possible[^2] (no absolute contra-indication). For DS-TB: do not use the regimen 2HPZ-Mfx/2HP-Mfx in pregnant or breastfeeding women.</td>
</tr>
<tr>
<td><strong>Bdq</strong></td>
<td>No evidence of fetal harm in animal studies. Associated with low birth weight in one observational study[^1]. As Bdq reduces mortality from DR-TB, the benefits often outweigh the risks. Avoid breastfeeding if possible (no absolute contra-indication).</td>
</tr>
<tr>
<td><strong>Lzd</strong></td>
<td>Few reported cases of use in pregnant women. Fetal harm in animal studies. As Lzd reduces mortality from DR-TB, the benefits often outweigh the risks. Avoid breastfeeding if possible (no absolute contra-indication).</td>
</tr>
<tr>
<td><strong>Cfz</strong></td>
<td>Despite common use for leprosy and MDR-TB in pregnant women, few data on pregnancy outcomes. Fetal harm in animal studies. Use during pregnancy only if the benefits outweigh the risks. Avoid breastfeeding if possible (no data). If used, inform mother of possible (and reversible) skin discolouration of the breastfed infant.</td>
</tr>
<tr>
<td><strong>Cs, Trd</strong></td>
<td>Use during pregnancy only if the benefits outweigh the risks (no data). No contra-indication during breastfeeding.</td>
</tr>
<tr>
<td><strong>Dlm</strong></td>
<td>Use during pregnancy only if benefits outweigh the risks (limited human data, fetal harm in animal studies). Avoid breastfeeding if possible (high concentrations in animal breast milk).</td>
</tr>
<tr>
<td><strong>Ipml/Cln, Mpm</strong></td>
<td>Use during pregnancy and breastfeeding only if the benefits outweigh the risks (no data).</td>
</tr>
<tr>
<td><strong>Am, S</strong></td>
<td>Contra-indicated in pregnancy. No contra-indication during breastfeeding[^2].</td>
</tr>
<tr>
<td><strong>Eto, Pto</strong></td>
<td>For DR-TB: contra-indicated in pregnancy (fetal harm in animal studies[^5]). In breastfeeding women, use only if the benefits outweigh the risks (limited data). For DS-TB: do not use the regimen 6HRZ-Eto in pregnant or breastfeeding women.</td>
</tr>
<tr>
<td><strong>PAS</strong></td>
<td>Use in pregnancy only if the benefits outweigh the risks (limited human data, no fetal harm in animal studies). Avoid breastfeeding if possible (no data).</td>
</tr>
<tr>
<td><strong>Pa</strong></td>
<td>Use during pregnancy and breastfeeding only if the benefits outweigh the risks (no human data, no fetal harm in animal studies[^6]).</td>
</tr>
<tr>
<td><strong>R, Rfb</strong></td>
<td>Not recommended during pregnancy and breastfeeding.</td>
</tr>
</tbody>
</table>

For more specific recommendations for pregnant and breastfeeding women see [Chapter 9](#), [Chapter 10](#), and [Appendix 10](#).

**References**

Appendix 12. Dose adjustments in renal insufficiency

Update: January 2022

12.1 Normal values for creatinine clearance (CrCl)

Women: 88 to 128 ml/minute  
Men: 97 to 137 ml/minute

12.2 Estimation of CrCl (Cockcroft-Gault method)

12.2.1 If serum creatinine is in µmol/litre

\[
\text{CrCl} = \frac{\text{Weight (kg)} \times (140 - \text{age}) \times \text{constant}}{\text{Serum creatinine (µmol/litre)}}
\]

The constant = 1.04 for women and 1.23 for men

12.2.2 If serum creatinine is in mg/dl

\[
\text{CrCl} = \frac{\text{Weight (kg)} \times (140 - \text{age})}{72 \times \text{serum creatinine (mg/dl)}}
\]

For women, the result must be multiplied by 0.85.

Example (calculation with serum creatinine in µmol/litre): 
A woman on cycloserine (Cs), 50 kg, 46 years, serum creatinine = 212 µmol/litre

• Step 1 - Calculate the CrCl:
  50 x (140 – 46) x 1.04 = 4,888
  4,888 ÷ 212 = 23.1
  For this patient, the CrCl is 23.1 ml/minute

• Step 2 - CrCl is < 30 ml/minute, administer 250 mg of Cs once daily or 500 mg 3 times a week.
• Step 3 - Adjust each drug as required according to the table below.

12.2.3 Overweight and obese patients

For overweight (BMI > 25) or obese (BMI > 30) patients, use the ideal body weight (IBW) rather than the actual body weight to avoid overestimation of the CrCl.

The IBW is calculated using the patient’s height:

• IBW women (kg) = 45.4 + 0.89 (height in cm – 152.4)
• IBW men (kg) = 49.9 + 0.89 (height in cm – 152.4)

Example:
A woman, weight 70 kg, height 160 cm (BMI = 27.3, i.e. overweight)

45.4 + 0.89 (160 – 152.4) = 45.4 + 0.89 (7.6) = 45.4 + 6.76 = 52.2

For this patient, the IBW is 52 kg.

12.3 Dosing of TB drugs in renal insufficiency
<table>
<thead>
<tr>
<th>Drugs</th>
<th>Dose and frequency if Clcr &lt; 30 ml/min</th>
</tr>
</thead>
<tbody>
<tr>
<td>H</td>
<td>No change</td>
</tr>
<tr>
<td>R</td>
<td>No change</td>
</tr>
<tr>
<td>Z</td>
<td>25 mg/kg 3 times a week (not daily)</td>
</tr>
<tr>
<td>E</td>
<td>15-25 mg/kg 3 times a week (not daily)</td>
</tr>
<tr>
<td>Rfb</td>
<td>No change</td>
</tr>
<tr>
<td>Mfx</td>
<td>No change</td>
</tr>
<tr>
<td>Lfx</td>
<td>750-1000 mg 3 times a week (not daily)</td>
</tr>
<tr>
<td>Bdq(^{(a)})</td>
<td>No change</td>
</tr>
<tr>
<td>Lzd</td>
<td>No change</td>
</tr>
<tr>
<td>Cfz</td>
<td>No change</td>
</tr>
<tr>
<td>Os(^{(b)})</td>
<td>250 mg once daily or 500 mg 3 times a week</td>
</tr>
<tr>
<td>Dlm(^{(a)})</td>
<td>No change</td>
</tr>
</tbody>
</table>
| Ip/Cln    | 750 mg every 12 hours for CrCl 20-40 ml/min  
500 mg every 12 hours for CrCl < 20 ml/min |
| Mpm       | 750 mg every 12 hours for CrCl 20-40 ml/min  
500 mg every 12 hours for CrCl < 20 ml/min |
| Am\(^{(c)}\) | 12-15 mg/kg 2 or 3 times a week (not daily) |
| S\(^{(c)}\) | 12-15 mg/kg 2 or 3 times a week (not daily) |
| Eto/Pto   | No change                              |
| PAS\(^{(d)}\) | 4 g 2 times daily                      |
| H\(^{h}\)  | No information                        |
| Amx/Clv\(^{(e)}\) | No change                       |
| P         | No change                              |
| Pa        | No information                        |

(a) Use with caution in case of severe renal insufficiency or dialysis (limited data).
(b) Monitor carefully for signs of neurotoxicity.
(c) Use with caution in case of severe renal insufficiency or dialysis (increased risk of nephrotoxicity and ototoxicity).
(d) Avoid sodium salt formulations of PAS in patients with severe renal disease (risk of excessive sodium load).
(e) On a case-by-case basis, consider once daily dosing (e.g. 500/125 mg every 24 hours) for patients with CrCl < 10 ml/minute.
Footnotes

(a) If possible use a calculator to avoid errors, e.g.: 
https://www.mdcalc.com/creatinine-clearance-cockcroft-gault-equation

(b) If possible use a calculator to avoid errors, e.g.: 
https://www.mdcalc.com/ideal-body-weight-adjusted-body-weight
Appendix 13. Daily dose of TB drugs using fixed-dose combinations

Update: October 2022

13.1 Conventional regimens for drug-susceptible tuberculosis

Intensive phase

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Paediatric formulations</th>
<th>Adult formulations</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>HZR 50/150/75 mg</td>
<td>E 100 mg</td>
</tr>
<tr>
<td>4-7</td>
<td>1 tab</td>
<td>1 tab</td>
</tr>
<tr>
<td>8-11</td>
<td>2 tab</td>
<td>2 tab</td>
</tr>
<tr>
<td>12-13</td>
<td>3 tab</td>
<td>2 tab</td>
</tr>
<tr>
<td>14-15</td>
<td>3 tab</td>
<td>3 tab</td>
</tr>
<tr>
<td>16-17</td>
<td>4 tab</td>
<td>3 tab</td>
</tr>
<tr>
<td>18-22</td>
<td>4 tab</td>
<td>–</td>
</tr>
<tr>
<td>23-29</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>30-34</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>35-39</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>40-54</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>55-70</td>
<td>–</td>
<td>–</td>
</tr>
<tr>
<td>&gt; 70</td>
<td>–</td>
<td>–</td>
</tr>
</tbody>
</table>

For example:
- A child weighing 9 kg takes 2 tablets of HZR (50 mg/150 mg/75 mg) + 2 tablets of E (100 mg) once daily.
- A child weighing 20 kg takes 4 tablets of HZR (50 mg/150 mg/75 mg) + 1 tablet of E (400 mg) once daily.

Note: ethambutol is not routinely given to all children; see Chapter 9.

Continuation phase
<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>Paediatric formulation HR 50/75 mg</th>
<th>Adult formulation HR 75/150 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>4-7</td>
<td>1 tab</td>
<td>−</td>
</tr>
<tr>
<td>8-11</td>
<td>2 tab</td>
<td>−</td>
</tr>
<tr>
<td>12-14</td>
<td>3 tab</td>
<td>−</td>
</tr>
<tr>
<td>15-21</td>
<td>−</td>
<td>2 tab</td>
</tr>
<tr>
<td>22-29</td>
<td>−</td>
<td>3 tab</td>
</tr>
<tr>
<td>30-34</td>
<td>−</td>
<td>2 tab</td>
</tr>
<tr>
<td>35-39</td>
<td>−</td>
<td>3 tab</td>
</tr>
<tr>
<td>40-54</td>
<td>−</td>
<td>3 tab</td>
</tr>
<tr>
<td>55-70</td>
<td>−</td>
<td>4 tab</td>
</tr>
<tr>
<td>&gt; 70</td>
<td>−</td>
<td>4 tab</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>Daily dosing in patients &lt; 30 kg</th>
<th>Daily dosing in patients ≥ 30 kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>E</td>
<td>15 to 25 mg/kg</td>
<td>15 to 25 mg/kg</td>
</tr>
<tr>
<td>H</td>
<td>7 to 15 mg/kg</td>
<td>4 to 6 mg/kg</td>
</tr>
<tr>
<td>Z</td>
<td>30 to 40 mg/kg</td>
<td>20 to 30 mg/kg</td>
</tr>
<tr>
<td>R</td>
<td>10 to 20 mg/kg</td>
<td>8 to 12 mg/kg</td>
</tr>
</tbody>
</table>

### 13.2 2HPZ-Mfx/2HP-Mfx regimen for drug-susceptible tuberculosis

<table>
<thead>
<tr>
<th>Weight (kg)</th>
<th>HP 300/300 mg</th>
<th>P 300 mg</th>
<th>Z 400 mg</th>
<th>Mfx 400 mg</th>
</tr>
</thead>
<tbody>
<tr>
<td>40-49</td>
<td>1 tab</td>
<td>3 tab</td>
<td>4 tab</td>
<td>1 tab</td>
</tr>
<tr>
<td>50-64</td>
<td>1 tab</td>
<td>3 tab</td>
<td>4 tab</td>
<td>1 tab</td>
</tr>
<tr>
<td>≥ 65</td>
<td>1 tab</td>
<td>3 tab</td>
<td>5 tab</td>
<td>1 tab</td>
</tr>
<tr>
<td>TB drugs</td>
<td>Daily dosing in patients &lt; 40 kg</td>
<td>Daily dosing in patients ≥ 40 kg</td>
<td></td>
<td></td>
</tr>
<tr>
<td>----------</td>
<td>---------------------------------</td>
<td>---------------------------------</td>
<td></td>
<td></td>
</tr>
<tr>
<td>H</td>
<td>–</td>
<td>300 mg</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Z</td>
<td>–</td>
<td>1600 to 2000 mg</td>
<td></td>
<td></td>
</tr>
<tr>
<td>P</td>
<td>–</td>
<td>1200 mg</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mfx</td>
<td>–</td>
<td>400 mg</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Appendix 14. Monitoring of patients on drug-susceptible TB treatment

Update: October 2022

A cross "X" with no brackets indicates that the exam should be performed in all patients.
A cross between brackets "(X)" indicates that the exam should only be performed in certain patients.

<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>Treatment</th>
<th>End of treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>W2</td>
<td>M1</td>
<td>M2</td>
</tr>
<tr>
<td><strong>Clinical visits</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Vital signs, weight, etc.</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Adverse events</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td><strong>Bacteriological tests</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rapid molecular tests (b)</td>
<td>X</td>
<td>(X)</td>
<td></td>
</tr>
<tr>
<td>Smear microscopy</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Culture and pDST (c)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td><strong>Other investigations</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Radiography (d)</td>
<td>(X)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Full blood count (e)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td>Liver function (f)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td>Serum creatinine (g)</td>
<td>(X)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HbA1c, blood glucose (h)</td>
<td>X</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HIV, HBV, HCV (i)</td>
<td>X</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CD4 and viral load (j)</td>
<td>(X)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

(a) For treatments longer than 6 months.
(b) Rapid molecular tests:
· Xpert MTB/RIF (or Ultra) and Xpert MTB/XDR (or GenoType MTBDRsl if Xpert MTB/XDR not available).
· Repeat RMTs if microscopy or culture is positive at Month 2 or later.

(c) Culture and pDST to first- and second-line drugs:
· At baseline if RMTs are not available, to detect rifampicin andisoniazid resistance or rifampicin resistance mutations not detected by RMTs.
· At Month 2 or later, if RMTs show a new resistance.
· At Month 4, if microscopy is still positive.

(d) Radiography:
· Chest: at baseline for children with presumptive PTB, patients with non-bacteriologically confirmed PTB, suspicion of other intra-thoracic TB, then if indicated (e.g. worsening respiratory symptoms, non-response to TB treatment).
· Bone: at baseline then every 6 months for patients with bone and joint TB.

(e) For patients on AZT or rifabutin.

(f) For patients with pre-existing hepatic disease: AST and ALT (and bilirubin if AST or ALT are elevated).

(g) For patients with renal insufficiency.

(h) For all patients to detect diabetes. If diabetes is detected, monitor according to standard protocols.

(i) For all patients, unless documented HIV, hepatitis B and C status; HIV test every 6 months in high HIV prevalence areas.

(j) For HIV-infected patients.
Appendix 15. Monitoring of patients on drug-resistant TB treatment

Update: June 2023

A cross "X" with no brackets indicates that the exam should be performed in all patients.
A cross between brackets "(X)" indicates that the exam should only be performed in certain patients.
<table>
<thead>
<tr>
<th></th>
<th>Baseline</th>
<th>Treatment</th>
<th></th>
<th>End of treatment</th>
<th>Post treatment&lt;sup&gt;a&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>W1</td>
<td>W2</td>
<td>W3</td>
<td>W4</td>
<td>W5</td>
</tr>
<tr>
<td><strong>Clinical visits</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Vital signs, weight, etc.</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Adverse events</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>BPNS&lt;sup&gt;b&lt;/sup&gt;</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td>Visual function tests&lt;sup&gt;c&lt;/sup&gt;</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td>Audiology&lt;sup&gt;d&lt;/sup&gt;</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td>ECG&lt;sup&gt;e&lt;/sup&gt;</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td><strong>Bacteriological tests</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Smear microscopy</td>
<td>X</td>
<td></td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Culture</td>
<td>X</td>
<td></td>
<td>X</td>
<td>X</td>
<td>X</td>
</tr>
<tr>
<td>Rapid molecular tests&lt;sup&gt;f&lt;/sup&gt;</td>
<td>X</td>
<td></td>
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<tr>
<td>Full pDST&lt;sup&gt;g&lt;/sup&gt;</td>
<td>X</td>
<td></td>
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<tr>
<td><strong>Other investigations</strong></td>
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<tr>
<td>Radiography&lt;sup&gt;h&lt;/sup&gt;</td>
<td>X</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Full blood count&lt;sup&gt;i&lt;/sup&gt;</td>
<td>X</td>
<td></td>
<td>(X)</td>
<td>(X)</td>
<td>(X)</td>
</tr>
<tr>
<td>Liver function&lt;sup&gt;j&lt;/sup&gt;</td>
<td>X</td>
<td></td>
<td>(X)</td>
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<tr>
<td>Serum creatinine and potassium&lt;sup&gt;k&lt;/sup&gt;</td>
<td>X</td>
<td></td>
<td>(X)</td>
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<td>(X)</td>
</tr>
<tr>
<td>HbA1c, blood glucose&lt;sup&gt;l&lt;/sup&gt;</td>
<td>X</td>
<td></td>
<td></td>
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<tr>
<td>Test</td>
<td>Frequency</td>
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<td>-------------------------------------</td>
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<td></td>
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<tr>
<td>HIV, HBV, HCV^{(m)}</td>
<td>X, If indicated</td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>CD4 and viral load^{(n)}</td>
<td>(X), (Every 6 months)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>TSH^{(o)}</td>
<td>(X), (Every 3 months)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pregnancy test^{(p)}</td>
<td>X, If indicated</td>
<td></td>
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<td></td>
</tr>
</tbody>
</table>

(a) Only for patients on BPaLM or BPaL regimen. For other regimens, only for operational research purposes.
(b) For patients on Lzd.
(c) For patients on E, Lzd or thionamides: visual acuity and colour vision deficiency.
(d) For patients on Am or S.
(e) Electrocardiogram, for patients taking:
- < 2 moderate or severe QT-prolonging TB drugs or < 3 QT-prolonging drugs (TB and non-TB): at baseline then monthly.
- ≥ 2 moderate or severe QT-prolonging TB drugs or ≥ 3 QT-prolonging drugs (TB and non-TB) or with other risk factors for QT prolongation or TdP: once a week for the first month, then once a month.
(f) Rapid molecular tests:
- Xpert MTB/RIF (or Ultra) and Xpert MTB/XDR (or GenoType MTBDRs if Xpert MTB/XDR not available).
- Repeat Xpert MTB/XDR (or GenoType MTBDRs) if culture or microscopy is positive at Month 4 or later.
(g) For first- and second-line drugs. Repeat if culture is positive at Month 4 or later.
(h) For all patients at baseline, then every 6 months.
(i) For all patients at baseline, then:
- Patients on Lzd: every 2 weeks for the first 2 months, then once a month.
- Patients on AZT: once a month for the first 2 months, then if indicated.
(j) For all patients: AST and ALT (and bilirubin if AST or ALT are elevated).
(k) For all patients at baseline. Repeat if indicated. For patients on Am or S: once a month or more frequently if indicated.
(l) For all patients to detect diabetes. If diabetes is detected, monitor according to standard protocols.
(m) For all patients, unless documented HIV, hepatitis B and C status; HIV test every 6 months in high HIV prevalence areas.
(n) For HIV-infected patients.
(o) For patients on thionamides or PAS.
(p) For adolescents and women of childbearing age. Repeat if indicated.
Appendix 16. Additional investigations in drug-resistant TB

Update: October 2022

16.1 Electrocardiogram (ECG)

The QT interval is measured in milliseconds (ms) from the start of the QRS complex to the end of the T wave of the ECG. Its value varies depending on the heart rate and should be corrected accordingly (QTc).

To calculate the QTc interval it is recommended to use the Fridericia formula (QTcF):  

\[ QTcF = \frac{QT \text{ interval}}{\sqrt[3]{RR}} \]

Normal QTc values:
- < 470 ms in women
- < 450 ms in men

16.2 Brief peripheral neuropathy screen (BPNS)

Adapted from AIDS Clinical Trial Group (ACTG)[1][2].

Step 1. Grade subjective symptoms

- Ask the patient to rate the severity of symptoms on a scale from 0 (no symptoms) to 10 (most severe symptoms) for right (R) and left (L) feet and legs.
- Enter the score for each symptom in the corresponding column.

<table>
<thead>
<tr>
<th>Symptoms</th>
<th>R</th>
<th>L</th>
</tr>
</thead>
<tbody>
<tr>
<td>a. Pain or burning sensation</td>
<td></td>
<td></td>
</tr>
<tr>
<td>b. Pins and needles sensation</td>
<td></td>
<td></td>
</tr>
<tr>
<td>c. Numbness (lack of feeling)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Symptoms may be unilateral or bilateral and of different intensity. Use the highest subjective sensory neuropathy score to obtain the severity grade.

<table>
<thead>
<tr>
<th>Subjective sensory neuropathy score</th>
<th>Severity grade</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>1-3</td>
<td>1</td>
</tr>
<tr>
<td>4-6</td>
<td>2</td>
</tr>
<tr>
<td>7-10</td>
<td>3</td>
</tr>
</tbody>
</table>

**Step 2. Evaluate vibration perception**

- Place the vibrating 128 Hz tuning fork on the top of the distal joint of the right and left big toes and begin counting the seconds.
- Ask the patient to say when they no longer feel the vibration.

There is a decrease in vibration perception if the patient feels the vibration for 10 seconds or less on both sides.

<table>
<thead>
<tr>
<th>Vibration perception</th>
<th>Result</th>
<th>Grade</th>
</tr>
</thead>
<tbody>
<tr>
<td>Felt &gt; 10 seconds</td>
<td>Normal</td>
<td>0</td>
</tr>
<tr>
<td>Felt 6-10 seconds</td>
<td>Mild loss</td>
<td>1</td>
</tr>
<tr>
<td>Felt &lt; 5 seconds</td>
<td>Moderate loss</td>
<td>2</td>
</tr>
<tr>
<td>Not felt</td>
<td>Severe loss</td>
<td>3</td>
</tr>
</tbody>
</table>

**Step 3. Evaluate tendon reflexes**

Using a reflex hammer, tap the Achilles tendon on each ankle.

**Step 4. Make a diagnosis**

Diagnosis of peripheral neuropathy is based on the combination of:

- subjective symptoms of grade 1, 2 or 3, and
- at least one bilateral objective finding:
  - reduced vibration perception (grade 1, 2 or 3), or
  - decreased reflexes (absent or hypoactive reflexes)

**16.3 Ishihara test**

The patient is asked to look at a set of plates with circles made of dots of different sizes and colours.

Some circles contain dots that form a number or a shape clearly visible to patients with normal colour vision. Patients who cannot see or have difficulty distinguishing numbers or shapes have a red-green colour vision defect.

Some circles contain dots that form a number or a shape visible to patients with red-green colour vision defect, but invisible to patients with normal colour vision.

The test should be performed as per the manufacturer's instructions.
Footnotes
(a) When possible, use a calculator to avoid errors, e.g. https://www.mdcalc.com/corrected-qt-interval-qtc

References


Appendix 16. Basic TB infection control risk assessment tool

Control risk assessment tool.pdf
Appendix 17. Management of adverse effects

Update: January 2022

- **Gastrointestinal disorders**
  - Abdominal pain
  - Diarrhoea
  - Epigastric pain
  - Hepatotoxicity
  - Metallic taste
  - Nausea and vomiting

- **Neurotoxicity**
  - Depression
  - Headache
  - Optic neuritis
  - Ototoxicity
  - Peripheral neuropathy
  - Psychosis
  - Seizures

- **Endocrine disorders**
  - Gynecomastia
  - Hypothyroidism

- **Dermatological disorders**
  - Alopecia
  - Fungal infection
  - Photosensitivity
  - Skin reactions

- **Musculoskeletal disorders**
  - Arthralgias
  - Tendinitis/tendon rupture

- **Miscellaneous**
  - Electrolyte disorders
  - Haematologic disorders
  - Lactic acidosis
  - Nephrotoxicity
  - QT prolongation
Gastrointestinal disorders

- Abdominal pain
- Diarrhoea
- Epigastric pain
- Hepatotoxicity
- Metallic taste
- Nausea and vomiting
Abdominal pain

Eto or Pto, PAS, Cfz, Lzd, FQs, H, Z

Abdominal pain is common with MDR/RR-TB treatment. It can be the early sign of severe adverse effects such as hepatitis, pancreatitis, or lactic acidosis.

Deposition of Cfz crystals may cause severe abdominal pain (presentation of acute abdomen). In this case, stop Cfz until symptoms resolve.
Diarrhoea

PAS, FQs, Eto or Pto, Amx/Clv, Ipm/Cln or Mpm

Diarrhoea, along with cramping, can cause significant difficulty and lead to discontinuation of treatment. PAS often causes diarrhoea at treatment initiation. It usually resolves or improves substantially after some weeks.

For diarrhoea with no blood in stools and no fever, loperamide PO (adult: 4 mg followed by 2 mg after each loose stool to a maximum of 10 mg daily) may be used intermittently, especially when the patient needs to attend social functions or return to work, but not on a daily basis.

Encourage the patient to tolerate some degree of diarrhoea. Prevent (encourage fluid intake including oral rehydration solution) or treat dehydration.

In the event of severe diarrhoea, particularly if associated with blood in stools, severe abdominal pain, or fever > 38.5 °C, consider other causes such as acute bacterial enteritis, or pseudo-membranous colitis (C. difficile) due to FQs. Do not use loperamide in bloody diarrhoea or diarrhoea associated with fever.

Monitor serum electrolytes in patients with severe diarrhoea on QT prolonging drugs.
Epigastric pain

PAS, Eto or Pto, FQs, E, Z

Gastritis (epigastric burning or cramp relieved by eating) or dyspepsia (epigastric pain or discomfort following meals, often accompanied by bloating, sensation of fullness and nausea) are frequent with PAS, Eto or Pto.

- For gastritis:
  - omeprazole PO: 20 mg once daily in the morning for 7 to 10 days. In severe or recurrent cases, dose may be increased to 40 mg once daily and the treatment may be prolonged for up to 8 weeks.
  - Histamine H2-antagonists (e.g., ranitidine) may be an alternative.

- For dyspepsia:
  - omeprazole PO: 10 mg once daily in the morning for 4 weeks

Haematemesis (vomiting of blood) and melena (black stools) are symptoms of a bleeding gastric ulcer and require urgent intervention.
Hepatotoxicity

Z, H, R, P, Eto or Pto, PAS, Bdq, Amx/Clv

All TB drugs may cause hepatotoxicity. However, certain drugs are likely more responsible than others for this adverse effect.

The liver function tests (LFTs) used for the diagnosis and monitoring of hepatotoxicity are serum levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST) and bilirubin.

A mild, transient elevation of ALT and AST may be observed during treatment and usually remains asymptomatic. Significant hepatotoxicity is usually symptomatic.

Clinical features resemble that of viral hepatitis. Early symptoms include malaise, fatigue, loss of appetite, muscle and joint pain. Nausea, vomiting and abdominal pain are common in severe toxicity. Jaundice, scleral icterus, dark (tea-coloured) urine and discoloured stool are signs of clinical worsening.

Differential diagnosis includes infections (e.g. viral hepatitis, cytomegalovirus, leptospirosis, yellow fever, rubella), chronic alcohol use and hepatotoxicity due to other drugs (e.g. anti-epileptics, paracetamol, sulfa drugs, erythromycin).

Clinical hepatitis can be fatal and action should be taken immediately.

1) General management

- Patient with symptoms of hepatitis:
  Stop all TB drugs and perform LFTs:
  a) AST or ALT or bilirubin ≥ 3 times upper limit of normal (ULN): wait for resolution of symptoms, perform LFTs weekly and restart TB treatment when LFTs are < 3 times ULN.
  b) AST, ALT and bilirubin < 3 times ULN and mild symptoms (no jaundice): restart TB treatment, closely monitor the patient and perform LFTs weekly. Continue TB treatment as long as LFTs levels remain < 3 ULN and there are no signs of worsening hepatitis.

- Patient without symptoms of hepatitis, but elevated LFTs:
  a) AST or ALT ≥ 5 times ULN or bilirubin ≥ 3 ULN: stop all TB drugs and perform LFTs weekly. Restart TB treatment when LFTs return < 3 times ULN.
  b) AST and ALT < 5 times ULN and bilirubin < 3 ULN: continue TB treatment and perform LFTs weekly.

If LFTs continue to increase after stopping TB treatment, then ongoing progressive drug-induced hepatitis or an unrelated cause of hepatitis should be suspected.

2) Patient on DS-TB treatment

In most cases, the same treatment can be resumed without incident. The objective is to resume the initial regimen or an alternative regimen as rapidly as possible.

If symptoms reappear or LFTs re-increase, try to reintroduce the TB drugs one by one. Start with E and R and reintroduce H three to 7 days later. If E, R and H have been introduced and the LFT abnormalities have not recurred, do not introduce Z as it is most likely the causative agent.

The alternative regimen depends on the drug causing hepatotoxicity:
- Z is involved: 2 (HR)E/7 (HR)
- H is involved: 6RZE-Lfx
- R is involved: treat as MDR/RR-TB

3) Patient on DR-TB treatment

When restarting TB treatment, start with the drugs least hepatotoxic (E, Lfx or Mfx, Cs or Trd, Dlm, Am or S, Ipm/Cln or Mpm), then drugs moderately hepatotoxic (Bdq, Cfz, Amx/Clav), then give the most hepatotoxic (Z, H, R, Eto or Pto, PAS). Add drugs one at a time every 5 to 7 days, and check LFTs.

The causative agent can generally be identified in this manner. It can be discontinued if not essential and replaced with another less hepatotoxic TB drug.
**Note:** hepatotoxicity may occur in patients receiving regimens containing pretomanid (Pa-Mfx-Z and Bdq-Pa-Lzd). However, the responsible drug has not been determined.
Metallic taste

Eto or Pto, FQs

Encourage the patient to tolerate this adverse effect. Normal taste returns when TB treatment is stopped.
Nausea and vomiting

Eto or Pto, PAS, Z, Amx/Clv, Cfz, Lzd, IpM/Cln or Mpm, Bdq

Nausea and vomiting are frequent, especially with Eto or Pto and PAS during the first few weeks of treatment. To avoid nausea and vomiting, these drugs can be initiated at low dose with gradual increase over one to 2 weeks.

- Always look for:
  - Signs of dehydration (thirst, dry mouth, sunken eyes)
  - Serum electrolytes disorders if vomiting
  - Signs of hepatitis
  - Haematemesis and melena
- Dehydration and electrolyte disorders should be corrected as necessary.
- Treat nausea and vomiting aggressively, using a stepwise approach:

First phase - Adjust administration of the responsible drug

- Administer the suspected drug(s) causing nausea at bedtime.
- Patient on Eto or Pto: stop for 3 to 4 days. If signs improve, gradually resume at a lower dose (250 mg, then if tolerated, 500 mg and so on until the full dose is reached).
- Patient on PAS: stop for 3 to 4 days. If signs improve, gradually resume at a lower dose (2 g, then if tolerated, 4 g and so on until the full dose is reached). Take PAS one hour after taking other TB drugs. If PAS is taken once daily, take in 2 divided doses.
- Encourage the patient: nausea and vomiting often improve over the first weeks and may resolve entirely with time.

Second phase - Administer an antiemetic

ondansetron PO 30 minutes before TB drugs:
Child 6 months to < 2 years: 2 mg once daily
Child 2 to < 4 years: 2 mg 2 times daily
Child 4 to < 12 years: 4 mg 2 times daily
Child ≥ 12 years and adult: 4 to 8 mg 2 times daily
Ondansetron is a QT prolonging drug and should be avoided in patients on Cfz, Bdq, Mfx, Dlm, Lfx.

In adults, when ondansetron is not available or is to be avoided:
metoclopramide PO:
Adult < 60 kg: 5 mg 3 times daily
Adult ≥ 60 kg: 10 mg 3 times daily
The interval between each dose should be at least 6 hours (even in the event of vomiting). Do not use metoclopramide if neurological problems develop.

or
promethazine PO 30 minutes before TB drugs:
Adult: 25 mg

Third phase - Reduce the dose or temporarily stop the responsible drug

- Patient on Eto or Pto: if the patient does not tolerate full dose, avoid giving an adult less than 500 mg daily.
- Patient on PAS: if the patient does not tolerate full dose, avoid giving an adult less than 6 to 8 g daily.
- Patient on Cfz: reduce the dose by half.
- In the event of intractable nausea and vomiting despite dose reduction or interruption of the suspected drug, stop all TB drugs for 3 to 4 days, until signs resolve.
Permanent interruption of a drug should only be considered if it is not essential to treatment.

Note: if there is excessive anxiety over the nausea caused by TB drugs, consider adding diazepam PO (adult: 5 mg 30 minutes before TB drugs). This can help to avoid “anticipation nausea”. The treatment must be short as benzodiazepines may cause dependence and tolerance. Do not exceed 10 days of treatment.
Neurotoxicity

- Depression
- Headache
- Optic neuritis
- Ototoxicity
- Peripheral neuropathy
- Psychosis
- Seizures
Depression

Cs or Trd, Eto or Pto

The treatment of MDR/RR-TB may contribute to depression. Depressive symptoms may fluctuate during TB treatment. History of depression may increase the risk of developing depression during treatment, but is not a contra-indication to use of any of the above TB drugs.

Consider lowering the dose or discontinuing a suspected TB drug, provided this does not compromise the effectiveness of TB treatment.

Other interventions include psychological support to patient (and family if needed) and, when necessary antidepressant treatment. Avoid selective serotonin reuptake inhibitors and tricyclic antidepressants with Lzd (risk of serotonin syndrome).

Suicidal ideation is more commonly associated with Cs or Trd. Evidence of suicidal ideation should prompt immediate action:

- Keep the patient in the hospital for surveillance.
- Stop Cs or Trd.
- Lower the dose of Eto or Pto to 500 mg daily until the patient is stable.
- Refer to mental health consultation.
Headache

Cs or Trd, Bdq, Dlm, FQs

Headache is common during the first months of treatment. It can be treated with analgesics. Headache due to Cs or Trd can be prevented by starting at low dose (250 to 500 mg daily), with gradual increase over 1 to 2 weeks.
Optic neuritis

Lzd, E; rarely H, Eto or Pto

This adverse effect is typically due to Lzd and E.

Symptoms include loss of red-green colour distinction, reduced visual acuity and central scotoma. Loss of red-green colour distinction is the first sign. In this case, stop the suspect drug immediately and permanently.

Symptoms are usually reversible after discontinuation of the drug, but optic neuritis due to Lzd may be irreversible.
Ototoxicity

Aminoglycosides; rarely: Cs or Trd, FQs, Eto or Pto, Lzd

Hearing loss, tinnitus and/or vestibular disorders (vertigo, dizziness, imbalance) are signs of ototoxicity. Ototoxicity is most commonly observed in patients receiving large cumulative doses of aminoglycosides. Concomitant use of loop diuretics (furosemide), particularly in patients with renal insufficiency, may exacerbate ototoxicity.

Baseline and follow-up audiometry is required to detect early hearing loss. Hearing loss in high frequencies (> 4000 Hz) is often the first sign of auditory toxicity due to aminoglycosides and can be unnoticed by the patient.

In case of hearing loss, tinnitus or vestibular disorders, discontinue the suspected drug if this does not compromise the effectiveness of TB treatment.

If no alternative is available, reduce the dose of aminoglycoside (3 times weekly rather than daily, e.g. on Monday, Wednesday and Friday). Continuation of aminoglycoside therapy despite hearing loss almost always results in deafness.

Tinnitus and vestibular disorders can rarely be due to the following drugs: Cs or Trd, FQs, Eto or Pto and Lzd. If stopping the aminoglycoside does not improve symptoms, other drugs can be discontinued to see if the symptoms improve, then reintroduced one by one to see if symptoms return.

Drug-induced tinnitus and vestibular disorders can be irreversible.
Peripheral neuropathy

Lzd, Cs or Trd, H, Eto or Pto; rarely E, FQs

Peripheral neuropathy refers to damage to the nerves located outside of the central nervous system. This adverse effect is associated to several TB drugs but is commonly due to Lzd, Cs or Trd and H.

Peripheral neuropathy occurs most commonly in the lower extremities. Signs and symptoms include sensory disturbances (e.g. numbness, tingling, burning, pain, loss of temperature sensation), difficulty walking, weakness and decreased or absent deep tendon reflexes. At times, sensory changes may occur in upper extremities.

Linezolid-induced neuropathy is extremely painful and may be non-reversible.

1) Patient on DS-TB treatment

- To prevent isoniazid-induced peripheral neuropathy:
  Administer **pyridoxine** PO to patients at risk (pregnant and breastfeeding women, neonates and breastfed infants, and patients with HIV infection, alcohol dependency, malnutrition, diabetes, chronic hepatic disease, and renal impairment) along with their TB treatment:
    - Neonate, infant and child < 5 kg: 5 mg once daily
    - Child ≥ 5 kg and adult: 10 mg once daily

- If peripheral neuropathy develops:
  Administer **pyridoxine** PO
    - Child < 12 years: 10 to 20 mg 2 times daily
    - Child ≥ 12 years: 50 mg 2 times daily
    - Adult: 50 mg 3 times daily
  For pain management: ibuprofen or paracetamol.

2) Patient on DR-TB treatment

- To prevent peripheral neuropathy:
  Administer **pyridoxine** PO:
    - Patient on H: all patients at risk, as for DS-TB.
    - Patient on Cs or Trd, Lzd, H⁰ and Eto or Pto:
      - Neonate, infant, child: 1 to 2 mg/kg (usual range in child: 10 to 50 mg) once daily
      - Adult: 100 mg once daily

- If peripheral neuropathy develops:
  - Patient on Lzd: stop Lzd immediately. For mild symptoms not requiring analgesics, Lzd can be restarted at a lower dose once symptoms subside. For moderate or severe symptoms, stop Lzd permanently. Consider additional TB drugs to reinforce the therapeutic regimen.
  - Patient on Cs or Trd or H⁰: stop these drugs. If they are essential to the regimen, they may be re-introduced once symptoms subside.

Other contributing causes should be addressed (e.g., diabetes or malnutrition).
Administer **pyridoxine** PO: 100 mg daily in adults until symptoms resolve.
For pain management: ibuprofen or paracetamol.
Physiotherapy may be of benefit.
If these measures are insufficient, treat as chronic neuropathic pain, but avoid tricyclic antidepressants in patients on Lzd (risk of serotonin syndrome).
Do not use carbamazepine (strong CYP450 inducer) in patients on Bdq or Dlm.
Psychosis

**Cs or Trd, FQs, H, Eto or Pto**

Visual or auditory hallucinations, delusions, paranoia and bizarre behaviour are hallmarks of psychosis. Health personnel should be familiar with these symptoms to allow early detection.

The most likely TB drug involved is Cs or Trd, but psychotic symptoms may occur with FQs, H, Eto or Pto.

History of psychosis is not a contra-indication to the use of the above-mentioned drugs, though psychiatric symptoms are more likely to occur in such circumstances.

Some patients may need antipsychotic treatment throughout the duration of TB treatment.

Psychosis is generally reversible upon discontinuation of TB treatment.

For acute psychosis:

- If patients are at risk of harming themselves or others: urgent hospitalisation.
- Stop Cs or Trd.
- Treat the acute psychosis.

Once psychotic symptoms have resolved, antipsychotic treatment can be tapered most of the time. Cs or Trd can be resumed, generally at lower dose.

Antipsychotic treatment should be continued until the end of Cs or Trd treatment and then can usually be stopped gradually (do not stop it abruptly).

If the patient does not tolerate the reintroduction of Cs or Trd, another TB drug should be considered.

Whenever psychosis occurs in a patient on Cs or Trd, check the serum creatinine. Cs or Trd is 100% renally excreted and a decrease in renal function can result in toxic levels of Cs or Trd. In this case, a temporary suspension of Cs or Trd and re-introduction at an adjusted dose may be needed ([Appendix 12](#)).
Seizures

Cs or Trd, H, Fqs, Eto or Pto, IpM/Cln or Mpm

All the above-mentioned drugs may cause seizures. However, rule out or treat other possible causes (e.g., epilepsy, meningitis, encephalitis, alcohol withdrawal, hypoglycaemia, stroke, cancer, or toxoplasmosis in HIV-infected patients).

In the event of seizures, measure blood glucose level and blood electrolytes. Measure also serum creatinine. With impaired renal function, TB drugs can reach toxic levels, causing seizures. Dosage adjustment may be necessary (Appendix 12).

A history of seizures is not an absolute contra-indication to the use of the above-mentioned drugs. However, do not use Cs or Trd if there is an alternative. In patients with epilepsy, seizures should be controlled with anti-epileptic therapy before starting TB treatment.

The use of TB drugs (especially H and R) in patients on antiepileptics may lead to decreased blood levels of antiepileptics and seizures.

In patients without history of seizures, a first episode of seizures on TB treatment is likely due to the TB drugs. However, none of the above drugs leave permanent damage.

If a patient has a seizure for the first time:

- Stop suspected TB drugs for a short period.
- Start antiepileptic treatment, especially in the event of repeated seizures after stopping suspected drugs. Do not use carbamazepine or phenytoin in patients receiving Bdq or Dlm (strong CYP450 inducers).
- Reintroduce TB drugs that are essential to TB treatment. Usually, they can be resumed at a lower dose, but the effective dose should be reached as soon as possible.

Antiepileptic treatment may be necessary until the end of the TB treatment.
Endocrine disorders

- Gynecomastia
- Hypothyroidism
Eto or Pto may cause breast enlargement in men and women. Galactorrhoea has been reported. Encourage the patient to tolerate this adverse effect. Symptoms resolve when Eto or Pto is stopped.
Hypothyroidism

Eto or Pto, PAS

Symptoms appear slowly, are nonspecific and may include fatigue, muscle weakness, daytime sleepiness, excessive sensitivity to cold, dry skin, coarse hair, constipation, facial puffiness, and depression. Thyroid enlargement and delayed deep tendon reflexes may be seen on examination.

The diagnosis is confirmed by a serum level of thyroid-stimulating hormone (TSH) ≥ 10 mIU/litre.

Eto or Pto and PAS may cause hypothyroidism, even more frequently when used together. If possible the responsible TB drugs should be replaced but may be continued if there is no alternative.

In both cases, replacement hormone therapy is required:

**Levothyroxine** PO

Adult < 60 years: initially 75 to 100 micrograms once daily then, adjust in 25 microgram increments every 4 to 12 weeks according to response. Usual maintenance dose is 100 to 200 micrograms daily.

Adult ≥ 60 years and/or with significant cardiovascular disease: initially 25 micrograms once daily then, adjust in 25 microgram increments every 4 to 12 weeks according to response. Usual maintenance dose is 100 to 125 micrograms daily.

The daily dose should be taken at the same time each day, 30 to 60 minutes before a meal or a caffeine-containing drink (e.g. coffee, tea) or other drugs to improve absorption.

Monitor TSH until it normalizes below 5 mIU/litre.

Thyroid dysfunction resolves upon discontinuation of TB treatment. Hormone replacement may be discontinued several months after TB treatment completion.
Dermatological disorders

- Alopecia
- Fungal infection
- Photosensitivity
- Skin reactions
Alopecia

H, Eto or Pto

Temporary and mild hair loss may (rarely) occur in the first months of treatment. Encourage the patient to tolerate this adverse effect. Symptoms resolve when TB treatment is stopped.
Fungal infection

FQs

Vaginal, penile, skin fold and oral candidiasis may occur in patients taking FQs. Topical antifungals or short-course oral antifungals are usually effective.
Photosensitivity

Cfx, FQs; rarely Z

Advise patient to avoid direct exposure to the sun, wear protecting clothes (e.g. long sleeves) and use sunscreen.
Skin reactions

All TB drugs

Skin reactions such as itch and skin rash may be hypersensitivity reactions due to any TB drug. General signs of hypersensitivity such as fever, dizziness, vomiting and headache may also occur.

Skin reactions usually appear early during treatment, often in the first month, but rarely during the first week. Most skin reactions are mild or moderate. Severe – even lethal – exfoliative dermatitis (Stevens Johnson’s syndrome) may occasionally occur, particularly if administration of the TB drug continues after first signs of hypersensitivity appear.

Minor skin reactions

- Simple itching: symptomatic treatment (e.g. antihistamine) without interrupting or modifying the TB treatment.
- Localised, mild skin rash, with or without itching:
  - Rule out other possible causes unrelated to TB drugs (i.e. scabies, contact dermatitis).
  - If no obvious other cause, stop all TB drugs.
  - Give symptomatic treatment (an antihistamine, no corticosteroids except in emergencies) and wait for disappearance of symptoms.
  - Once the reaction has resolved, try to determine which drug caused the reaction (see re- challenge of TB drugs below).

Major skin reactions

- Stop all TB drugs.
- In the event of anaphylaxis, manage according to standard emergency protocol (epinephrine, etc.).
- For severe generalised rash, a parenteral corticosteroid may be needed.
- Once the reaction has resolved, try to determine which TB drug caused the reaction (see re- challenge of TB drugs below).
- Never re-introduce any drug resulting in Stevens-Johnson syndrome or anaphylaxis.

Rechallenge of TB drugs

Each TB drug can be reinstated as a “challenge” (a test-dose). Introduce one drug at a time, starting with the drugs least likely to have caused the reaction.

Give the drugs in a setting where a health care provider can respond to any severe allergic reaction.

If a test-dose of any drug causes a reaction, discontinue this drug, unless it is deemed essential to the regimen (in this case, desensitisation can be considered).

- First-line TB drugs
  - Start with isoniazid over 3 days then add rifampicin over 3 days, etc.

<table>
<thead>
<tr>
<th>Drug</th>
<th>Likelihood</th>
<th>Trial dose 1</th>
<th>Trial dose 2</th>
<th>Trial dose 3</th>
</tr>
</thead>
<tbody>
<tr>
<td>H</td>
<td>Least likely</td>
<td>50 mg</td>
<td>Full dose</td>
<td>Full dose</td>
</tr>
<tr>
<td>R</td>
<td>Least likely</td>
<td>75 mg</td>
<td>300 mg</td>
<td>Full dose</td>
</tr>
<tr>
<td>Z</td>
<td>Likely</td>
<td>250 mg</td>
<td>1000 mg</td>
<td>Full dose</td>
</tr>
<tr>
<td>E</td>
<td>Likely</td>
<td>100 mg</td>
<td>500 mg</td>
<td>Full dose</td>
</tr>
</tbody>
</table>

**Note:** if the initial reaction to treatment is severe, a weaker trial dose should be used (approximately 1/10th of the dose indicated for trial dose 1).

- Second-line TB drugs
  - Start with the most important drug in a regimen unless there is suspicion that it is the cause of the reaction. Restart each TB drug...
one after the other, starting at about 1/10 of the dose on Day 1, half-dose on Day 2 and full dose on Day 3.
Musculoskeletal disorders

- Arthralgias
- Tendinitis/tendon rupture
Arthralgias

Z, Rfb, H, Bdq, Fqs

Arthralgias generally diminish over time. Serum uric acid levels are frequently elevated, but this is of little clinical relevance. Anti-hyperuricaemic therapy is of no proven benefit in these patients.

Begin therapy with an anti-inflammatory agent, e.g. ibuprofen PO (adult: 400 to 800 mg 3 times daily). Paracetamol PO (adult: 500 to 1000 mg 3 times daily) may also help bring relief when given together with an anti-inflammatory drug.

If symptoms fail to resolve, consider lowering the dose of the suspected agent (most often Z), if this does not compromise the effectiveness of TB treatment.
Tendinitis/tendon rupture

FQs

In the acute phase, the main symptom of tendinitis is pain when moving the affected joint or palpating the tendon. In later phase, continuous pain and tendon thickening or nodularity may be present. The Achilles tendon is involved in most cases, but other joints may be affected (shoulder, hand, etc.). New and intense physical activities are not recommended during a treatment with a FQ. Tendinitis is more common in older patients, patients with renal insufficiency or under corticosteroids.

Tendon rupture is a complication of tendinitis. Signs and symptoms include a snap or pop sound at the time of rupture, bruising, inability to move the joint and a lack of continuity of the tendon on palpation.

Early detection of tendinitis, symptomatic treatment, and discontinuation of FQ can prevent tendon rupture. If the TB treatment is likely to fail without the FQ, try to continue the FQ. Inform the patient that tendon rupture may occur, but that FQ is essential to prevent TB treatment failure.

Symptomatic treatment:

- Rest the joint involved.
- Pain management: application of ice, and ibuprofen PO:
  
  Adult: 400 to 600 mg every 4 to 6 hours when required, maximum dose: 2400 mg daily.
Miscellaneous

- Electrolyte disorders
- Haematologic disorders
- Lactic acidosis
- Nephrotoxicity
- QT prolongation
Electrolyte disorders

Aminoglycosides

Electrolyte disorders can occur with the aminoglycosides and are typically reversible with discontinuation of therapy.

Other potential causes (vomiting and diarrhoea) should be treated if present.

If clinical signs of mild to moderate hypokalaemia develop (i.e. muscle cramps, spasms or weakness) or if serum potassium level is between 2.5-3.4 mmol/litre, potassium replacement is required:

**potassium chloride PO:**
- Child under 45 kg: 2 mmol/kg (2 ml/kg) daily in divided doses
- Child 45 kg and over and adult: 30 mmol (30 ml) 3 times daily

If clinical signs of severe hypokalaemia develop (i.e. marked muscle weakness, cardiac arrhythmias) or if serum potassium level is < 2.5 mmol/litre, hospitalise and urgently administer potassium chloride by slow IV infusion.

For a patient with hypokalaemia:
- Monitor serum potassium levels and QT interval until they return to normal.
- Consider magnesium PO if serum magnesium cannot be measured. Untreated hypomagnesaemia may lead to “resistance” to correction of hypokalaemia. Magnesium should be taken at least 2 hours before or 4 to 6 hours after the FQs.
Haematologic disorders

Lzd, R, P, Rfb, E

Most TB drugs can cause hematological disorders that may involve any blood cells (red cells, white cells, platelets). However, the TB drugs most involved are Lzd and rifamycins.

<table>
<thead>
<tr>
<th>Severity grade in adults (a)</th>
<th>Anaemia</th>
<th>Neutropenia</th>
<th>Thrombocytopenia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mild</td>
<td>10.5 - 9.5 g/dl</td>
<td>1500 - 1000/mm³</td>
<td>100,000 - 75,000/mm³</td>
</tr>
<tr>
<td>Moderate</td>
<td>&lt; 9.5 - 8.0 g/dl</td>
<td>&lt; 1000 - 750/mm³</td>
<td>&lt; 75,000 - 50,000/mm³</td>
</tr>
<tr>
<td>Severe</td>
<td>&lt; 8.0 - 6.5 g/dl</td>
<td>&lt; 750 - 500/mm³</td>
<td>&lt; 50,000 - 20,000/mm³</td>
</tr>
<tr>
<td>Life-threatening</td>
<td>&lt; 6.5 g/dl</td>
<td>&lt; 500/mm³</td>
<td>&lt; 20,000/mm³</td>
</tr>
</tbody>
</table>

(a) Adapted from NIAID Division of Microbiology and Infectious Diseases, severity scale, Nov-2007.

1) Patient on DS-TB treatment

Rifamycins can cause potentially life-threatening thrombocytopenia. This is more common when used intermittently. Clinical features may include minor haemorrhage (e.g. epistaxis) or severe haemorrhage and thrombocytopenic purpura.

Measure platelets when thrombocytopenia is suspected:

- Moderate thrombocytopenia: stop the rifamycin and monitor platelets weekly until > 75,000/mm³.
- Severe thrombocytopenia: stop all TB drugs. Hospitalise. Treat shock or severe haemorrhage.

In any event rifamycins should not be reintroduced.

2) Patient on DR-TB treatment

Lzd may cause anemia, neutropenia and/or thrombocytopenia.
<table>
<thead>
<tr>
<th>Toxicity</th>
<th>Management</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Mild to moderate</strong></td>
<td>• In all cases:</td>
</tr>
<tr>
<td></td>
<td>▪ Monitor carefully.</td>
</tr>
<tr>
<td></td>
<td>▪ Consider reduction of dose of Lzd (e.g. 300 mg once daily or 600 mg 3 times weekly in adults).</td>
</tr>
<tr>
<td></td>
<td>• For moderate anemia: consider adding erythropoietin (EPO).</td>
</tr>
<tr>
<td></td>
<td>• For moderate neutropenia:</td>
</tr>
<tr>
<td></td>
<td>▪ Stop Lzd.</td>
</tr>
<tr>
<td></td>
<td>▪ Restart at reduced dose once toxicity has decreased to ‘mild’.</td>
</tr>
<tr>
<td><strong>Severe</strong></td>
<td>• In all cases:</td>
</tr>
<tr>
<td></td>
<td>▪ Stop Lzd and monitor carefully.</td>
</tr>
<tr>
<td></td>
<td>▪ If Lzd is essential to the regimen, restart at reduced dose once toxicity has decreased to ‘mild’.</td>
</tr>
<tr>
<td></td>
<td>• For severe anemia: consider adding EPO.</td>
</tr>
<tr>
<td><strong>Life-threatening</strong></td>
<td>• Stop Lzd and monitor carefully.</td>
</tr>
<tr>
<td></td>
<td>• Hospitalise.</td>
</tr>
<tr>
<td></td>
<td>• Perform blood transfusion</td>
</tr>
<tr>
<td></td>
<td>• If Lzd is essential to the regimen consider restarting at reduced dose once toxicity has decreased to ‘mild’.</td>
</tr>
</tbody>
</table>
Lactic acidosis

Lzd

Lactic acidosis is a rare but potentially life-threatening increase of lactic acid in the bloodstream, that can be due to mitochondrial toxicity of certain TB drugs, usually Lzd.

Signs and symptoms include nausea and vomiting, abdominal pain, extreme fatigue, muscle cramps and increased respiratory rate.

If lactic acidosis is suspected, measure blood lactate and pH. Blood lactate ≥ 4 mmol/litre and pH < 7.35 confirm the diagnosis. Stop Lzd and hospitalise for adequate management.

Note that lactic acidosis may also be due to ART (NRTIs).
Nephrotoxicity

Aminoglycosides

Nephrotoxicity is diagnosed by a rise in serum creatinine above baseline. In its early form it is usually asymptomatic, which means it is very important to monitor serum creatinine while on aminoglycosides.

Symptomatic cases may present with decreased urine output, evidence of volume overload (edema, anasarca or shortness of breath) or uremic symptoms such as mental status changes (confusion, somnolence).

Comorbidities such as diabetes or chronic renal failure are not a contra-indication to treatment with aminoglycosides, though caution must be exercised in such circumstances.

• If renal failure occurs:
  ▪ Stop the aminoglycoside.
  ▪ Rule out other causes of renal failure (e.g. diabetes, dehydration, other drugs, congestive heart failure, urinary obstruction, urinary tract infection, prostate hypertrophy).
  ▪ Adjust doses of other TB drugs to creatinine clearance (Appendix 12).
  ▪ Monitor serum creatinine and electrolytes every 1 to 2 weeks until stable.

• If renal function stabilises or improves and if the drug is essential, resume the aminoglycoside adjusted to creatinine clearance (Appendix 12).
QT prolongation

Cfz, Mfx\(^h\), Bdq, Mfx, Dlm, Lfx

Some TB drugs may cause QT prolongation and predispose to torsades de pointes, arrhythmias, and sudden death.
ECG should be performed before starting TB treatment then monitored throughout the course of treatment in patients taking these drugs.
Possible other causes include other QT prolonging drugs ([Appendix 19](#)), hypothyroidism and genetic causes such as long QT syndrome.

Mild or moderate QT prolongation (QTcF > 470 in women and > 450 ms in men and ≤ 500 ms) is common. Severe QT prolongation (QTcF > 500 ms or increase > 60 ms from baseline) is relatively rare.

- In all cases:
  - Measure serum electrolytes and correct electrolyte disorders if necessary.
  - Measure thyroid stimulating hormone (TSH) and, if necessary, treat hypothyroidism.
- For mild and moderate QT prolongation: monitor ECG at least weekly.
- For severe QT prolongation: stop QT prolonging drugs, hospitalise, perform continuous ECG monitoring until QT returns to normal. Once the patient is stable (normal QTcF and no electrolyte disorders), critical QT prolonging TB drugs can be reintroduced:
  - Patient on Bdq: consider resuming while suspending all other QT prolonging drugs.
  - Patient on Mfx: use Lfx instead.
  - Patient on Cfz or Dlm: consider stopping if alternatives are available.
  - Patient on QT prolonging non-TB drug: consider stopping it.
Appendix 17. Air change per hour (ACH) measurement recommendations

The ACH in a mechanically ventilated room should remain more or less constant, whereas natural ventilation will vary according to:

- Whether the doors/windows/vents in that room are open or not;
- Wind speed and direction;
- Temperature and humidity differential between inside and outside.

The ACH rate is one tool among others to assess if:

- The efficiency of the system in delivering the outdoor air and in removing the pollutants to each location in the room;
- The overall airflow direction is from clean to dirty zones.

To calculate the ACH in a given room:

- Start by drawing a sketch of the room;
- Measure the dimensions of the room and calculate the volume (in m³);
- Measure the surface (in m²) of all the openings/vents in the room and air direction across the openings/vents;
- Measure the air speed (in meters per second) using an anemometer.

\[
\text{ACH} = \frac{0.65 \times \text{air speed (m/s)} \times \text{opening area (m}^2\text{)} \times 3600}{\text{Room volume (m}^3\text{)}}
\]

Summary of proposed specifications:

<table>
<thead>
<tr>
<th></th>
<th>Surface (m²)</th>
<th>Height (m)</th>
<th>ACH</th>
<th>Opening window surface area (m²)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Single rooms</strong></td>
<td>&gt; 7.5 (2.5 x 3)</td>
<td>&gt; 3</td>
<td>&gt; 12</td>
<td>&gt; 25%</td>
</tr>
<tr>
<td><strong>Wards</strong></td>
<td>4.5 m²/patient</td>
<td>&gt; 3.5</td>
<td>&gt; 12</td>
<td>&gt; 15%</td>
</tr>
<tr>
<td><strong>Waiting rooms</strong></td>
<td>3 m²/patient</td>
<td>&gt; 3.5</td>
<td>&gt; 12</td>
<td>&gt; 15%</td>
</tr>
<tr>
<td>(preferably outside)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Sputum collection areas</strong></td>
<td>&gt; 1.5</td>
<td>&gt; 2.5</td>
<td>&gt; 20</td>
<td>&gt; 50%</td>
</tr>
<tr>
<td>(preferably outside)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Toilets</strong></td>
<td>&gt; 1.2</td>
<td>&gt; 2.5</td>
<td>&gt; 12</td>
<td>&gt; 25%</td>
</tr>
<tr>
<td><strong>Consultation rooms</strong></td>
<td>&gt; 7.5 (2.5 x 3)</td>
<td>&gt; 3</td>
<td>&gt; 12</td>
<td>&gt; 25%</td>
</tr>
<tr>
<td><strong>Central corridors</strong></td>
<td>&gt; 2</td>
<td>&gt; 3</td>
<td>&gt; 12</td>
<td>&gt; 25%</td>
</tr>
<tr>
<td>(avoid in new buildings)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

There are two main techniques to measure the ventilation. The most commonly used is the anemometer that measures the velocity (speed) of air (see manufacturer’s recommendations for various types of anemometers). The technique using the gas analyser is difficult and should only be used by trained staff.
Appendix 18. Compassionate use

18.1 Definitions

The term “compassionate use” refers to the use of potentially life-saving experimental treatments to patients suffering from a disease for which no satisfactory authorised therapy exists and/or who cannot enter a clinical trial. For many patients, these treatments represent their last hope.

Experimental treatment is below referred to as investigational new drug (IND).

18.2 Indications

Both MDR-TB and XDR-TB can be life-threatening diseases for which approved drugs alone may be ineffective. In some cases, experimental TB drugs, used in combination with approved drugs, could potentially be effective or life-saving.

Compassionate use may be considered for patients presenting with a life-threatening condition (e.g. deteriorating clinical condition due to TB and/or severe immune depression) when:

- Available treatments have failed or are very likely to fail (e.g. regimen comprises less than 3 highly likely effective drugs and/or clinical evolution shows that the treatment is not effective).
- No medical or surgical options are appropriate.
- At least one highly likely effective drug is available (based in the DST result and previous use by the patient). The IND should never be used in monotherapy. It should always be used in conjunction with other drug(s) with proven or probable efficacy in order to prevent emergence of resistance to the IND. In that respect, will be taken into consideration on a case by case basis:
  - the number of remaining drug(s) and their bacteraicidal or bacteriostatic activity: at least one bactericidal or 2 bacteriostatic drugs could be considered as a minimum;
  - the reliability of the DST to the remaining drug(s), treatment history prior to the last DST result;
  - the vulnerability to resistance amplification of the IND if known;
  - the use of the IND does not result in the discontinuation of an essential effective drug. Special attention will be paid if the use of the IND imposes the replacement of an anti- TB drug by a less effective one.

Compassionate use might be considered for a single patient or a group of patients presenting similar characteristics.

The use of two INDs would basically follow the same indications and conditions. Possible interactions and overlapping toxicity between the INDs have to be taken into consideration.

18.3 Minimal requirements

Compassionate use should only be considered if conditions for an adequate management of DR-TB patients are in place: optimal treatment regimen; clinical, biological and bacteriological monitoring; adherence support and follow-up. Results of DST by a validated laboratory are critical to decision making.

In addition to the basic components of regular DR-TB case management a specific monitoring might be required for the use of an IND.

It is essential that a reporting system is in place in order to diligently report any adverse events.

18.4 National regulations

In most countries, only drugs for which a marketing authorization has been granted by the national regulatory agency can be used in humans. Some national regulatory agencies have developed mechanisms to facilitate the access to new drugs at different stages of development, but before market approval. In this case, a party can apply for approval of an IND and then seek the proper permission to import the drug to a country. The use of an IND requires permission from the proper national regulatory authorities and/or country ethic boards.
## Appendix 18. Advantages and disadvantages of ventilation techniques

<table>
<thead>
<tr>
<th></th>
<th>Installation/equipment</th>
<th>Climate</th>
<th>Technical considerations</th>
<th>Cost</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Cold</td>
<td>Hot</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Natural ventilation</strong></td>
<td>Windows and doors</td>
<td>no</td>
<td>yes</td>
<td>simple</td>
<td>very simple</td>
</tr>
<tr>
<td></td>
<td>Whirly birds</td>
<td>no</td>
<td>yes</td>
<td>very simple</td>
<td>very simple</td>
</tr>
<tr>
<td></td>
<td>Chimney</td>
<td>no</td>
<td>yes</td>
<td>very simple</td>
<td>simple</td>
</tr>
<tr>
<td><strong>Assisted natural ventilation</strong></td>
<td>Ceiling, wall and desk fans</td>
<td>no</td>
<td>yes</td>
<td>simple</td>
<td>simple</td>
</tr>
<tr>
<td></td>
<td>Extractors/exhaust fans</td>
<td>no</td>
<td>yes</td>
<td>simple</td>
<td>very simple</td>
</tr>
<tr>
<td><strong>Mechanical</strong></td>
<td>Heating ventilation and air conditioning</td>
<td>yes</td>
<td>ideal</td>
<td>difficult</td>
<td>difficult</td>
</tr>
</tbody>
</table>
Appendix 19. Drug interactions and overlapping toxicities

Update: October 2022

19.1 Interactions between cytochrome P450 inducers/inhibitors and bedaquiline

Drugs interfering with the cytochrome P450 (CYP450) enzyme system should be avoided with bedaquiline.

<table>
<thead>
<tr>
<th>Strong CYP450 inducers</th>
<th>Moderate CYP450 inducers</th>
<th>Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rifampicin</td>
<td>Efavirenz</td>
<td>Decrease bedaquiline plasma concentrations</td>
</tr>
<tr>
<td>Phenytoin</td>
<td>Rifapentine</td>
<td></td>
</tr>
<tr>
<td>Carbamazepine</td>
<td>Rifabutin</td>
<td></td>
</tr>
<tr>
<td>Phenobarbital</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Strong CYP450 inhibitors</th>
<th>Moderate CYP450 inhibitors</th>
<th>Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atazanavir</td>
<td>Erythromycin</td>
<td>Increase bedaquiline plasma concentrations</td>
</tr>
<tr>
<td>Itraconazole</td>
<td>Fluconazole</td>
<td></td>
</tr>
<tr>
<td>Clarithromycin</td>
<td>Verapamil</td>
<td></td>
</tr>
<tr>
<td>Lopinavir</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Nelfinavir</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ritonavir</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Drugs metabolized by CYP</th>
<th>Effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>Emtricitabine</td>
<td>Can increase bedaquiline plasma concentrations</td>
</tr>
</tbody>
</table>

This list is not exhaustive. Clinicians should be informed of any cytochrome P450 inducers and inhibitors their patients may be taking.

19.2 Overlapping toxicity of QT-prolonging drugs

**TB drugs** (mean QT interval prolongation)
- Mild QT prolongation: delamanid (8.6 ms)\(^1\), levofloxacin (4.6 ms)\(^2\),
- Moderate QT prolongation: bedaquiline (12.3 ms)\(^2\), moxifloxacin (12.3 ms)\(^3\).
- Strong QT prolongation: clofazimine (28.5 ms)\(^4\), moxifloxacin high dose (23.14 ms)\(^3\).

**Non-TB drugs**\(^5\)
- Antimalarials: artemisinine derivatives (high risk), quinine
- Antipsychotics: haloperidol (high risk), chlorpromazine, fluphenazine, olanzapine, risperidone
- Cardiac drugs: amiodarone (high risk), beta-blockers, digoxin
- Oral azole antifungals: fluconazole, itraconazole
- Macrolides: azithromycin, clarithromycin, erythromycin
- Anti-nausea drugs: ondansetron
- Antiretrovirals: boosted protease inhibitors, efavirenz

This list is not exhaustive. Clinicians should be informed of any QT-prolonging drugs their patients may be taking.

19.3 Interactions between TB and antiretroviral drugs
AZT: zidovudine; ATV: atazanavir; 3TC: lamivudine; RAL: raltegravir; ABC: abacavir; DTG: dolutegravir; FTC: emtricitabine; TDF: tenofovir disoproxil fumarate; LPV/r: lopinavir/ritonavir; EFV: efavirenz; RTV or r: ritonavir.
R: rifampicin; Rfb: rifabutin; P: rifapentine; Bdq: bedaquiline.

For more information, see University of Liverpool HIV Drug Interaction Checker: [https://www.hiv-druginteractions.org/checker](https://www.hiv-druginteractions.org/checker).

### 19.4 Overlapping toxicities of antiretrovirals and TB drugs

Drugs strongly associated with the listed toxicities appear in bold lettering.

<table>
<thead>
<tr>
<th>TB drugs</th>
<th>NRTI (ABC, 3TC, TDF, AZT)</th>
<th>INI (DTG, RAL)</th>
<th>NNRTI (NVP, EFV)</th>
<th>Boosted PI (LPV/r, ATV/r, DRV/r)</th>
</tr>
</thead>
</table>
| **R**[8][7] | All NRTI  
- Can be combined.  
- No dose adjustment.  | DTG  
- Can be combined.  
- Double the dose of DTG(a).  | NVP  
- Do not combine.  
- Replace NVP with DTG or EFV.  
- If not possible, replace R with Rfb.  
- EFV  
- Can be combined.  
- No dose adjustment.  | ATV/r or DRV/r  
- Do not combine.  
- Replace R with Rfb.  |
| **Rfb**[7] | All NRTI  
- Can be combined.  
- No dose adjustment.  | All INI  
- Can be combined.  
- No dose adjustment.  | NVP  
- Can be combined.  
- No dose adjustment.  
- Monitor Rfb toxicity.  
- EFV  
- Do not combine.  | All boosted PI  
- Can be combined.  
- Reduce the dose of Rfb by half  
- Monitor Rfb toxicity.  |
| **P**[6][7] | All NRTI  
- Can be combined.  
- No dose adjustment.  | All INI  
- Can be combined.  
- No dose adjustment.  | NVP  
- Do not combine.  
- Replace NVP with DTG or EFV.  
- EFV  
- Can be combined.  
- No dose adjustment.  | All boosted PI  
- Do not combine.  |
| **Bdq**[7][8] |  
- Can be combined.  
- No dose adjustment.  | All INI  
- Can be combined.  
- No dose adjustment.  | NVP  
- Can be combined.  
- No dose adjustment.  
- EFV  
- Do not combine.  
- Replace EFV with DTG or NVP.  | All boosted PI  
- Do not combine.  
- Replace boosted PI with DTG.  
- If no alternative, closely monitor ECG.  |

(a) DTG: administer 50 mg 2 times daily, rather than the usual dose of 50 mg once daily.
(b) RAL: e.g. administer 12 mg/kg 2 times daily, rather than the usual dose of 6 mg/kg 2 times daily.
(c) LPV/r:  
- Child: increase the dose of RTV to obtain a one-to-one (1:1) LPV/r ratio  
- Adult: double the dose (e.g. 800/200 mg 2 times daily, rather than the usual dose of 400/100 mg 2 times daily)
<table>
<thead>
<tr>
<th>Toxicity</th>
<th>ARVs</th>
<th>TB drugs</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Abdominal pain</td>
<td>All ARVs</td>
<td>Eto or Pto, PAS, Cz, Lzd, FQs, H, Z</td>
<td>Common. Often benign, but can be an early symptom of severe adverse effects (<a href="#">Appendix 17</a>).</td>
</tr>
</tbody>
</table>
| Depression               | EFV, DTG                  | Cs or Trd, FQs, Eto or Pto, H | • EFV: consider replacing EFV in the event of severe depression.  
  • DTG: can cause depression, but less frequently[^9].                                |
| Diarrhoea                | All PI, DTG               | Eto or Pto, PAS, FQs, Amx/Clav, Ipm/Cln | Common. Also consider opportunistic infections as a cause of diarrhoea or *Clostridium difficile* infection (pseudomembranous colitis). |
| Electrolyte disorders    | TDF (rare)                | Am, S                     | See Nephrotoxicity.                                                                                                                                 |
| Haematological disorders | AZT                       | Lzd                       | • Monitor blood count.  
  • Replace AZT in the event of bone marrow suppression.  
  • For Lzd, see [Appendix 17](#).  
  • If the patient takes CMX, also consider CMX as a cause of haematological disorders. |
| Headache                 | AZT, EFV, DTG             | Cs or Trd, Bdq, Dlm       | Rule out bacterial or cryptococcal meningitis, toxoplasmosis, etc. Headache secondary to AZT, EFV, DTG and Cs or Trd are usually transient. |
| Hepatotoxicity           | NVP, EFV, boosted PIs, DTG | Z, H, R, E, PAS, Eto or Pto, Bdq, Amx/Clav | • If severe, stop ART and TB drugs. When treatment is resumed, start the TB drugs first ([Appendix 17](#)).  
  • If the patient takes CMX, also consider CMX as a cause of hepatotoxicity. |
| Nausea and vomiting      | RTV, NVP, and most other ARVs | Eto or Pto, PAS, Z, Amx/Clav, Cz, Lzd, Ipm/Cln, Bdq | Persistent vomiting can be a result of more severe conditions, such as lactic acidosis and/or drug-induced hepatitis. |
| Nephrotoxicity           | TDF                       | Am, S                     | • Avoid TDF in patients on aminoglycosides.  
  • If an aminoglycoside is essential:  
    ▪ For patients already on ART, replace TDF with ABC  
    ▪ For new patients, start with AZT or ABC.  
  • If TDF and aminoglycoside cannot be avoided, monitor serum creatinine, creatinine clearance and electrolytes at least every 2 weeks. |
| Neurotoxicity            | EFV, DTG                  | Cs or Trd, H, Eto or Pto, FQs | • EFV: numerous transient effects on the central nervous system during first 2-3 weeks of treatment. If they do not resolve, consider replacing EFV. There is limited data on the use of EFV with Cs or Trd; concomitant use is accepted practice provided the patient is closely monitored for neurotoxicity. |
### References


Appendix 19. Upper room ultraviolet germicidal irradiation (UVGI) system

The use of UVGI in the upper part of rooms may be effective in killing or inactivating *M. tuberculosis* generated by infected persons.

### 19.1 Mechanism of action

UV lamps are installed into fixtures suspended from a ceiling or mounted on a wall. Fixtures are shielded with louvers or bafflers in order to block radiation below the horizontal plane of the fixtures. UV lights create in the upper portion of the room a germicidal zone where the bacilli are killed (Figure 1). Patients in the lower portion of the room are not exposed to UVGI lights. Good air mixing is needed to transport the air (and thereby the bacilli) to the upper portion of the room. Disinfection is achieved through the rapid dilution of contaminated lower room air with clean irradiated upper room air.

#### Figure 1

*1 feet = 0.3048 m*

*From the WHO, Implementing the WHO Policy on TB Infection Control in Health-Care Facilities, Congregate Settings and Households*

The lamps should irradiate the entire surface of the upper part of the room (Figure 2), in order to disinfect the largest possible volume of air mixed at a low speed between the upper and lower part of the room.

### UVGI Upper-room Irradiation

#### Figure 2
From Guidelines for the Utilization of Ultraviolet Germicidal Irradiation technology in controlling transmission of tuberculosis in health care facilities in South Africa

Several factors influence the efficiency of UVGI systems:

- **Ventilation rate:** In controlled environment, at rates up to 6 air change per hour (ACH), UVGI systems increase the effect of air cleaning to > 12 ACH. But when ventilation rates are increased above 6 ACH, UVGI system effectiveness could be reduced because the time for bacteria irradiation is shorter.

- **Effective mixing within the room:** May be provided by natural convection currents or fans, preferably ceiling ones. Low velocity ceiling fans boosted UVGI system's effectiveness up to 33% when ACH was below 6.

- **Relative humidity:** Studies have reported rapidly decreasing air cleaning effectiveness in UVGI systems when the relative humidity goes above 70%.

- **Installation:** The height of the room should be minimum 2.5 m and UVGI fixtures should be installed at the minimum height of 2.1 m. As a thumb rule, a 30W lamp should be sufficient for 18 m² of surface, but room shape and type of fixture should be taken into consideration when calculating the needs. For instance, wall-mounted lamps would have a smaller germicidal area than ceiling-mounted ones. Lamps should be on whenever there is a risk of TB transmission. For example, in rooms with hospitalized patients, the lamps should be turned on 24 hours a day.

- **Maintenance:** See below.

### 19.2 Maintenance

Dust-covered and/or old UVGI lamps are less effective, hence the need for a careful maintenance, including regular cleaning:

- Lamps and fixture surfaces should be wiped at least monthly (more often if necessary) with a cloth dampened with 70% alcohol. Do not use water and soap or any detergent. The cleaning should be performed when lamps and fixtures are cool.

- Measurement of UVGI level must be done at installation and at least once a year. A UV light meter programmed to detect UV light on a wavelength of 254 nm is needed. Measurements should be performed at eye level in the occupied zone (~1.6 m) and in upper irradiated portion of the room, at a distance of 1.2 m from the fixture in all possible directions (imitating a circle with measurements done while moving in circumference spaced of 1 m). Ideally, all upper room measurements should be around 30 μW/cm² to 50 μW/cm². Persons doing these measurements should wear protective equipment (UV protective glasses, clothing made of tightly woven fabric, soft cotton gloves) and cover exposed skin with opaque creams with solar-protection factors > 15.

- **UV lamps last between 5 000 and 10 000 hours of continue use (7 to 14 months).** Check manufacturer's information. After this period, UV lamps rapidly lose effectiveness and need to be changed.

### 19.3 Disposal

UV lamps contain mercury and quartz and are considered as hazardous waste. Disposal is extremely difficult in many countries; this should be considered before implementing them. If adequate disposal of the lamps by specialized enterprises is not possible in the country, neither their repatriation; UV lamps should be disposed of by encapsulation (sealed in a metal 200 litre drum filled with concrete and then buried away from water sources).

**Safety considerations**

Reflecting surfaces in the irradiation area of UV lamps must be avoided (i.e. oil painted ceilings, etc.).

At certain wavelengths (including UV-C) UV exposure may be harmful. Skin exposure can produce sunburn (erythema). Exposure of the eyes can produce conjunctivitis (feeling of sand in the eyes, tearing) and/or keratitis (intense pain, sensitivity to light). These symptoms typically commence 6 to 12 hours after exposure.

Despite the fact that these are reversible conditions, health care workers should immediately report them to the IC officer. This could mean that UV irradiation is higher than previously thought at lower room level (lamp poorly positioned? Reflecting surface?).

The USA National Institute for Occupational Safety and Health (NIOSH) states that safe exposure limits are set below those found to initiate eye irritation, the body surface most susceptible to UV. Next table shows the permissible exposure times for given effective irradiances at 254 nm wavelength.
Exposures exceeding this limit would require the use of personal protection equipment to protect the skin and eyes.

In order to avoid overexposure of UVGI, education of health care workers should include basic information on UVGI systems and their potential harmful effects if overexposure occurs.

**References**


5. Riley RL, Permutt S. *Room air disinfection by ultraviolet irradiation of upper air. Air mixing and germicidal effectiveness*. Arch Environ Health


Appendix 20. Treatment supporters

Update: January 2022

Treatment supporters need specific training to know and understand their role in order to provide the patient with adequate treatment education and support. They should be compensated for their time and services and reimbursed for expenses incurred.

20.1 Selecting a treatment supporter

The treatment supporter:

- Is someone from the patient’s community;
- Is preferably a community health worker or a person with a background in health (e.g. pharmacist), but can also be a non-health worker (co-worker or neighbour);
- Is chosen by, or is acceptable to, the patient and their family (e.g. supporter and patient of the same sex);
- Is able to observe the patient’s confidentiality;
- Has a stable living situation;
- Has basic literacy skills (can read and write and has basic numeracy skills);
- Is motivated to care for TB patients and committed to supporting them for the full duration of treatment;
- Lives near enough to the patient to be able to make regular visits (daily or weekly) and go to their home immediately in the event of an emergency;
- Is in good physical condition and not immunosuppressed.

It is usually not recommended to have family members as treatment supporters. The family relationship may interfere with the ability to administer TB treatment, especially if the patient is a child.

20.2 Roles and responsibilities

Role and responsibilities of a treatment supporter may include:

- Supervision of all drug intakes and keeping records on TB treatment card.
- Detection of adverse effects and, when necessary, prompt referral of the patient to a health facility.
- Accompanying the patient to medical consultations.
- Collection and transport of sputum specimens for smear and culture.
- Provision of health education to family members, including the risk of transmission and implementation of infection control measures in the home.
- Detection of signs and symptoms of TB in family members.
- Participation in refresher trainings.

Footnotes


(b) The most common cause of immunosuppression is HIV infection, but chronic illnesses such as diabetes also alter the immune system and are a risk factor for TB infection and active TB.
Appendix 21. Patient therapeutic education

Update: March 2023

Therapeutic education should be provided promptly after diagnosis, then at each clinical visit and whenever considered necessary by the patient or the healthcare team, until the end of treatment. Interviews are done either by the prescribing clinician alone, or with the help of a specially trained staff member or counsellor. Patients may bring someone with them if they wish.

21.1 Initial therapeutic education

Two individual interviews should be organised:

- The first interview, before the start of treatment, aims to provide essential information to help the patient understand and manage the disease and treatment.
- The second interview, one week later, aims to verify that the information given previously has been assimilated, and if necessary, complete or clarify it.

21.1.1 First interview

- Plan a 30 to 45 minute session.
- Adapt the information according to the:
  - Stage of disease (latent TB infection, active TB).
  - Site of TB (pulmonary, extrapulmonary).
  - Resistance pattern (drug-susceptible, drug-resistant).
  - Treatment regimen.
  - Comorbidities, especially HIV infection.
- Explain:
  - The disease and how it is transmitted.
  - The treatment:
    - Total duration; duration of phases (intensive/continuation) if relevant.
    - Clinical and bacteriological monitoring.
    - Treatment administration (self-administered or directly observed).
  - The TB drugs:
    - Where, when, and from whom to get them.
    - How to take them: number of tablets (or doses) per day; with or without food, etc.
    - Storage: e.g. not removed from blister pack ahead of time.
    - Main adverse effects and what to do if they occur.
    - Special precautions according to the situation (e.g. concomitant treatment, pregnancy).
- Measures the patient and their household members should take to prevent the spread of TB or, if the patient is hospitalised, the hospital infection prevention and control measures in place (Chapter 14).
- The importance of HIV testing if not already performed.
- Explain the importance of adherence (regular treatment without omission or interruption). Address patient issues. Identify barriers to adherence and possible solutions. Explain what enablers the patient is eligible for and how to access them (Chapter 13).
- Answer any questions.
- Give the date of the second interview.

21.1.2 Second interview

- Plan a 30-minute session.
- Review patient’s knowledge (disease, treatment and other information provided at the first interview).
- Answer any questions.
- Assess adherence (Appendix 22) and address problems if any (Chapter 13).
- Give/remind the date of the next clinical visit.
21.2 Continuing therapeutic education

An individual interview should be organised at each clinical visit. For the schedule see Appendix 14 or Appendix 15. These interviews aim to consolidate the patient's skills and update them if necessary, especially when there is a modification in the treatment regimen, e.g. when moving from intensive to continuation phase; when replacing a treatment regimen with another; when transitioning to outpatient treatment after hospitalisation.

- Explain the changes in treatment if any (composition, duration, adverse effects, precautions, monitoring schedule, etc.).
- Answer any questions.
- Assess adherence (Appendix 22) and address problems if any (Chapter 13).
- Give/remind the date of the next clinical visit.

Additional sessions should be scheduled as needed, e.g. if there are learning difficulties or significant changes in the patient's life.
Appendix 23. Treatment card for patients on first-line anti-TB therapy
Appendix 24. Tuberculosis register for patients on first-line anti-TB therapy

Tuberculosis register for patients on first-line anti-TB therapy.pdf
Appendix 25. Treatment card for patients on second-line anti-TB therapy

Treatment card for patients on second-line anti-TB therapy.pdf
Appendix 26. Tuberculosis register for patients on second-line anti-TB therapy

Tuberculosis register for patients on second-line anti-TB therapy.pdf
Appendix 27. Respirators

Update: January 2022

27.1 Introduction

Respirators are masks designed to protect the wearer from inhaling bacilli.

Staff must wear a respirator when the risk of TB transmission is high (Chapter 14).

Visitors and attendants must wear a respirator when entering a ward or room of infectious TB patients.

Recommended respirators include:

- The CE-certified filtering facepiece EN 149 FFP2, filtering efficiency 94% if challenged with 0.4µm particles;
- or
- The United States Centre for Disease Control and Prevention/National Institute for Occupational Safety and Health (NIOSH) certified N95, filtering efficiency > 95% if challenged with 0.3µm particles.

27.2 Instructions for use

Respirators are for personal use. The same respirator cannot be shared between staff members or between caregivers.

The respirator should be put on before entering the room and removed after exiting the room.

Respirators must be worn covering the nose, mouth and chin and provide a tight seal around the edge. Every time that a respirator is put on, a seal check has to be performed:

- Fully open the respirator and slightly bend the nose wire to form a curve.
- Separate the two elastic straps and position the respirator under the chin.
- Stretch the two straps over the head, place the first strap at neck-height and the second strap across the top of the head.
- Model the nose wire around the bridge of the nose and secure the edges until you achieve a perfect
- Check for leaks by covering the respirator with both hands and forcefully inhaling and exhaling several times. The respirator should collapse when inhaling and expand when exhaling and no air leak between the face and the respirator should be Otherwise, straps should be readjusted and/or the respirator repositioned until is sealed properly.

Different factors may not allow proper sealing of respirators to the face: respirator size and/or model; respirator wearer's facial features, including beard and facial hair; headscarves, etc.

There is limited evidence on the acceptable length of time a respirator can be worn with maintained efficiency. The filter materials remain functional for weeks or months, but with frequent wearing the respirator will become less adjusted.

An extensively used respirator should be discarded after 7 days. However, if for example, it is only used a few hours 2 to 3 times a week, it can be reused for several weeks[1]. During this period, staff can reuse their respirator provided it is not wet or damaged and its straps are not loosened. Each staff member should keep their respirator in the pocket of their personal gown without creasing it. If the filter material is damaged or the mask has loose straps, the respirator should be discarded immediately.

Note: TB bacillus is trapped in the filter of a mask and will not be released with shaking or other physical movements of the mask.

27.3 Storage

Store in a dry, well ventilated place. Respirators should not be crushed during storage.

27.4 Disposal

Respirators are disposed of as “soft waste” and do not need to be disinfected before being discarded.

27.5 Fit testing
Proper fit of a respirator is critical to ensure respiratory protection. Therefore, all staff members who could be exposed to *M. tuberculosis* should before being required to wear a respirator perform a “fit testing” to determine if the respirators being used fit them properly.

At least two models of respirators should be available. If a worker cannot be fitted with one model, the other one should be used.

Testing is performed using a fit testing kit. The kit contains all the supplies and instructions needed to perform the test.

**Fit testing kit**

![Fit testing kit](image)

**References**

   
   [https://openres.ersjournals.com/content/6/1/00317-2019](https://openres.ersjournals.com/content/6/1/00317-2019)
Appendix 28. Surgical masks

Update: January 2022

28.1 Introduction
The purpose of surgical masks is to catch droplet nuclei that patients expel while talking, breathing or coughing. Surgical masks should be worn by contagious or potentially contagious patients (confirmed or presumed cases) when they leave their rooms to go to another department or any other enclosed area, or when they take care of young children.

The terms “surgical”, “medical” or “procedure” are sometimes used interchangeably to qualify masks. Only masks that conform to the norms EN 14683 or ASTM F2100 should be used.

28.2 Instructions for use
Surgical masks are for personal use. The same mask cannot be shared.
- Open the mask.
- Bend the nasal bar (if included).
- Put the chin into the mask.
- Attach the two straps behind the head or over the ears.

Surgical masks must be replaced at least once a day and when they become wet or damaged.

It is not recommended to wear masks for large portions of the day or while sleeping, as they restrict air movement and are not comfortable.

28.3 Storage
Store in a dry, well ventilated place.

28.4 Disposal
Masks are disposed of as “soft waste” and do not need to be disinfected before being discarded.
Appendix 29. BCG vaccine

Update: January 2022

Composition, forms and route of administration

- Live attenuated bacterial vaccine
- Powder for injection, to be dissolved with the entire vial of the specific solvent supplied by the manufacturer, in multidose vial, for intradermal injection

Dosage and vaccination schedule

Refer to national recommendations. In countries with a high incidence of TB (> 40 cases per 100,000), WHO recommends[^1]:
- Child under 12 months: 0.05 ml single dose as soon as possible after birth
- Child 12 months and over[^a] and adult: 0.1 ml single dose

Technique and site of administration

- Clean the injection site with clean water. Do not use antiseptics as risk of inactivation of vaccine). Allow to dry.
- Administer intradermally. If the injection is correctly performed, an "orange-skin" papule measuring 5-8 mm in diameter should appear at the injection site.
- The vaccine is administered in the deltoid region of the arm, about one-third down the upper arm over the insertion of the deltoid muscle.
- The vaccine should be injected in the same place for each child so that the BCG scar is easier to locate.

Contra-indications

- Do not administer to patients with congenital or acquired immunodeficiency (e.g. HIV infection or serologic status unknown, but symptoms consistent with HIV infection, immunosuppressive therapy, malignant haemopathy).
- Postpone vaccination until recovery in the event of acute extensive dermatosis, acute complicated malnutrition or severe acute febrile illness (minor infections are not contra-indications).

Adverse effects

- Local reaction 2-4 weeks after injection: papule that ends up as an ulcer and usually heals spontaneously (dry dressing only) after 2 to 5 months, leaving a permanent
- Complications requiring no specific treatment and which almost always evolve favourably:
  - persistent ulcer with serous discharge for over 4 months after injection;
  - non-suppurated adenitis, most often axillary, sometimes cervical;
  - abscess at the injection site due to infection (red, hot and painful abscess) or inadvertent intradermal injection (cold and painless abscess).
- Uncommon complications:
  - suppurative lymphadenitis, mostly observed in neonates, usually due to inadvertent intradermal injection. The lymph node, which can have a diameter of over 3 cm, evolves toward softening and fistulisation with chronic osteomyelitis in exceptional cases.
  - disseminated BCG disease[^b], most commonly in immunocompromised children under 2 years old (mortality rate > 70%)[^2].

Precautions

- If administered simultaneously with other vaccines, use different syringes and injection sites. Do not mix with other vaccines in the same syringe.
- Pregnancy: CONTRA-INDICATED
- Breastfeeding: no contra-indication

Storage
- Reconstituted vaccine: between 2 °C and 8 °C for 6 hours max.
- Powder: between 2 °C and 8 °C.
- Solvent: a cold chain is not required for However, at least 24 hours before reconstitution of the vaccine, the solvent must be refrigerated between 2 °C and 8 °C so that the solvent and lyophilised powder are at the same temperature: a temperature difference during reconstitution may reduce vaccine efficacy. Do not freeze.

Footnotes
(a) BCG vaccine provides high protection for neonates, but only moderate for school age TST negative children.

(b) If disseminated BCG disease is diagnosed, a 6-month TB treatment should be administered.

References

Appendix 29. Sputum smear microscopy register

Sputum smear microscopy register.pdf
Appendix 30. Xpert MTB/RIF register

Xpert MTB/RIF register.pdf
Appendix 32. Quarterly report

Quarterly report.pdf
Appendix 33. Report on detection and enrolment of TB cases with rifampicin and multidrug-resistance
Appendix 34. Request form for smear microscopy and Xpert assays

Update: March 2023

Request form for smear microscopy and Xpert assays.pdf

Request form for smear microscopy and Xpert assays.docx
Appendix 34. Report of final outcomes of drug-resistant tuberculosis

[Link to Report of final outcomes of drug-resistant tuberculosis.pdf]
Appendix 35. Request form for culture, pDST, LPA, genome sequencing

Update: August 2023

Request form for culture, pDST, LPA, genome sequencing.pdf

Request form for culture, pDST, LPA, genome sequencing.docx
Appendix 35. Check-list for the evaluation of a TB service

Check-list for the evaluation of a TB service.pdf